

## Supplementary Materials

### Improved Access to Chiral Tetranaphthoazepinium-Based Organocatalysts

#### Using Aqueous Ammonia as Nitrogen Source

Auraya Manaprasertsak, Sorachat Tharamak, Christina Schedl, Alexander Roller and  
Michael Widhalm

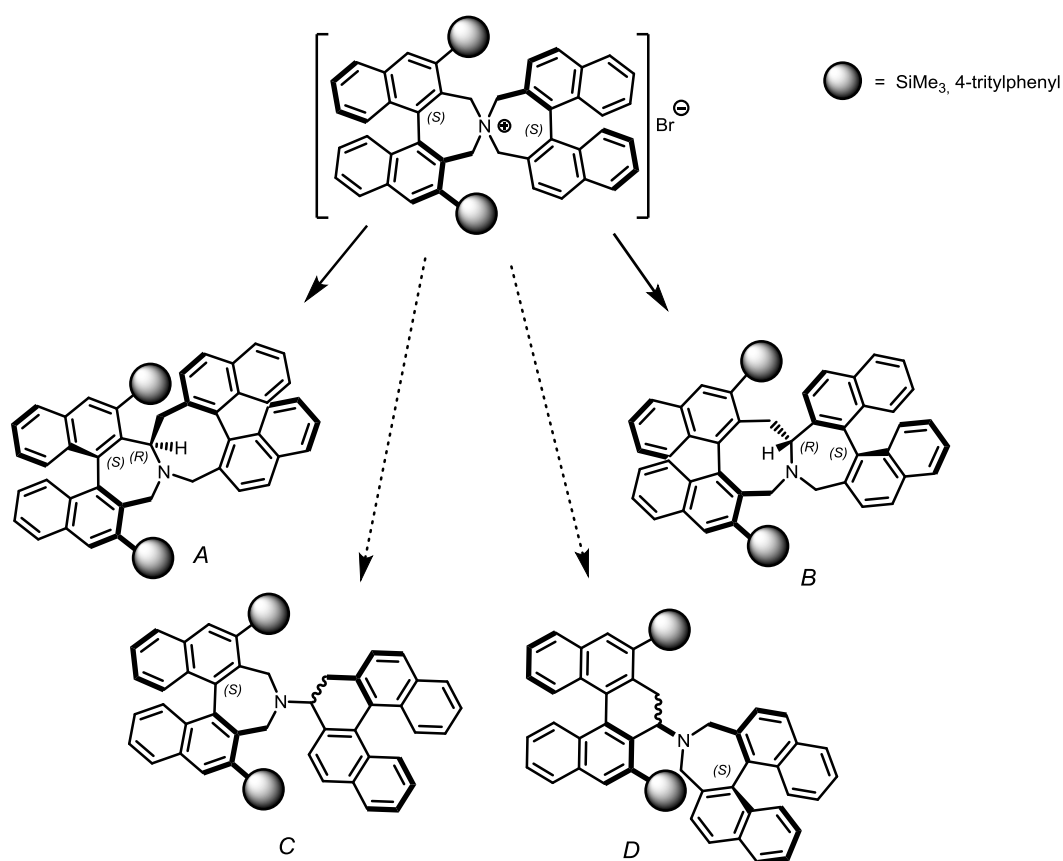
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Proposed sequence products of a Stevens rearrangement of **1e** and **5**, respectively

During attempted spirocyclisation of (*S*)-**9e** with (*S*)-2,2'-bisbromomethyl-1,1'-binaphthyl in MeCN the target compound (*S*)(*S*)-**1e** was isolated in small amounts (<10%, ~80% purity). Instead considerable amounts of dinaphthooxepine along with a sequence product **1e-S** from **1e** resulting from Stevens rearrangement was detected. Feasible structures *A-D* are depicted in Scheme S1. Based on the <sup>1</sup>H NMR spectrum showing seven aliphatic protons in different environment (2x AB, 1x ABX system) structure *C* and *D* can be excluded. From steric reasons an equatorial attack of the CH<sub>2</sub> group will be favored resulting in an axial methine proton in *A* or *B*. Due to the complexity of NMR spectra in the aromatic range only partial assignment of signals was done and no decision between structure *A* and *B* for **1e-S** is possible at the time.

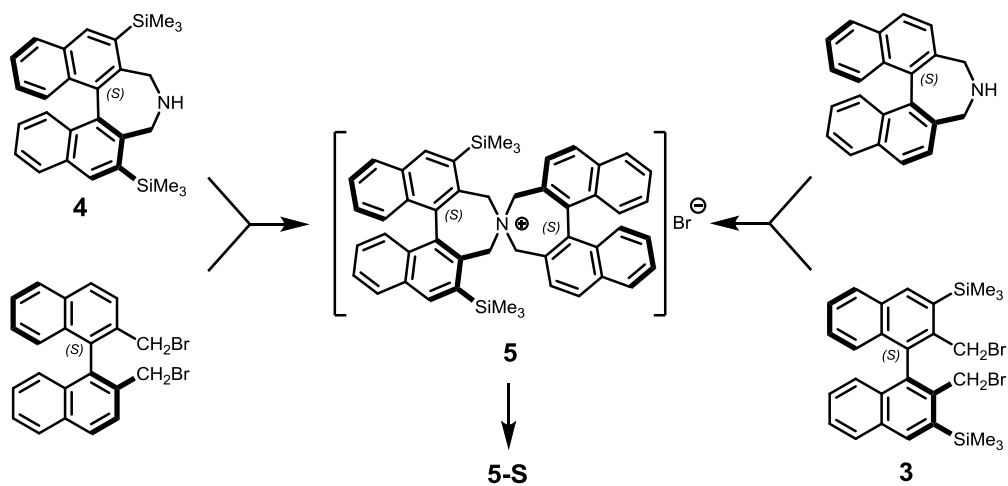
**Scheme S1.** Stevens Rearrangement of Postulated Dinaphthoazepinium Bromides **1e** and **5** with Bulky Substituents.



Also in case of (*S*)-**5** reaction of **4** with (*S*)-2,2'-bisbromomethyl-1,1'-binaphthyl afforded a rearranged product **5-S** with similar <sup>1</sup>H NMR spectrum and correct HRMS. It is worth to note that the same product **5-S** was also obtained from unsubstituted dinaphthoazepine and dibromide **3**, indicative for running

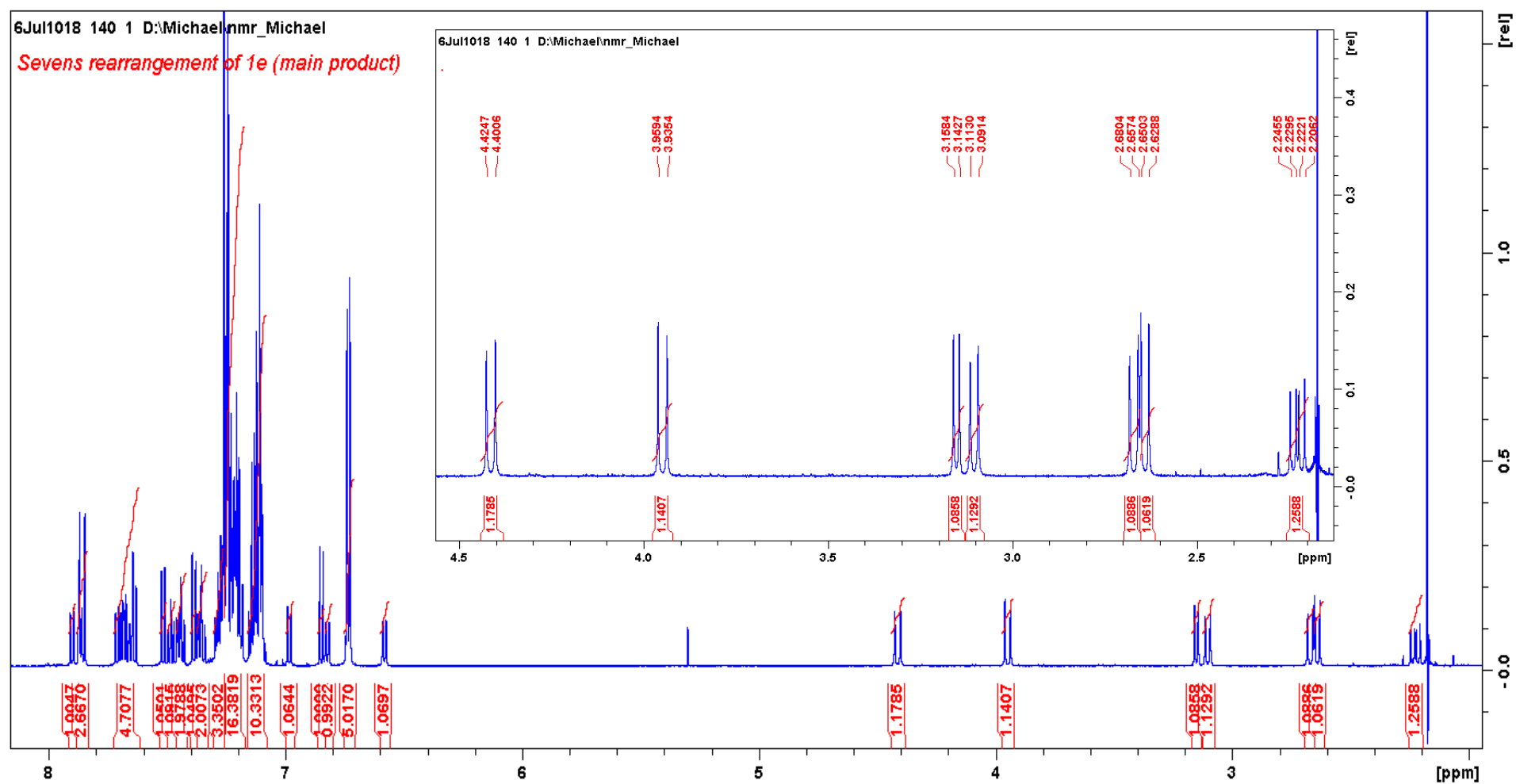
through a common intermediate **5** (Scheme S2). Attempted transformation into corresponding methylammonium salts failed.

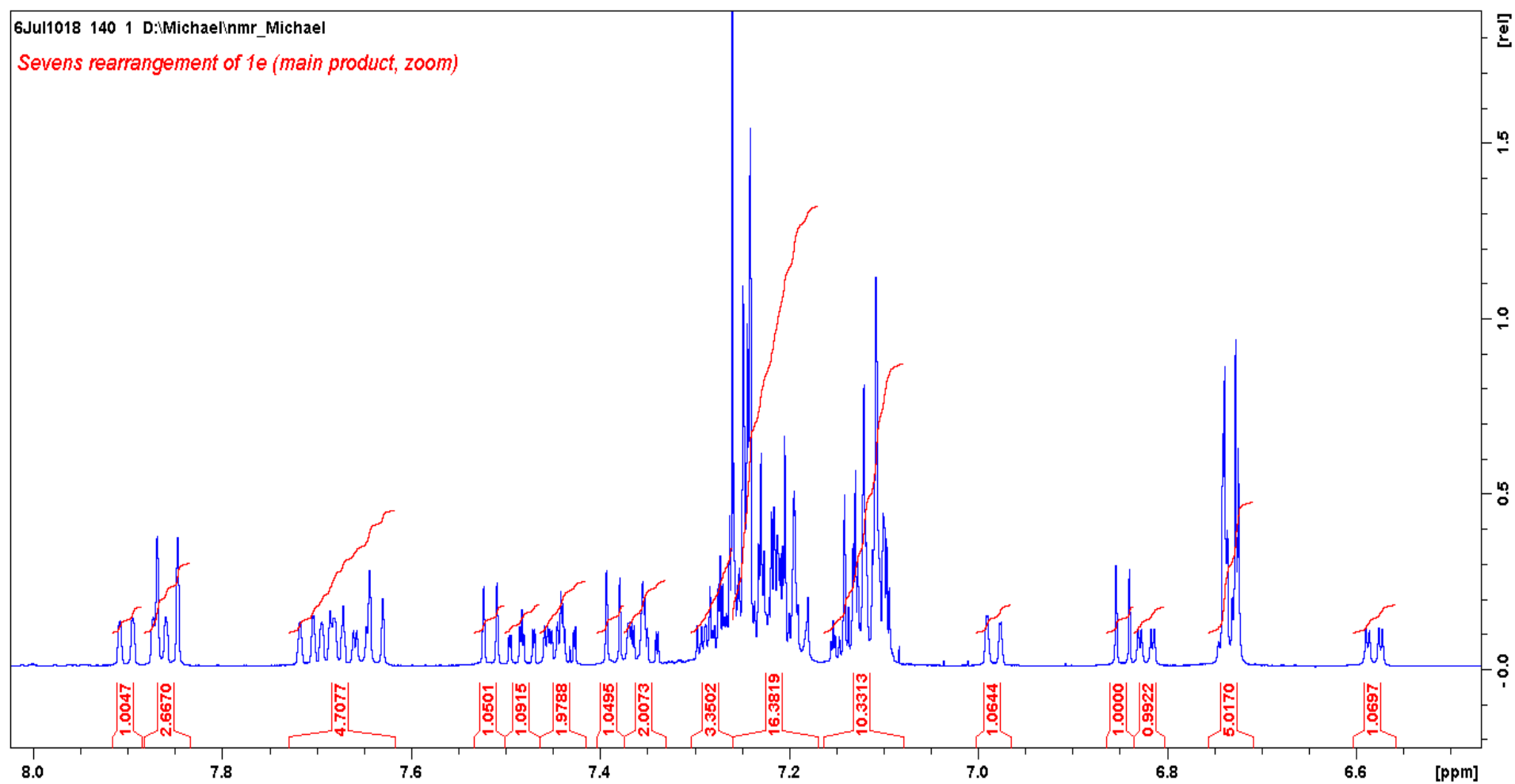
**Scheme S2.** Access of **5-S** from different precursors.



