

## Supporting Contents

# Dereplication of Components Coupled with HPLC-qTOF-MS in the Active Fraction of *Humulus japonicus* and it's Protective Effects Against Parkinson's Disease Mouse Model

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## Table of Contents

### Experimental section

Fig. S1. HR-ESI-MS of compound **1**.

Fig. S2. <sup>1</sup>H NMR spectrum (CD<sub>3</sub>OD, 500 MHz) of compound **1**.

Fig. S3. <sup>13</sup>C NMR spectrum (CD<sub>3</sub>OD, 125 MHz) of compound **1**.

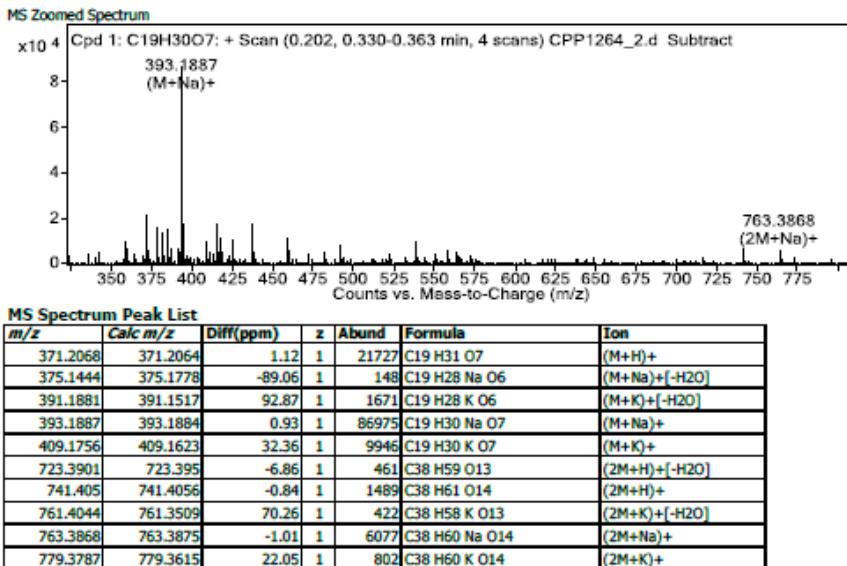
Fig. S4. HSQC spectrum (CD<sub>3</sub>OD, 500 MHz) of compound **1**.

Fig. S5. HMBC spectrum (CD<sub>3</sub>OD, 500 MHz) of compound **1**.