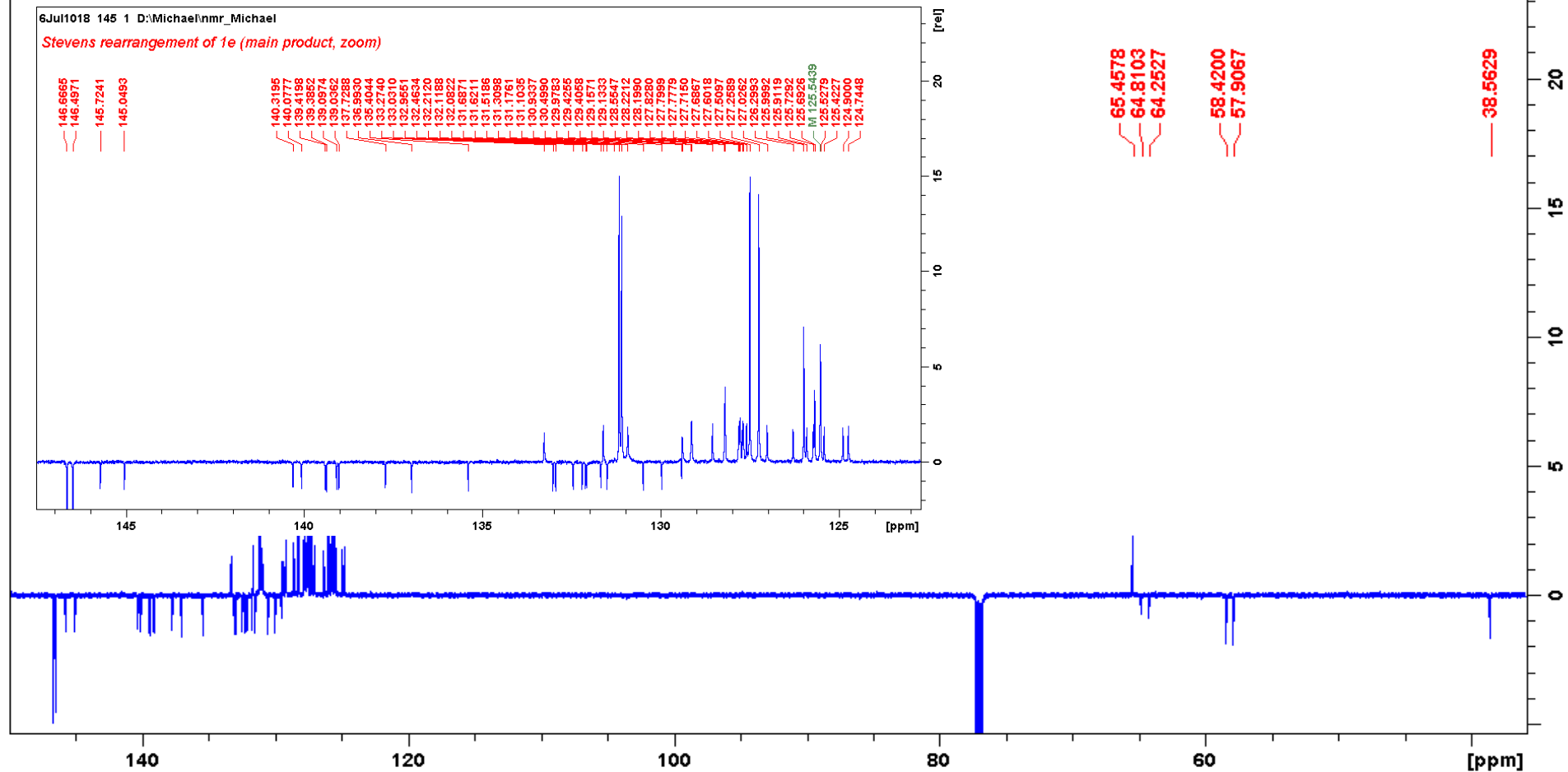
**1e-S**: HRMS: Calcd for  $\text{C}_{94}\text{H}_{68}\text{N}$   $[\text{M} + \text{H}]^+$ : 1210.5347, found: 1210.5337.

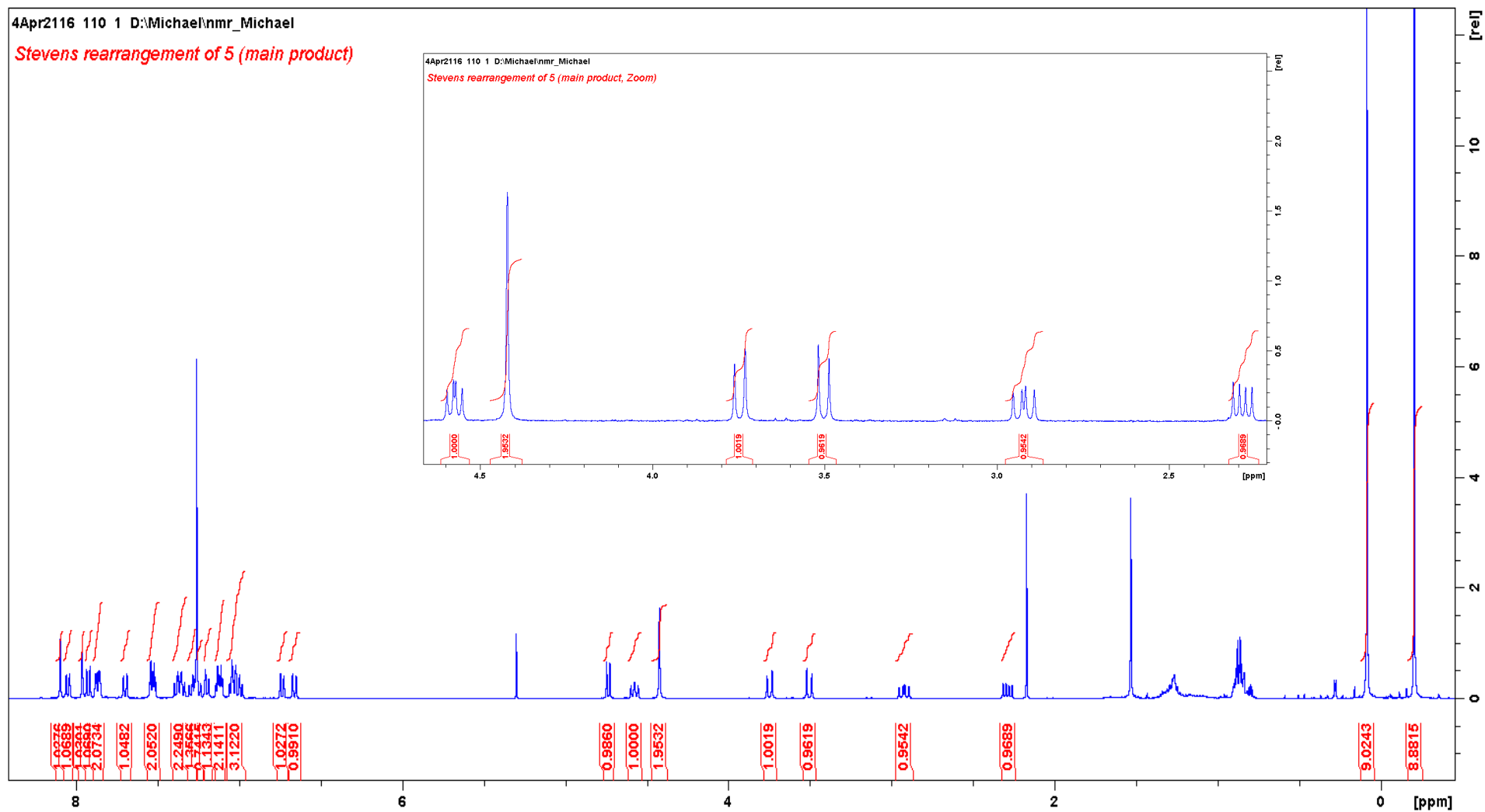
**5-S**: HRMS: Calcd for  $\text{C}_{50}\text{H}_{47}\text{NSi}_2$   $[\text{M} + \text{H}]^+$ : 718.3325, found: 716.3314.

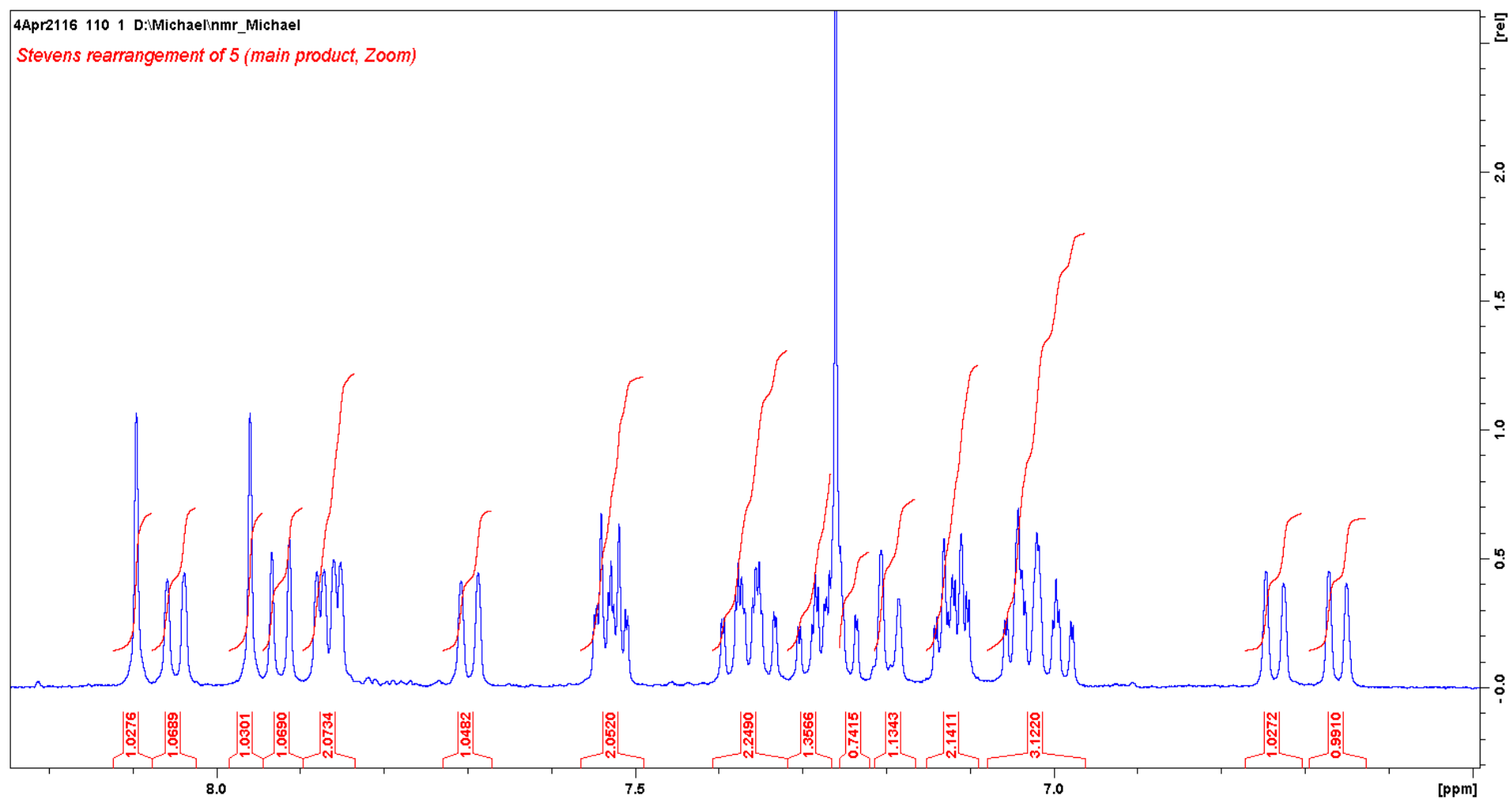




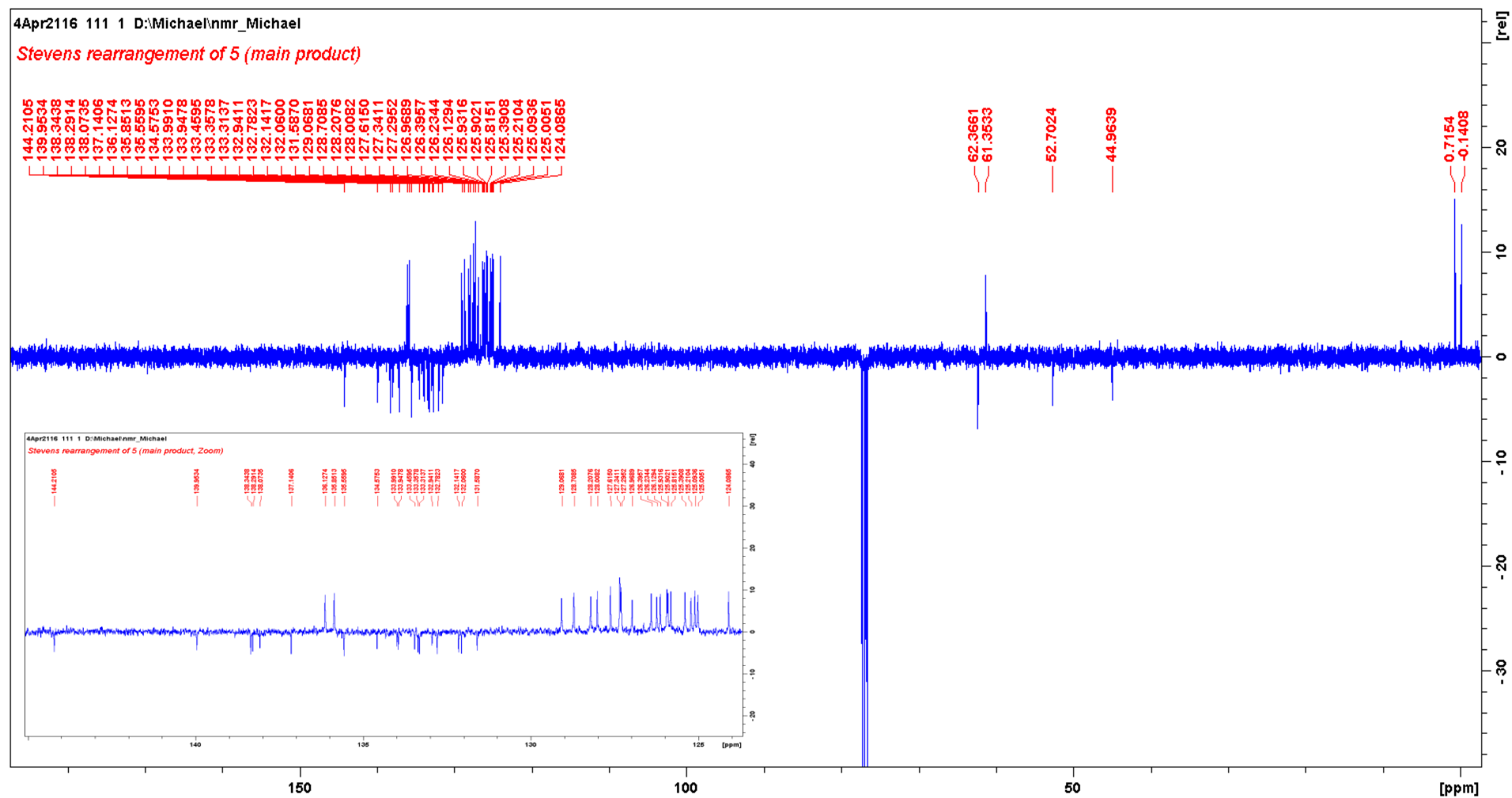
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*Stevens rearrangement of 1e (main product)*



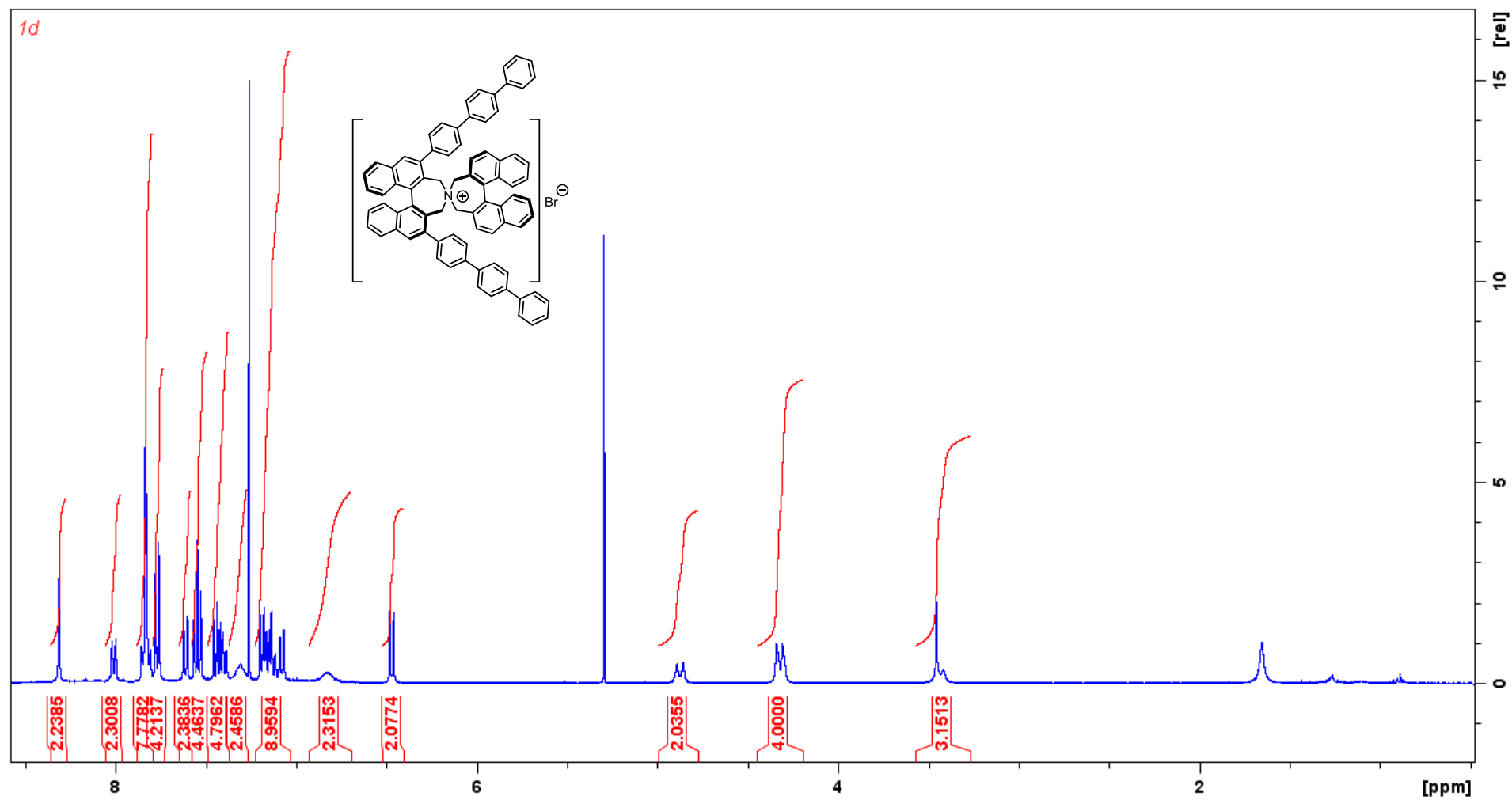


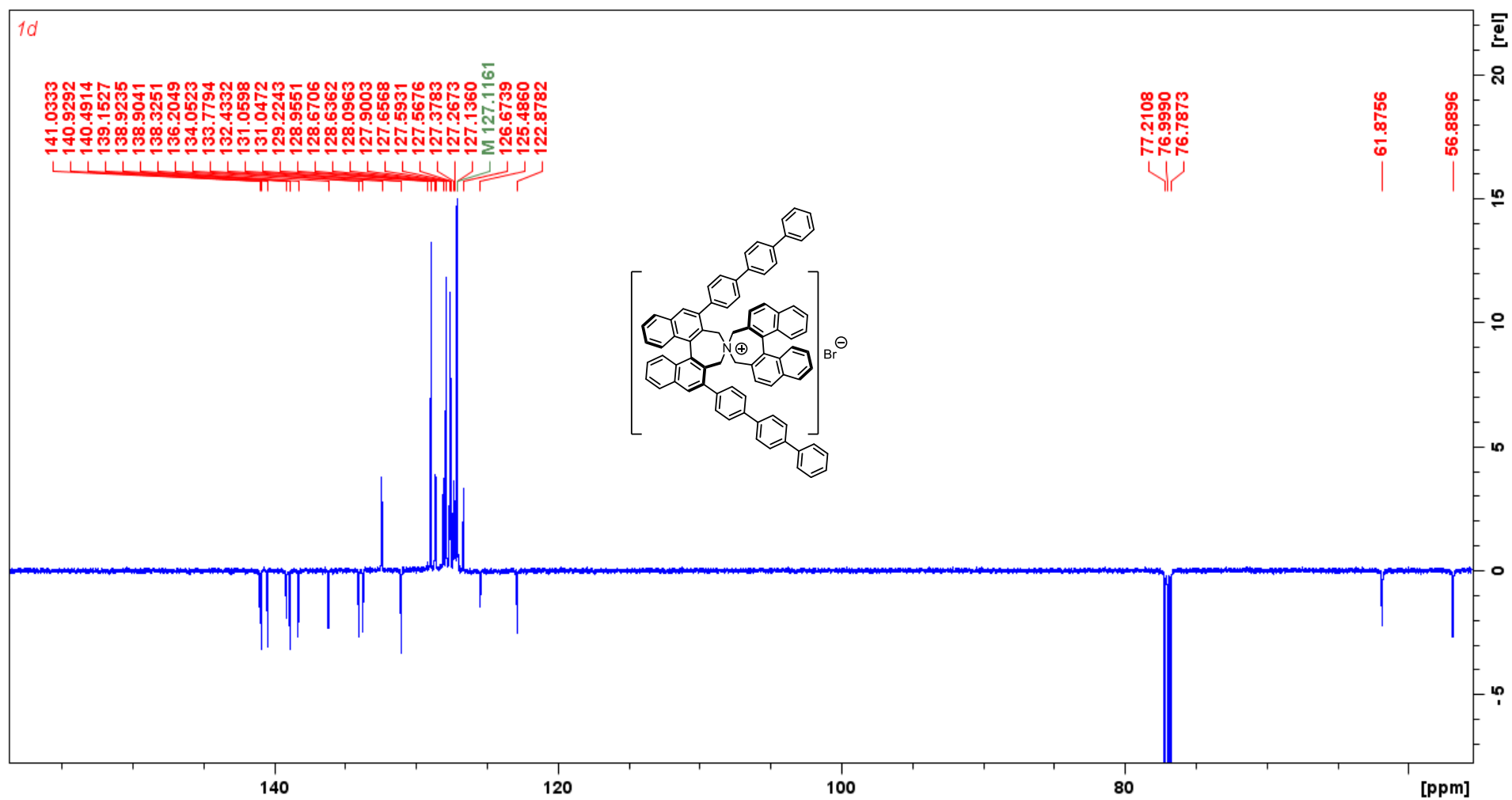


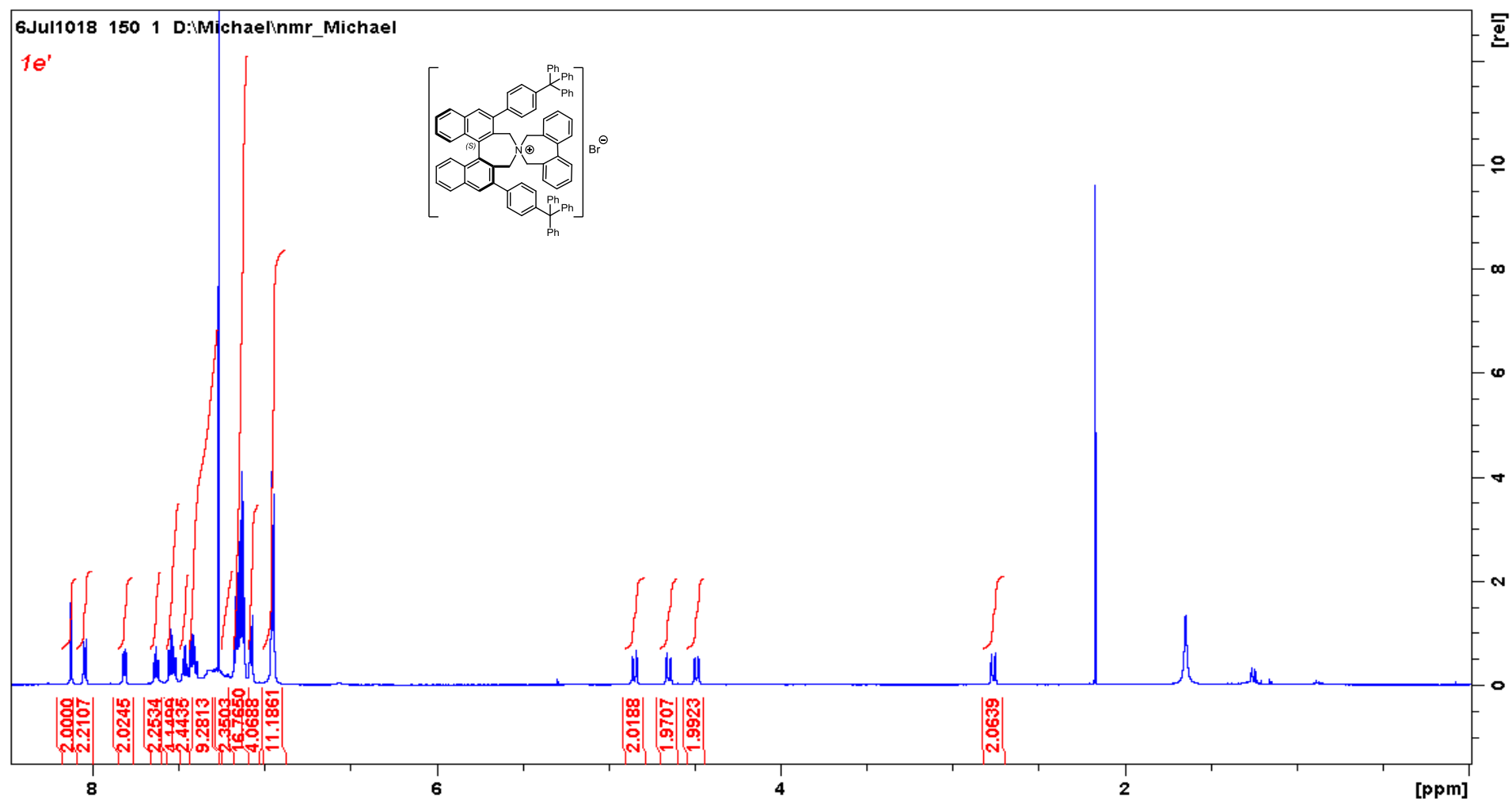


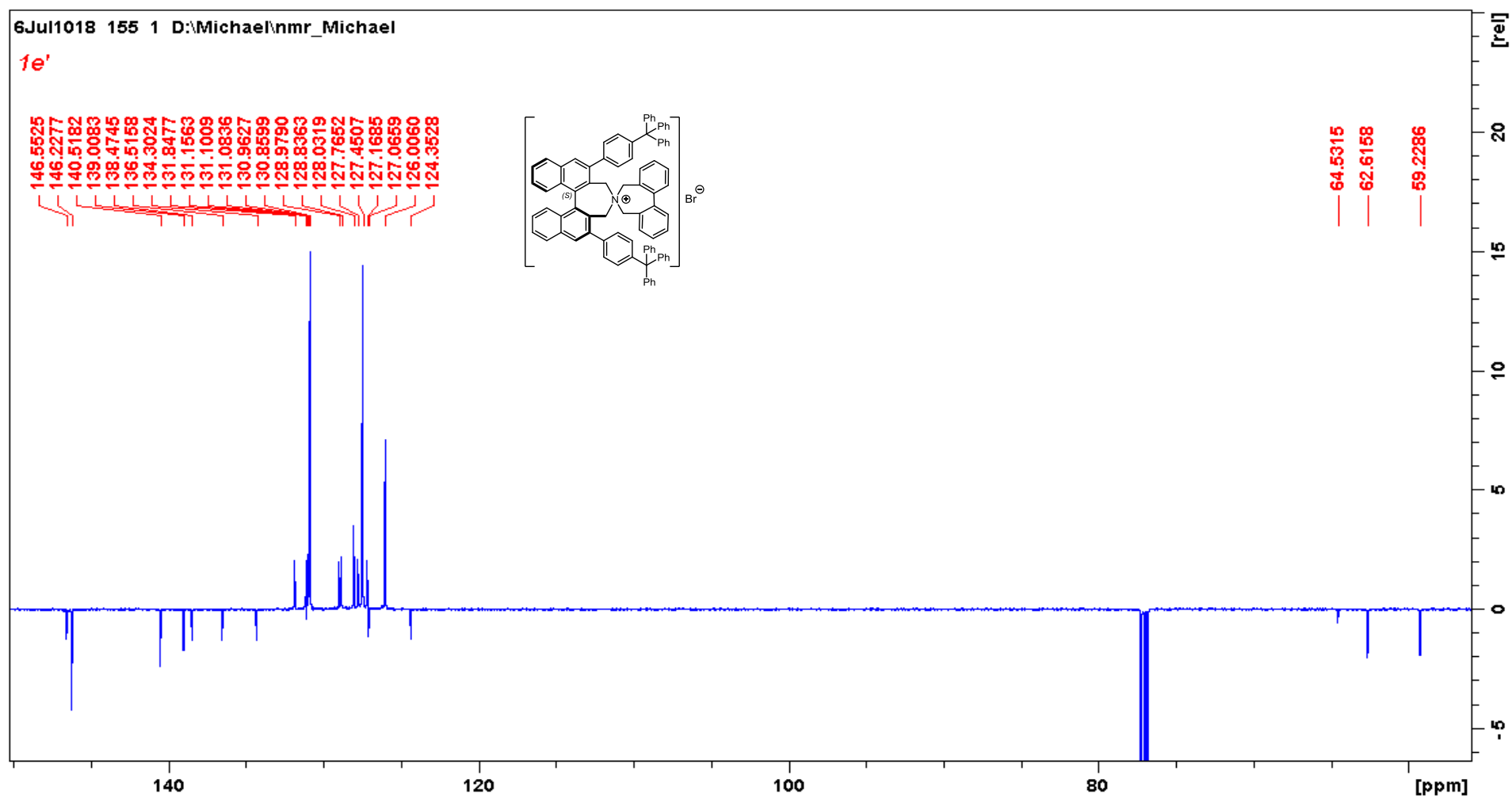
### $^1\text{H}$ - and $^{13}\text{C}$ NMR spectra