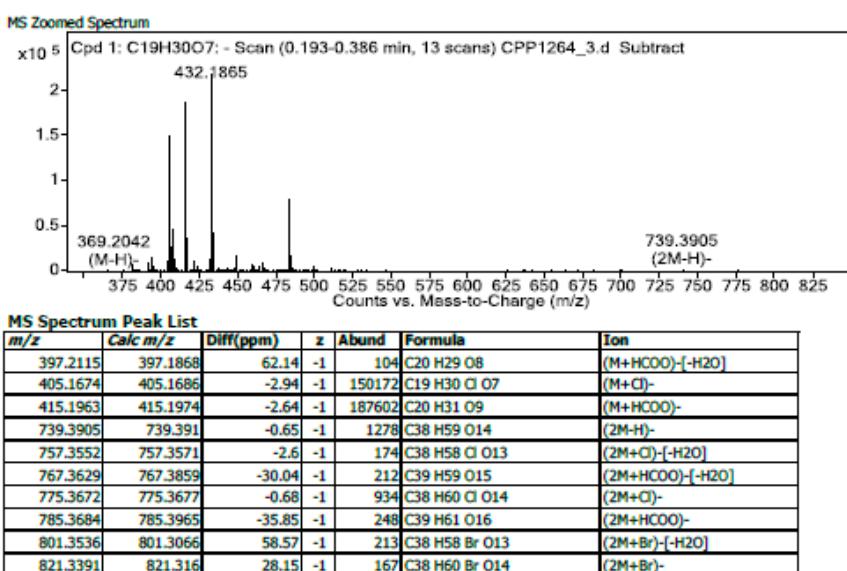
## Experimental section

### *HPLC analysis*

The HPLC system was equipped with an Agilent Series 1260 liquid chromatography, equipped with G1322A vacuum degasser, G1312C binary pump, G1329B autosampler, and G1315D DAD detector, connected to Agilent ChemStation software (Agilent Technologies Co. Ltd., Waldbronn, Germany). An INNO C18 column (4.6 x 250 mm, 5 μm, Young Jin Biochrom, Korea) were used. The gradient profile was as follows: 0–3 min, initial mobile phase 0.1% formic acid in acetonitrile/0.1% formic acid in water (10:90, %, v/v); 3–43 min, linear gradient 90:10 (%), v/v); 43–52 min, isocratic 100:0 (%), v/v); and stable steps to initial condition for 8 min. Flow rate was 0.6 ml/min and detection was at 254 nm. The injection volume of standards (luteolin-7-O-glucoside and apigenin-7-O-glucoside), HJ extracts and fraction was 10 μl, respectively (HPLC chromatogram data not shown).