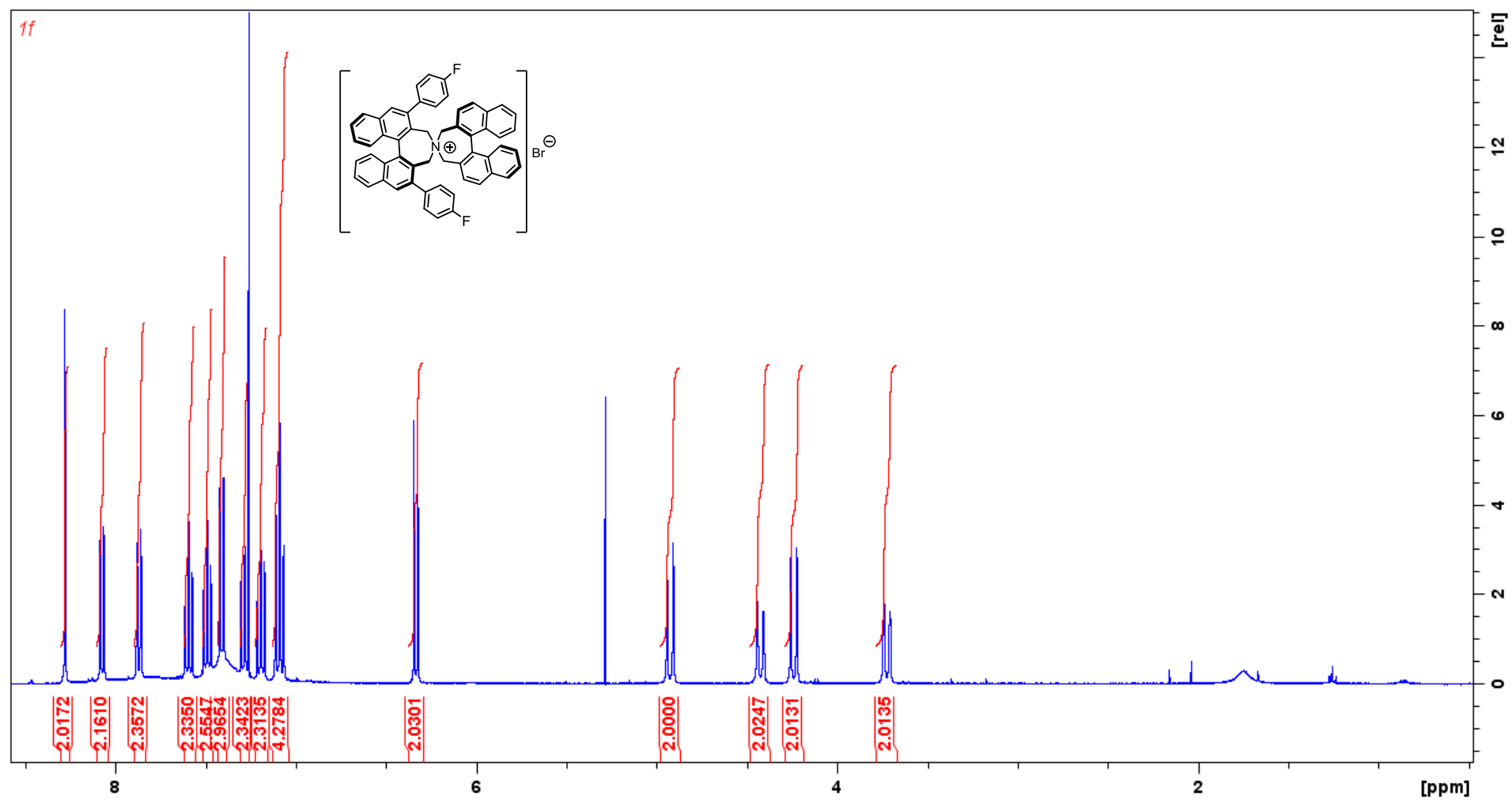
(If not otherwise noted spectra are recorded at room temperature in  $\text{CDCl}_3$ )

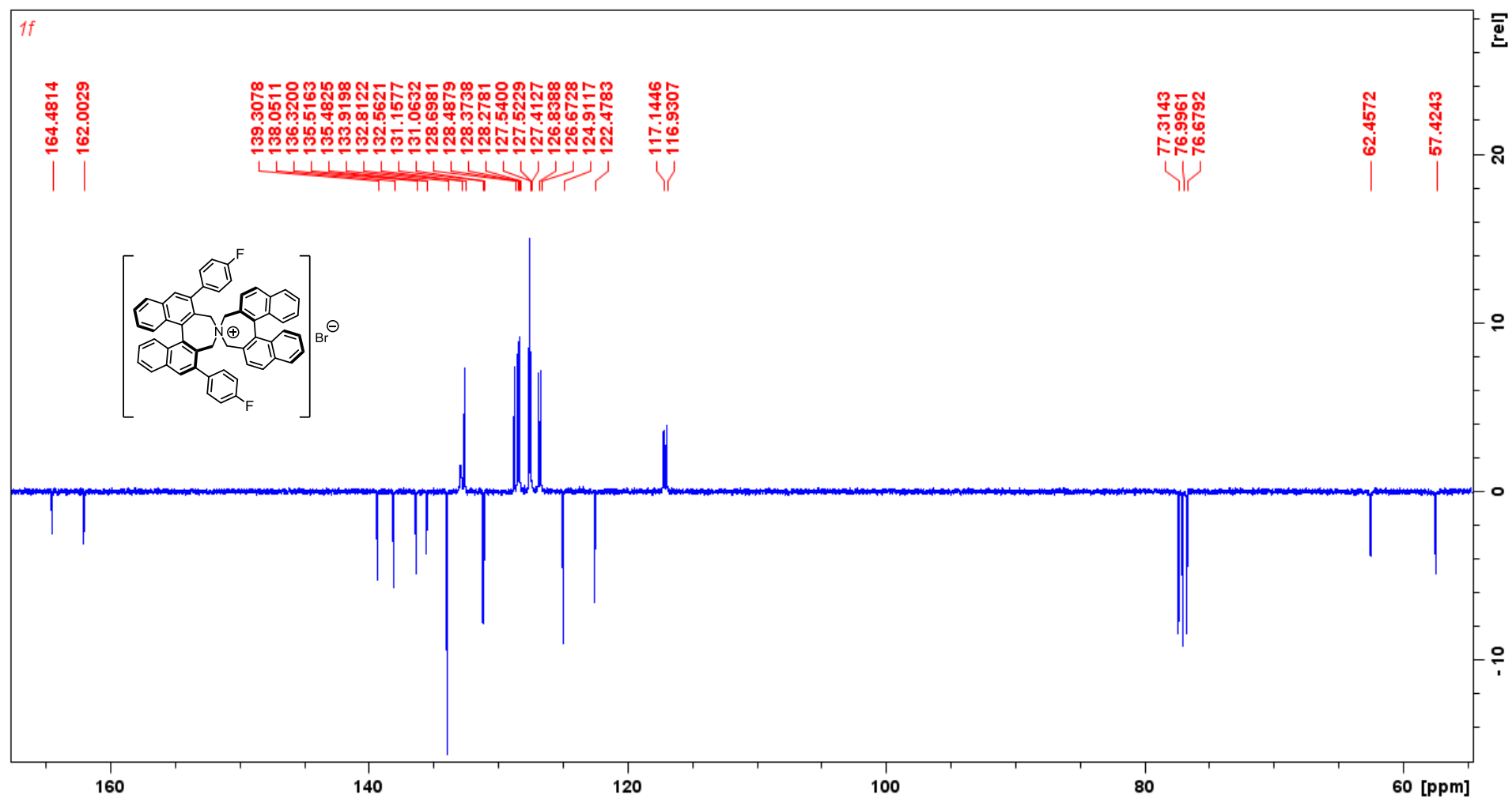


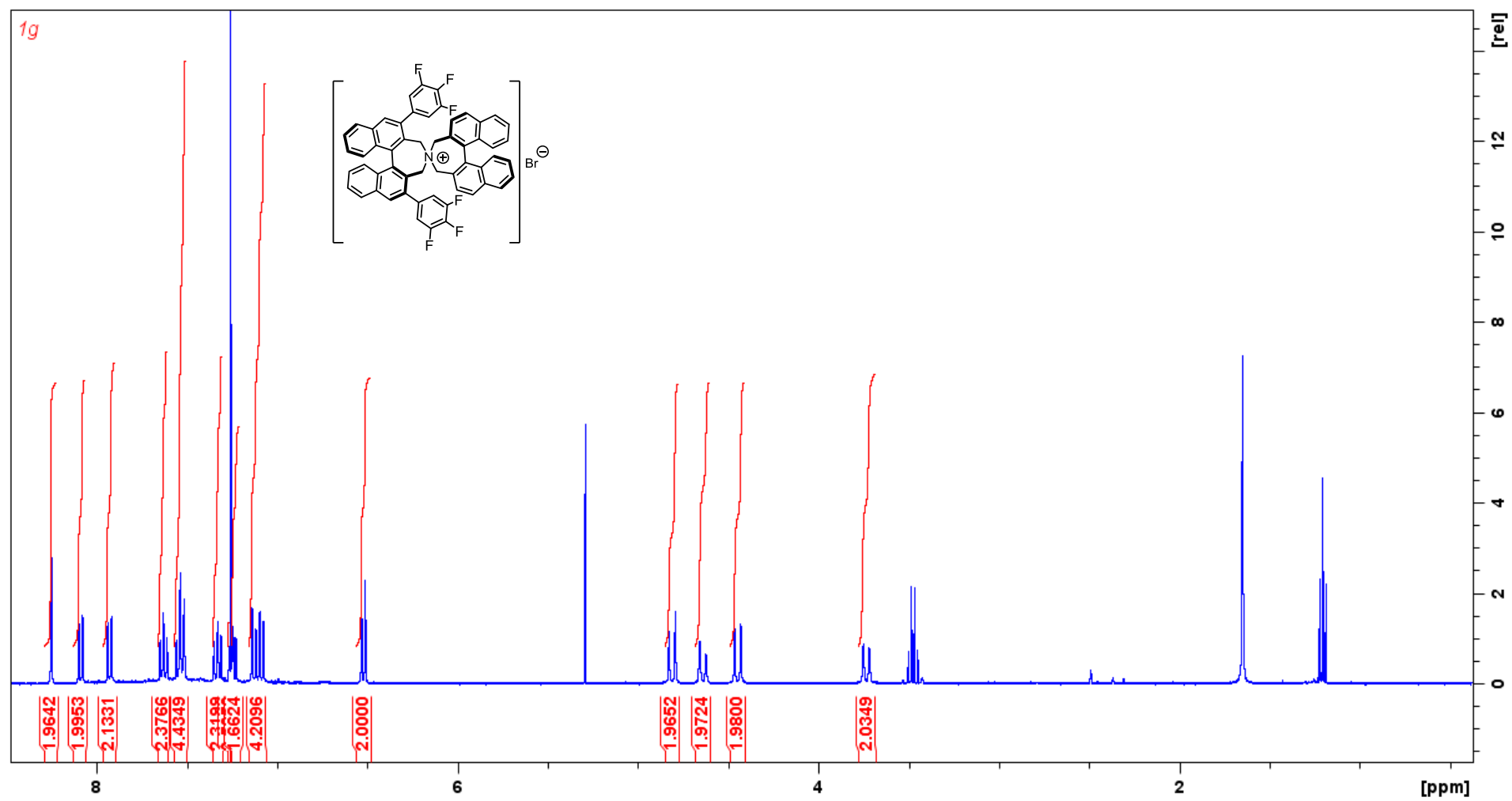




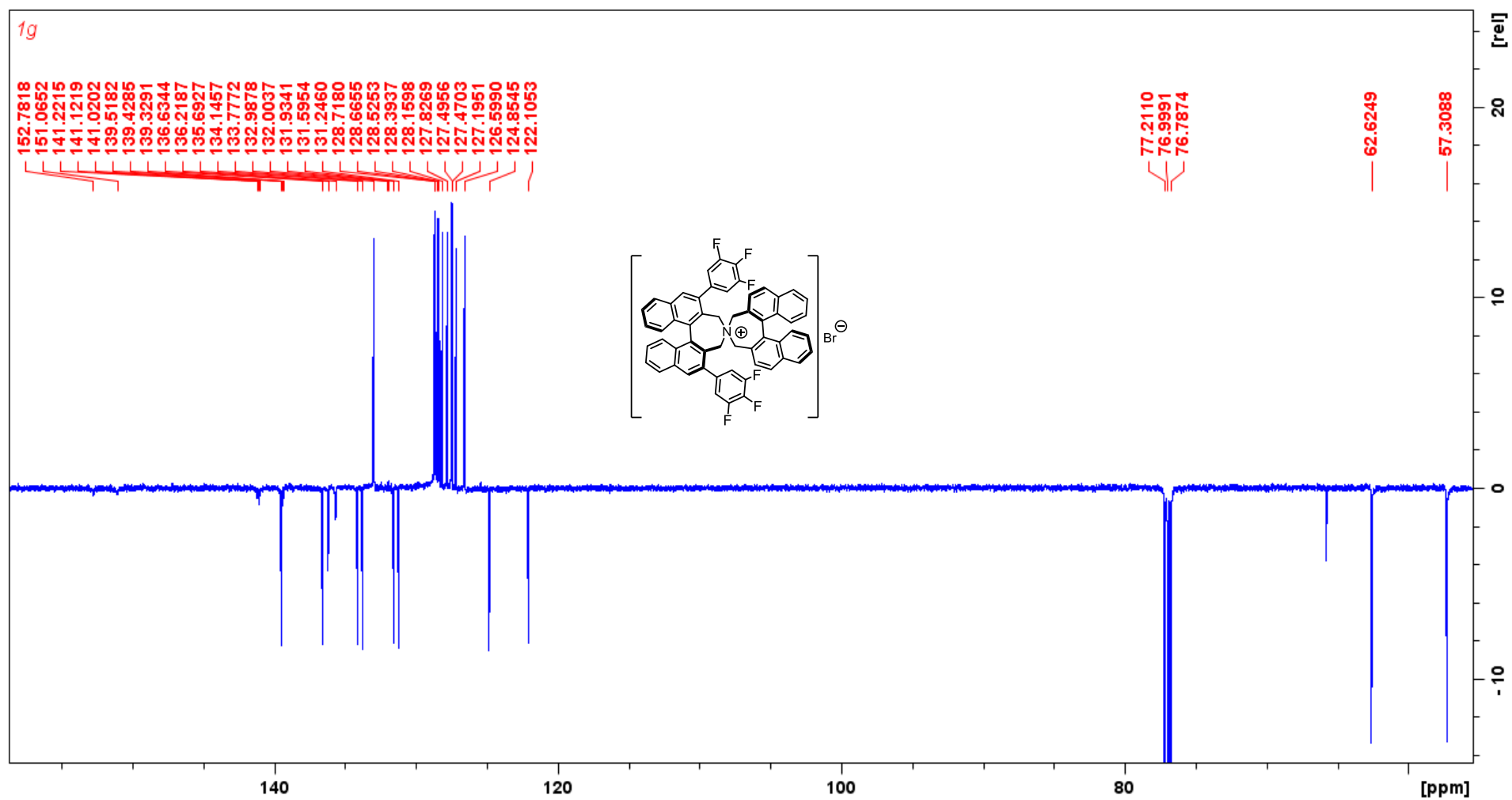


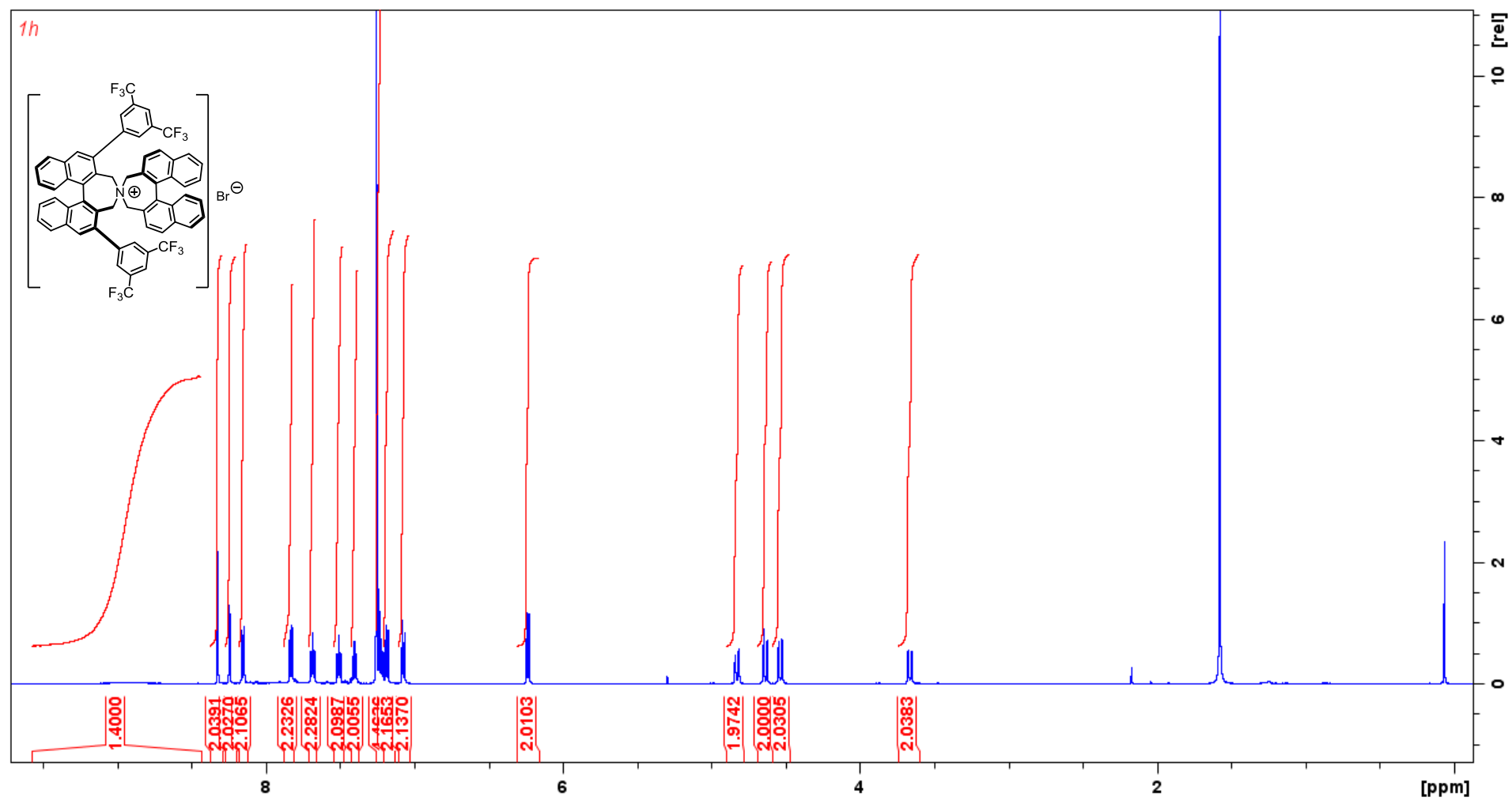