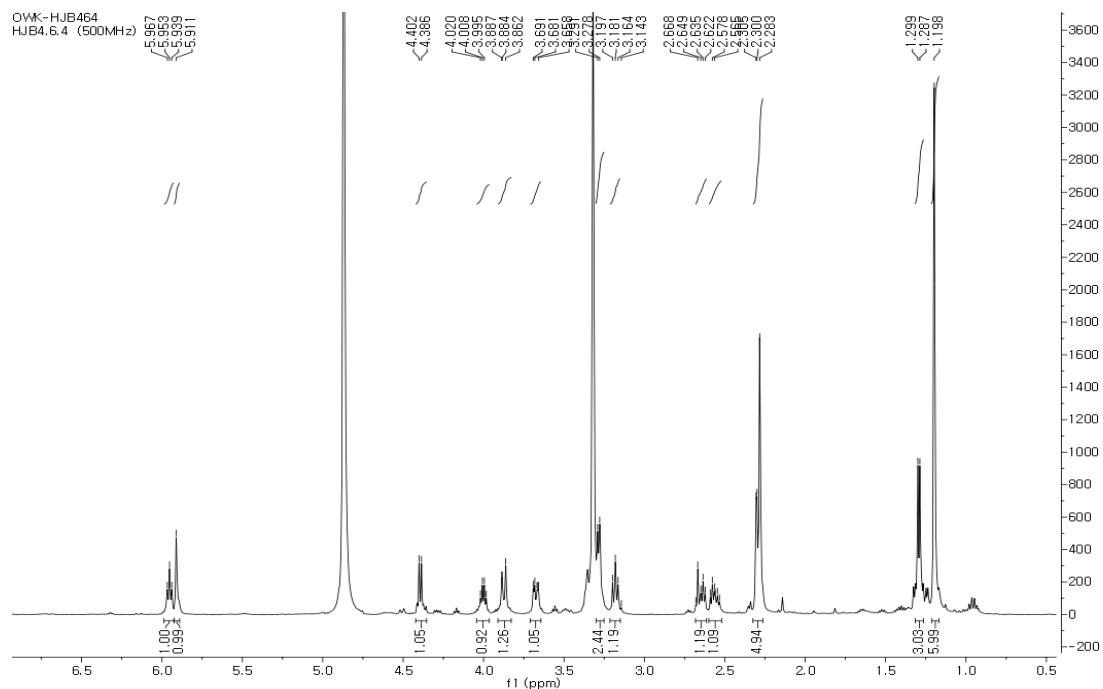


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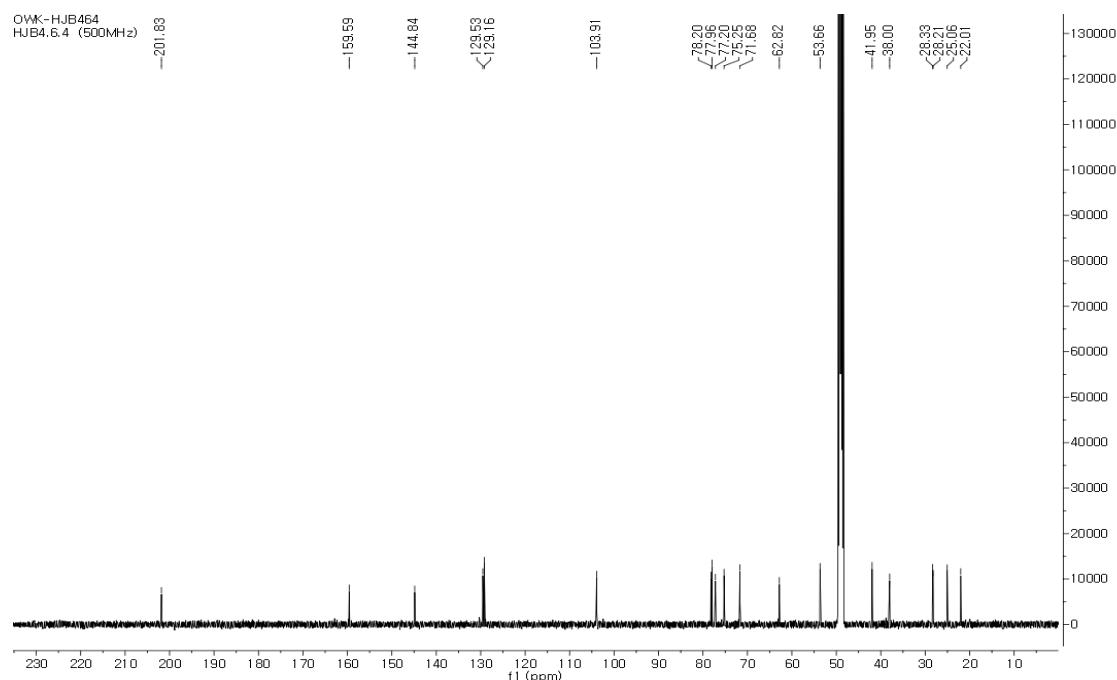


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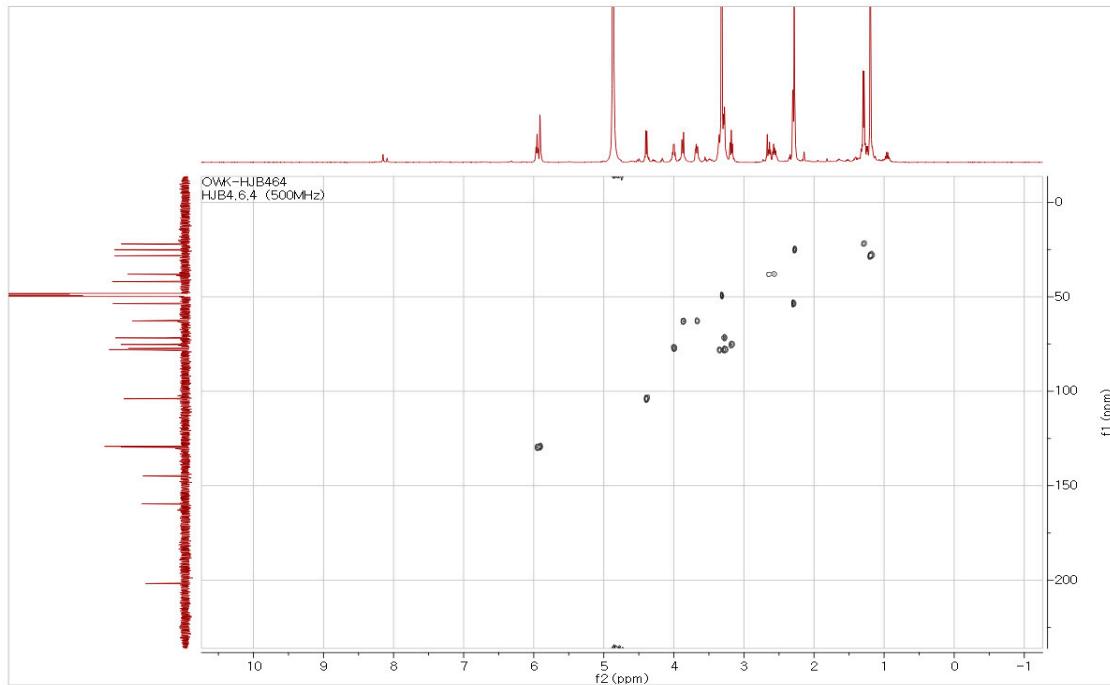
**Fig. S1.** HR-ESI-MS of compound 1.