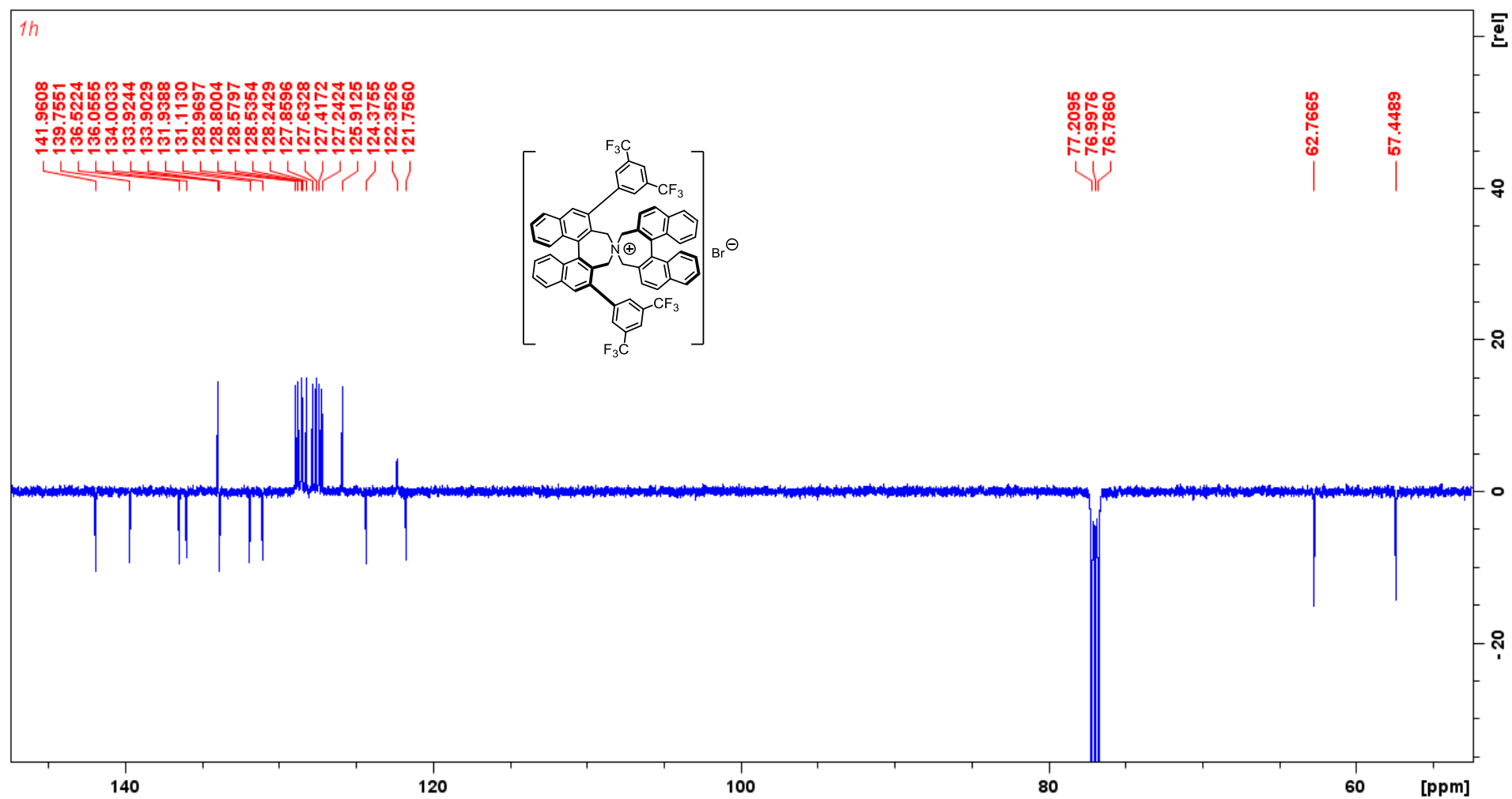


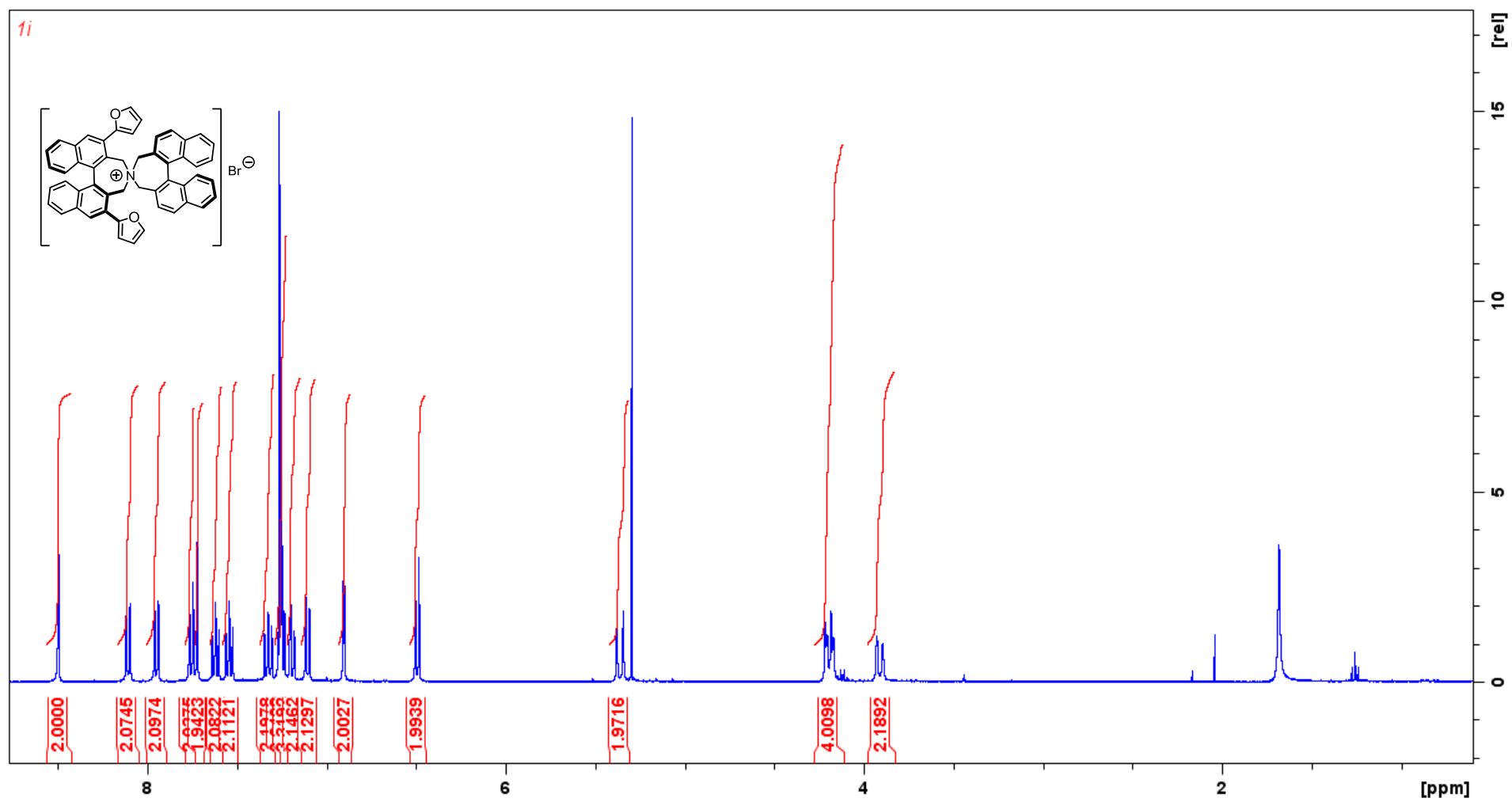


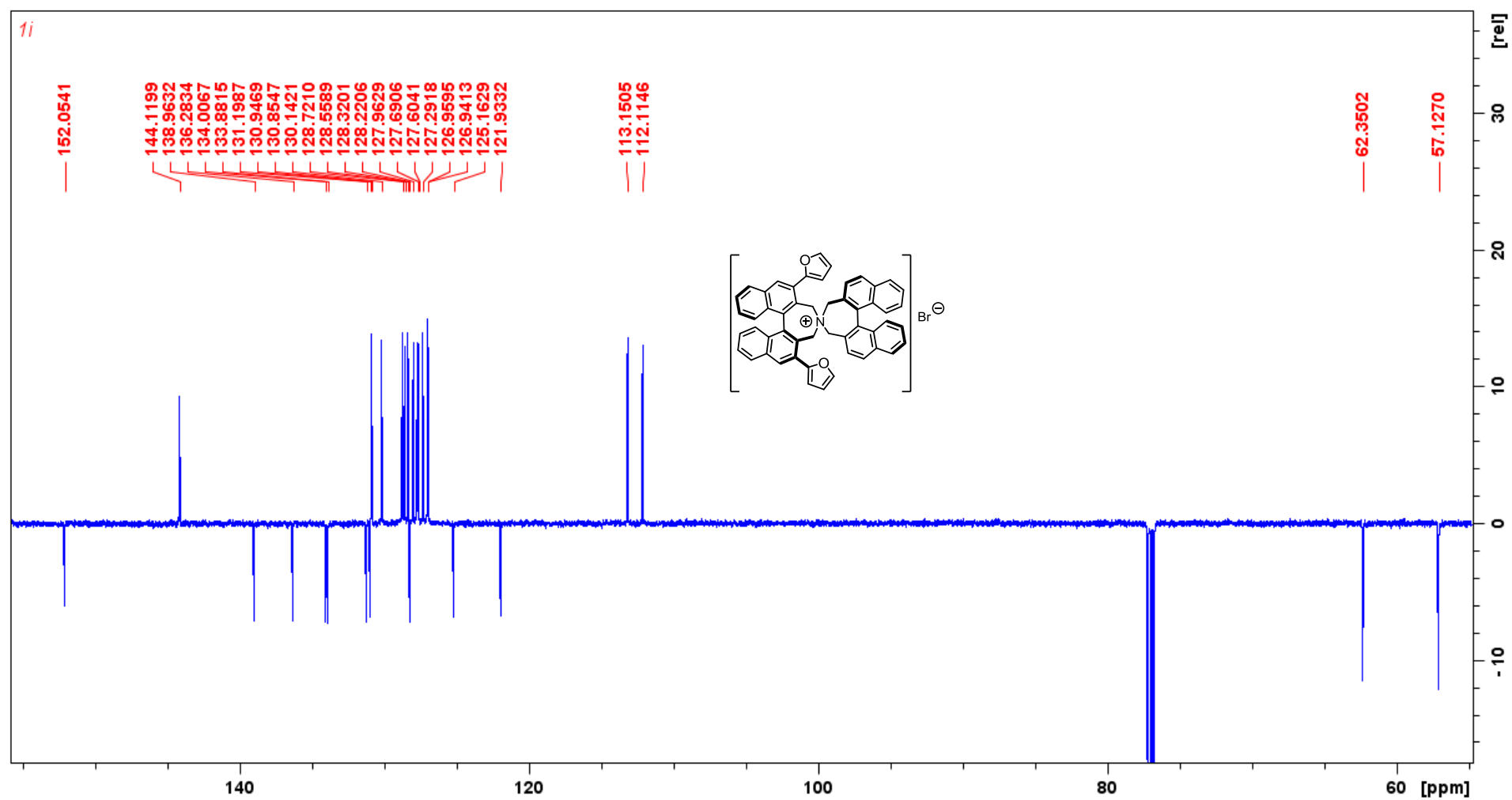


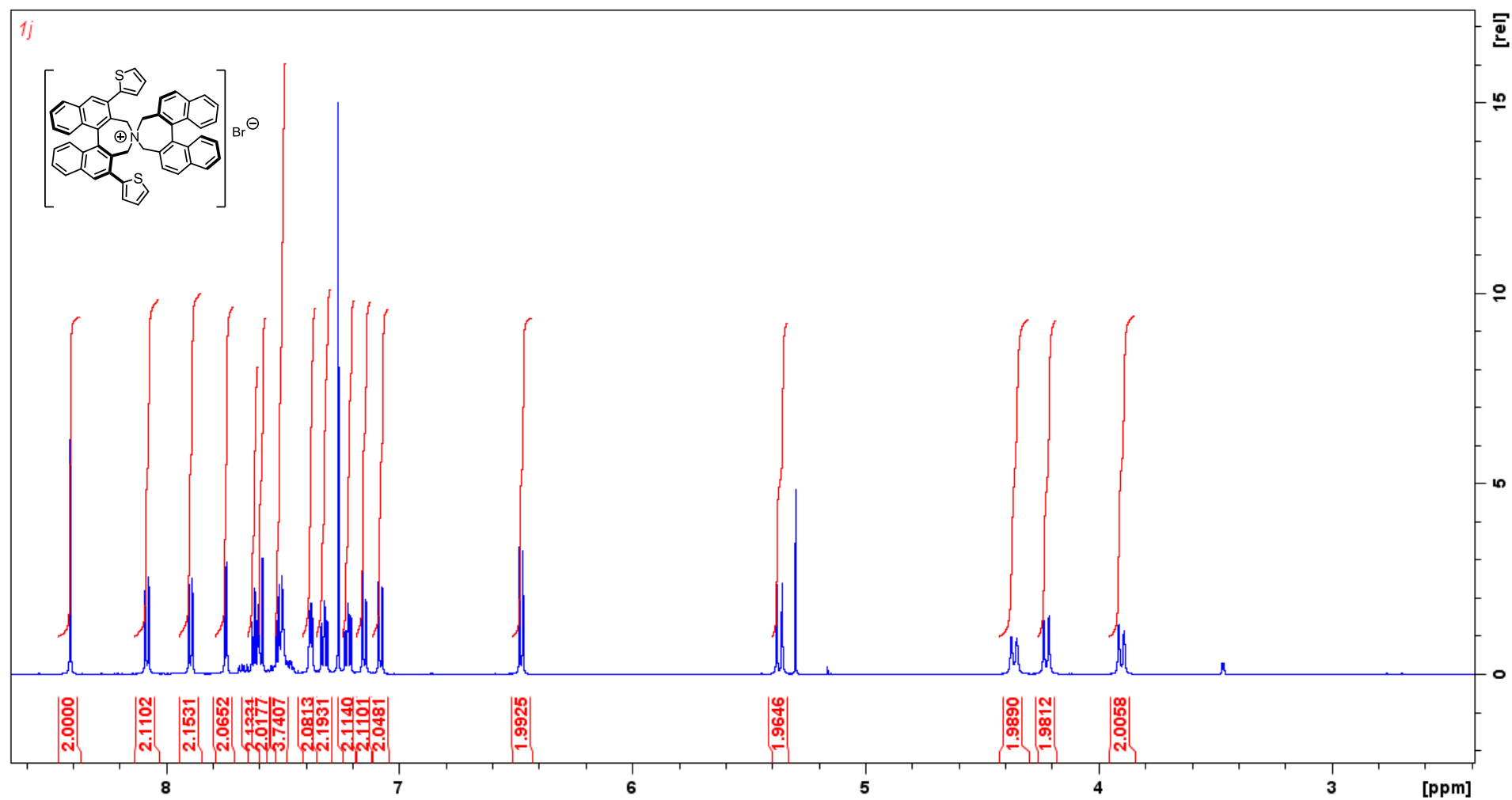


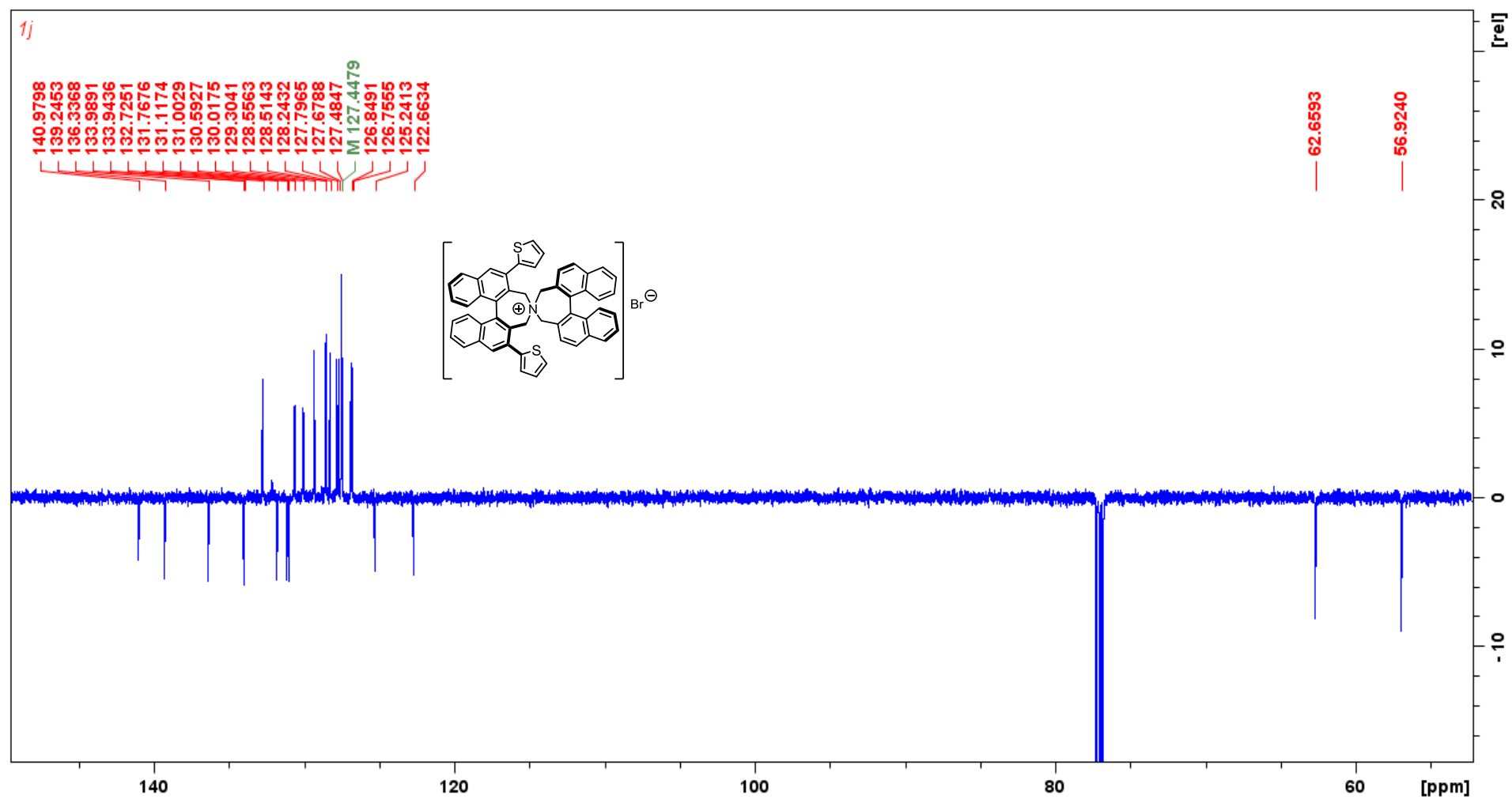


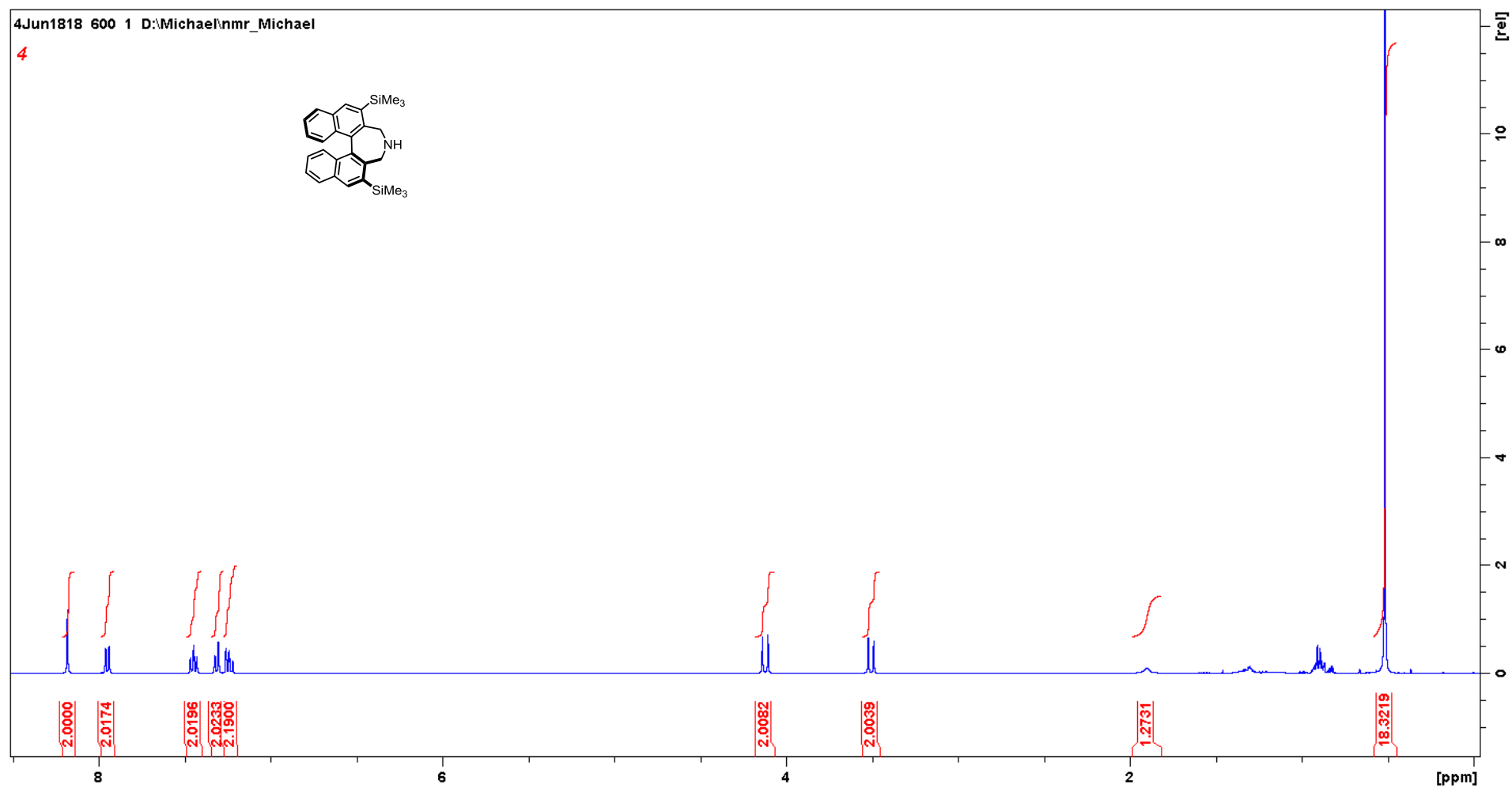




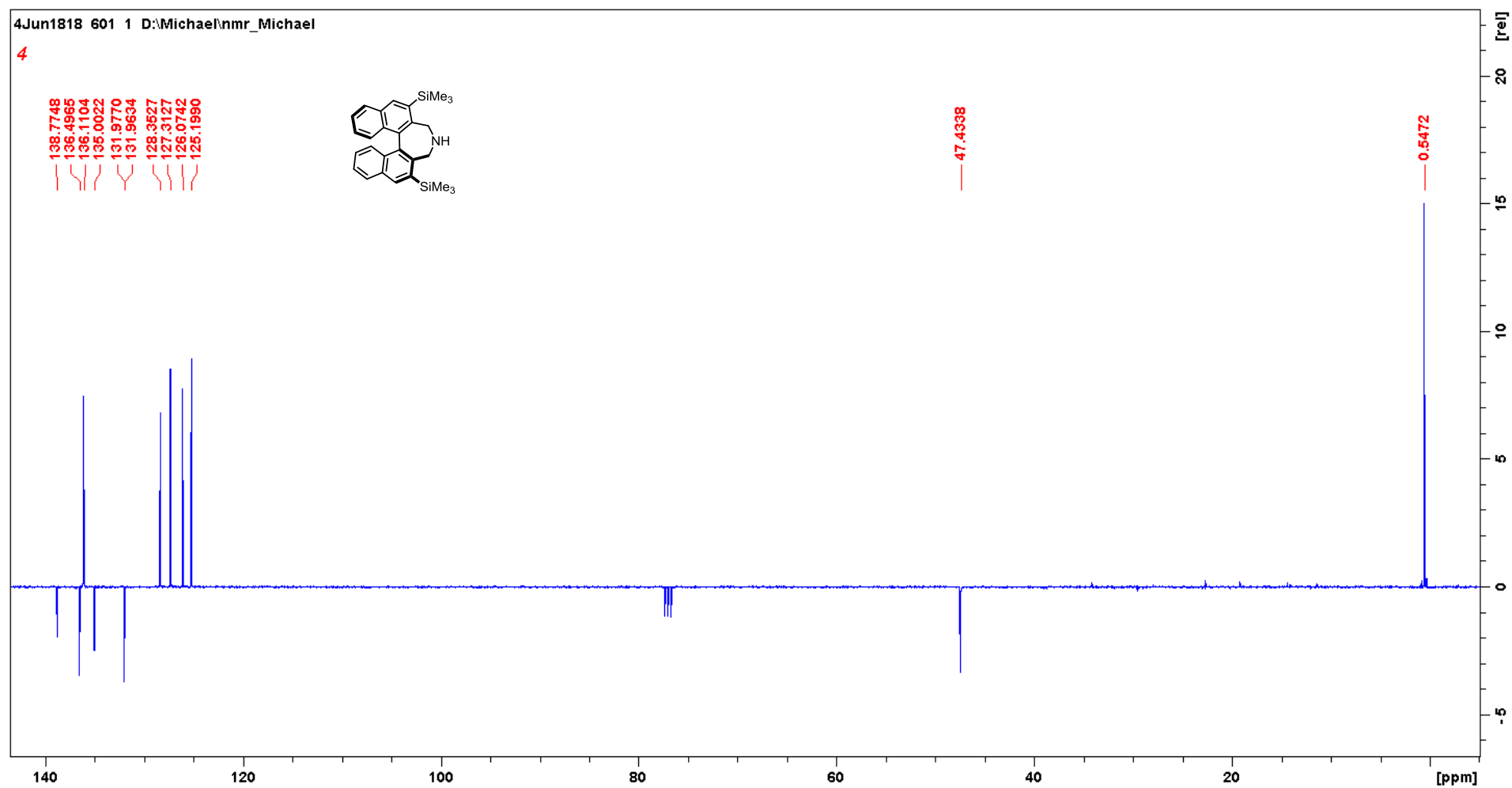


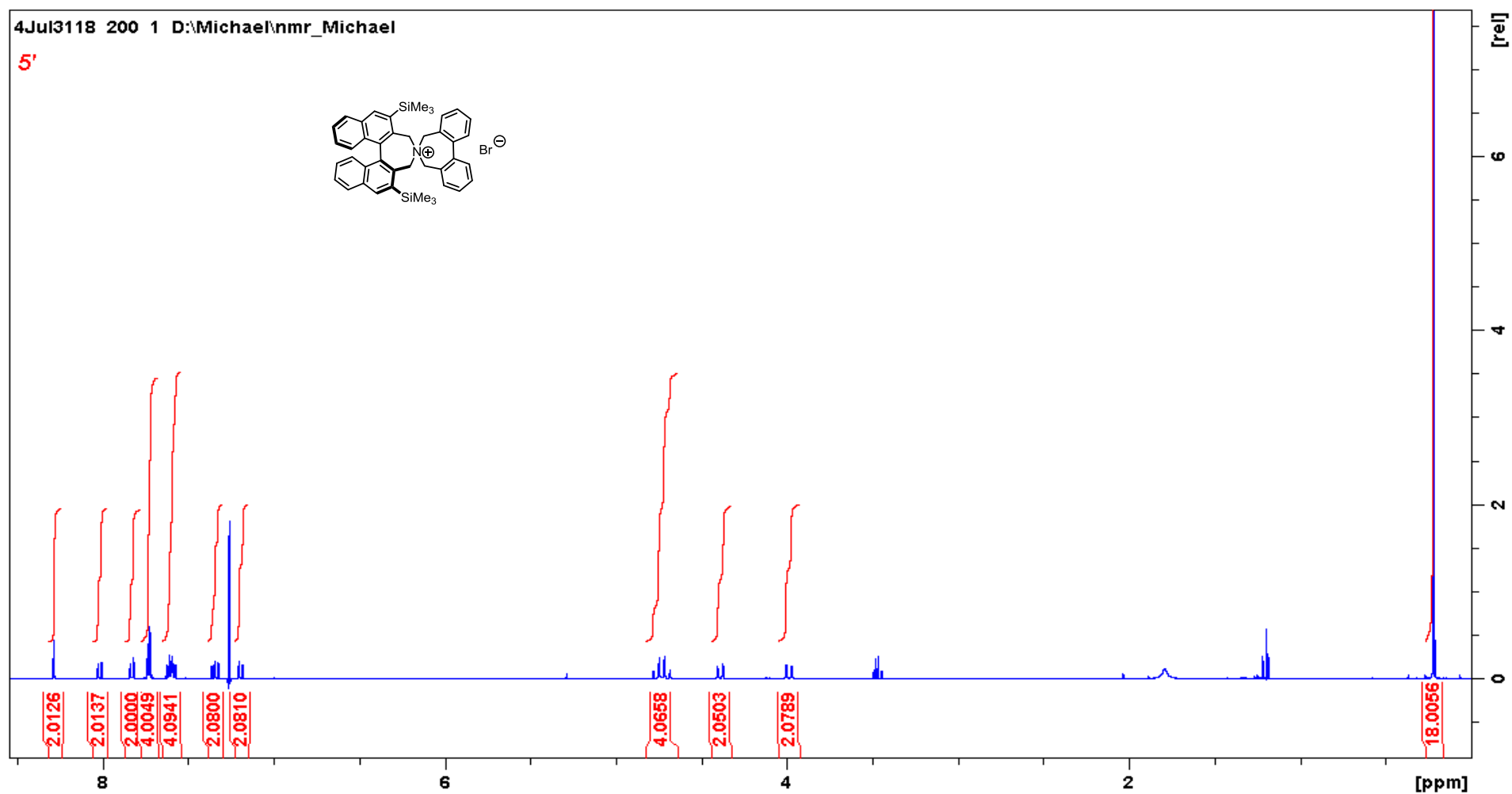


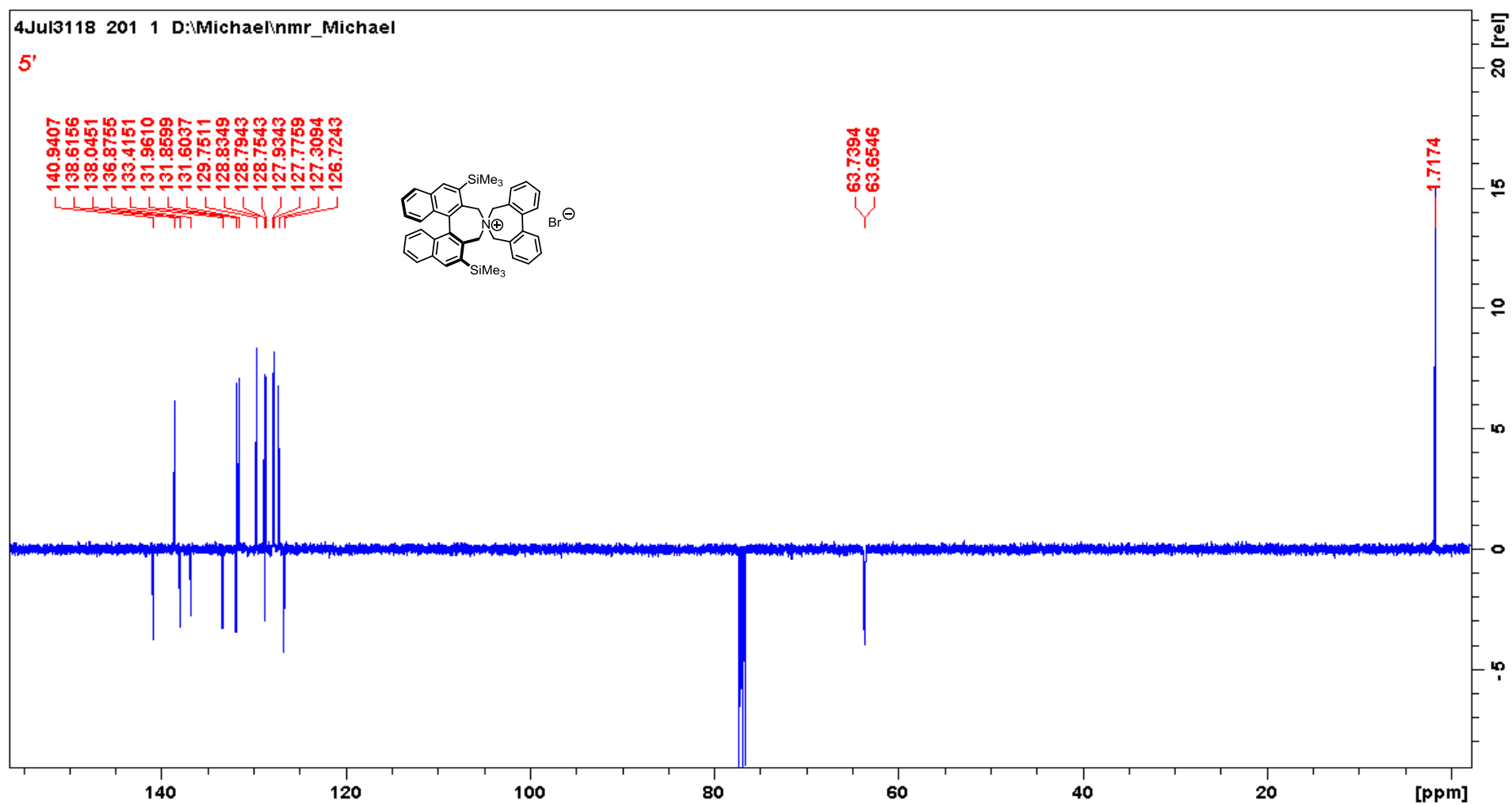


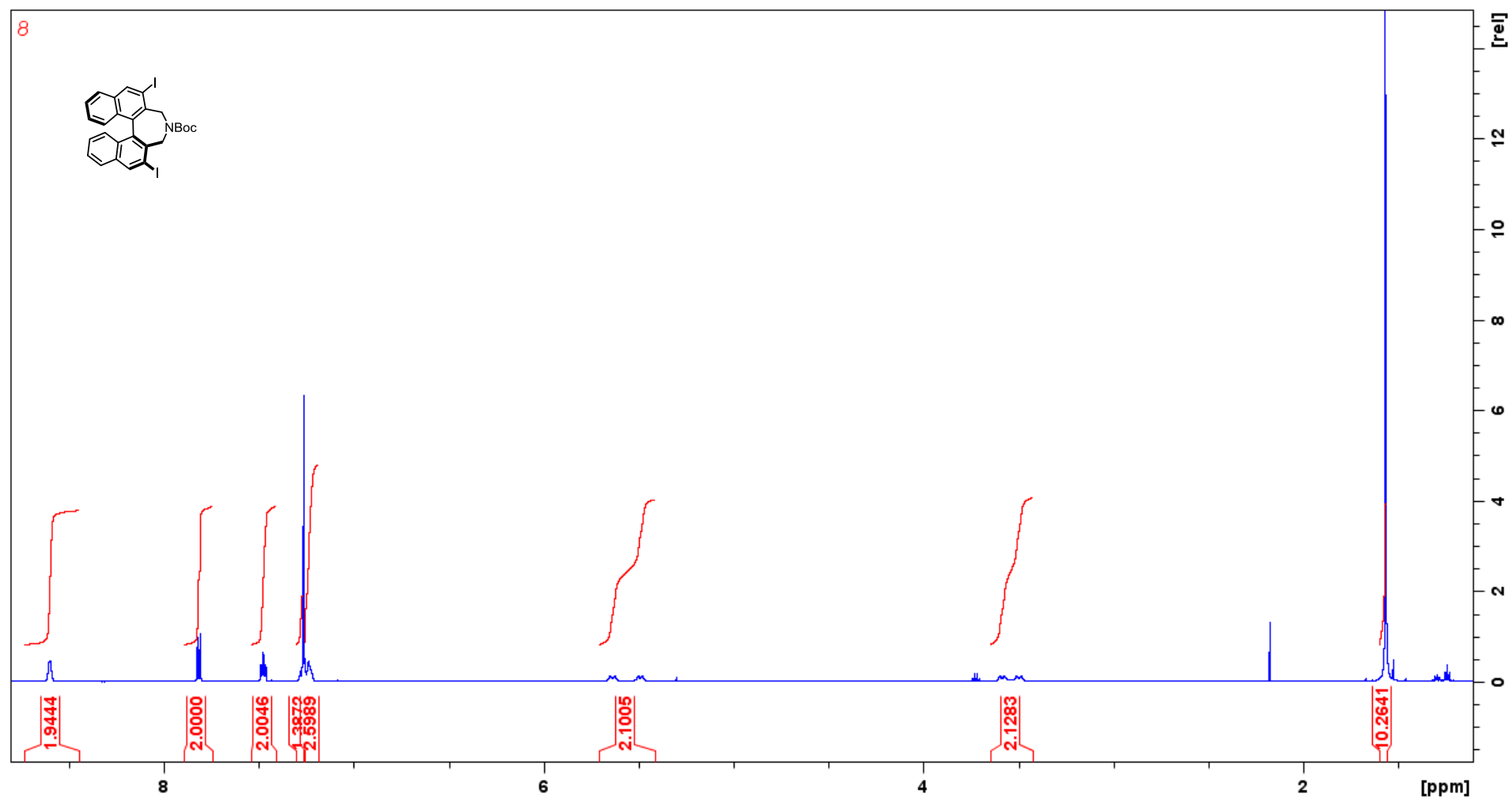