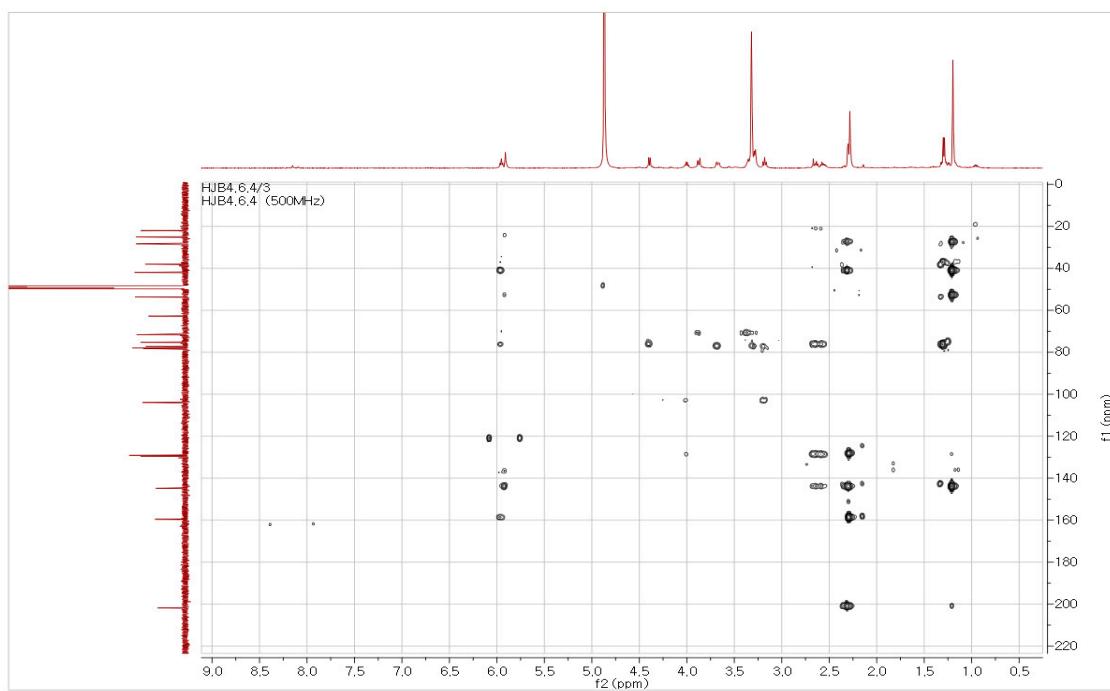
**Fig. S2.**  $^1\text{H}$  NMR spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of compound 1.



**Fig. S3.**  $^{13}\text{C}$  NMR spectrum ( $\text{CD}_3\text{OD}$ , 125 MHz) of compound 1.



**Fig. S4.** HSQC spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of compound **1**.



**Fig. S5.** HMBC spectrum ( $\text{CD}_3\text{OD}$ , 500 MHz) of compound **1**.