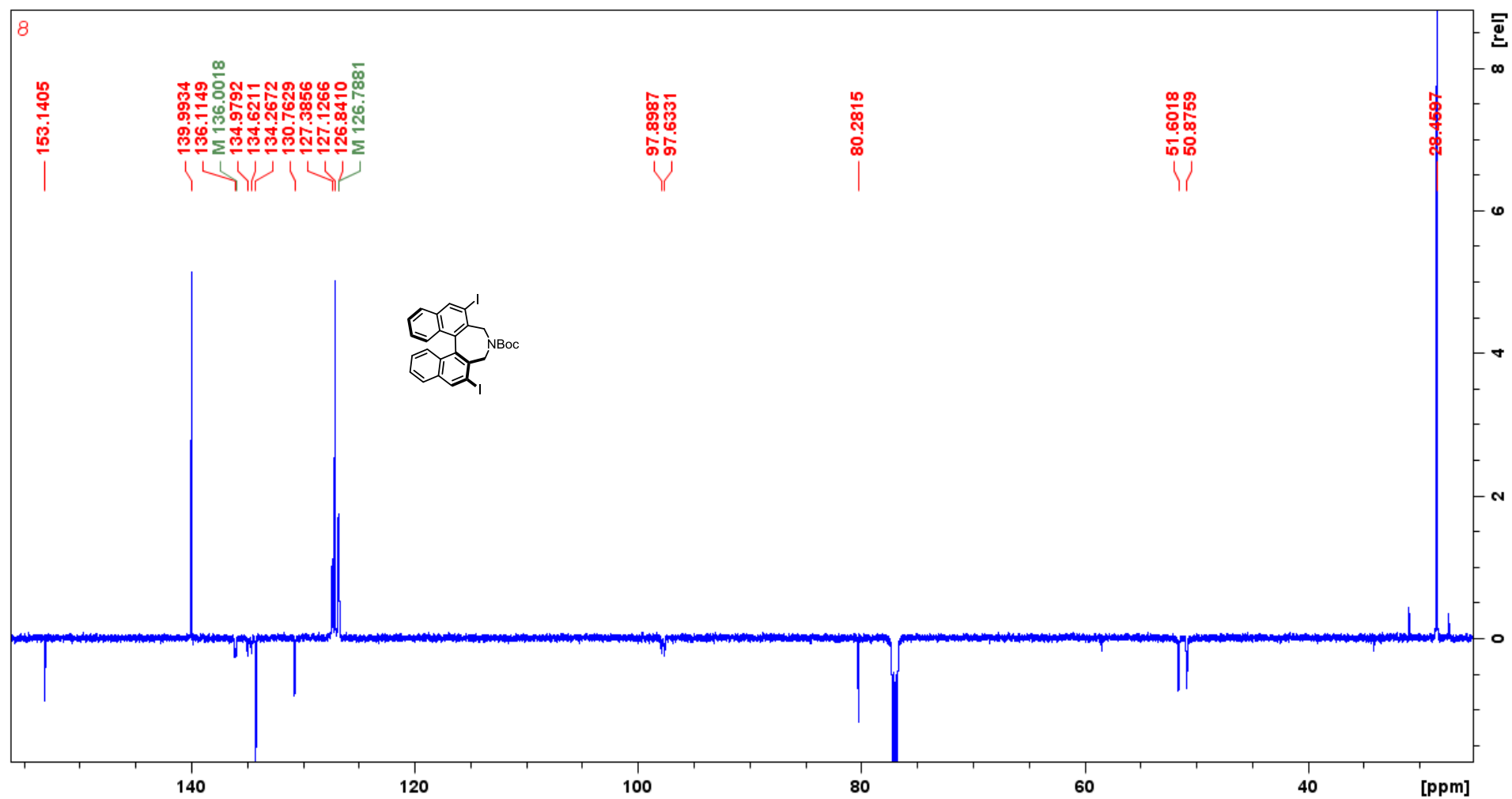


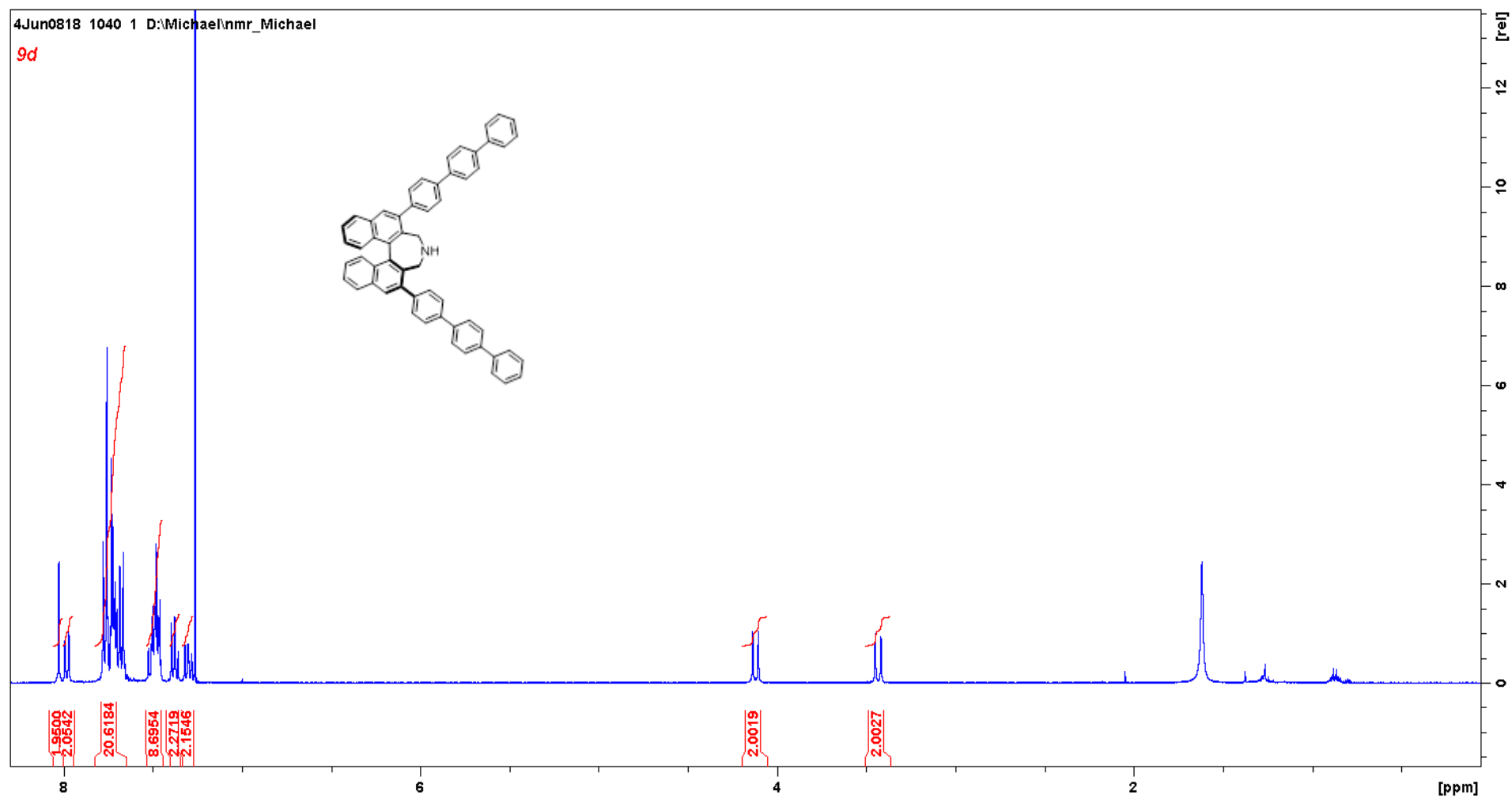


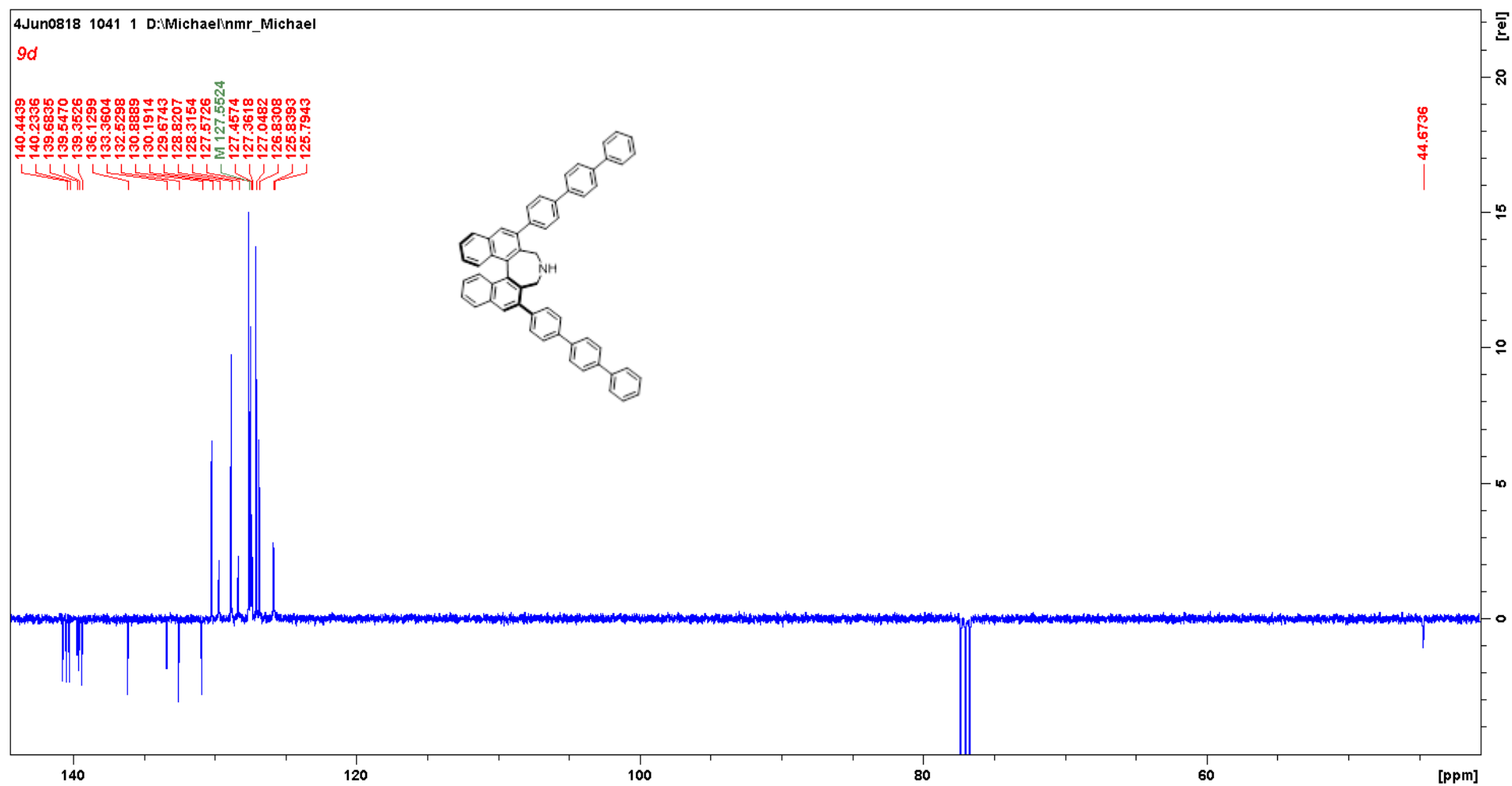


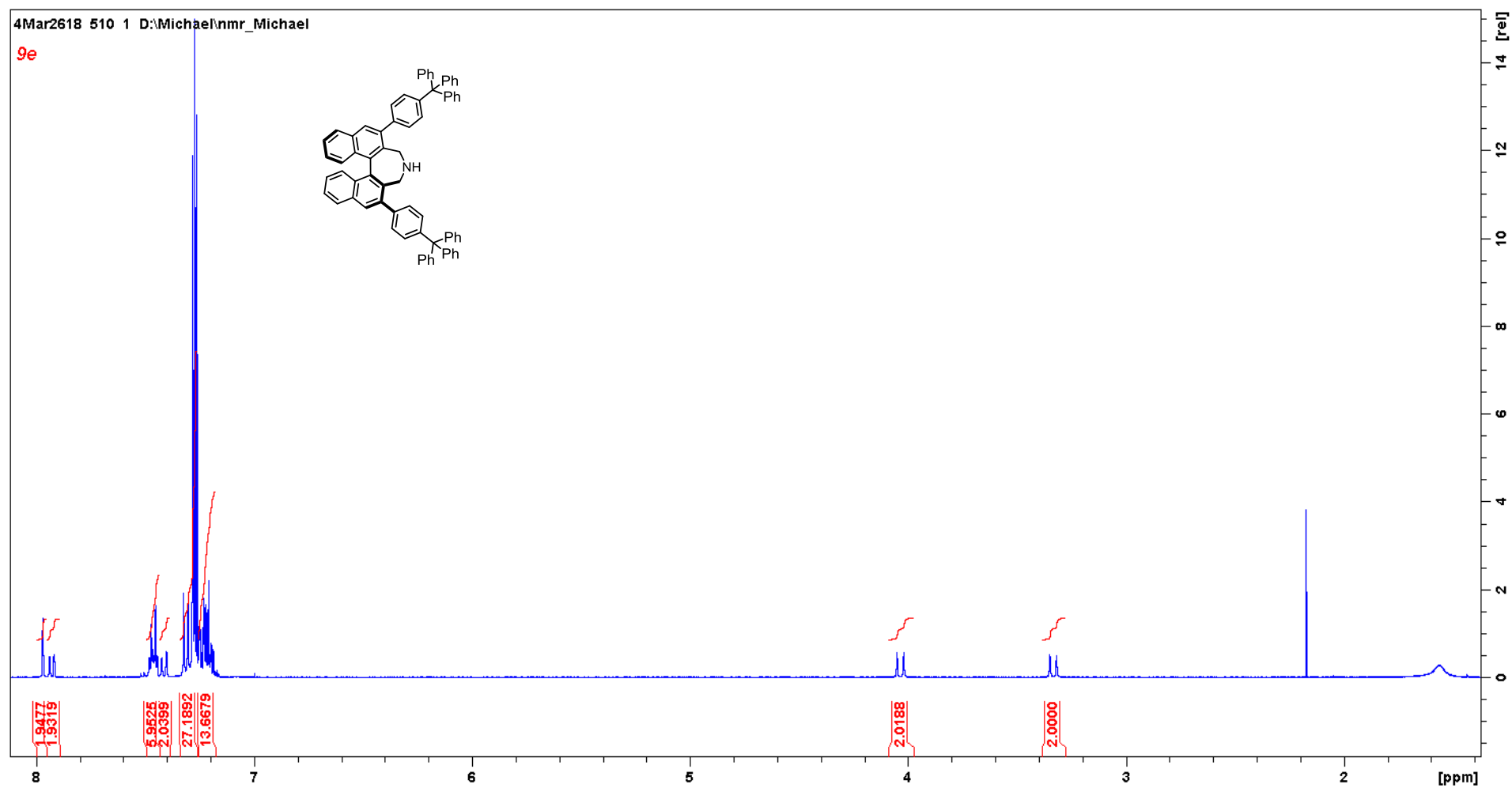




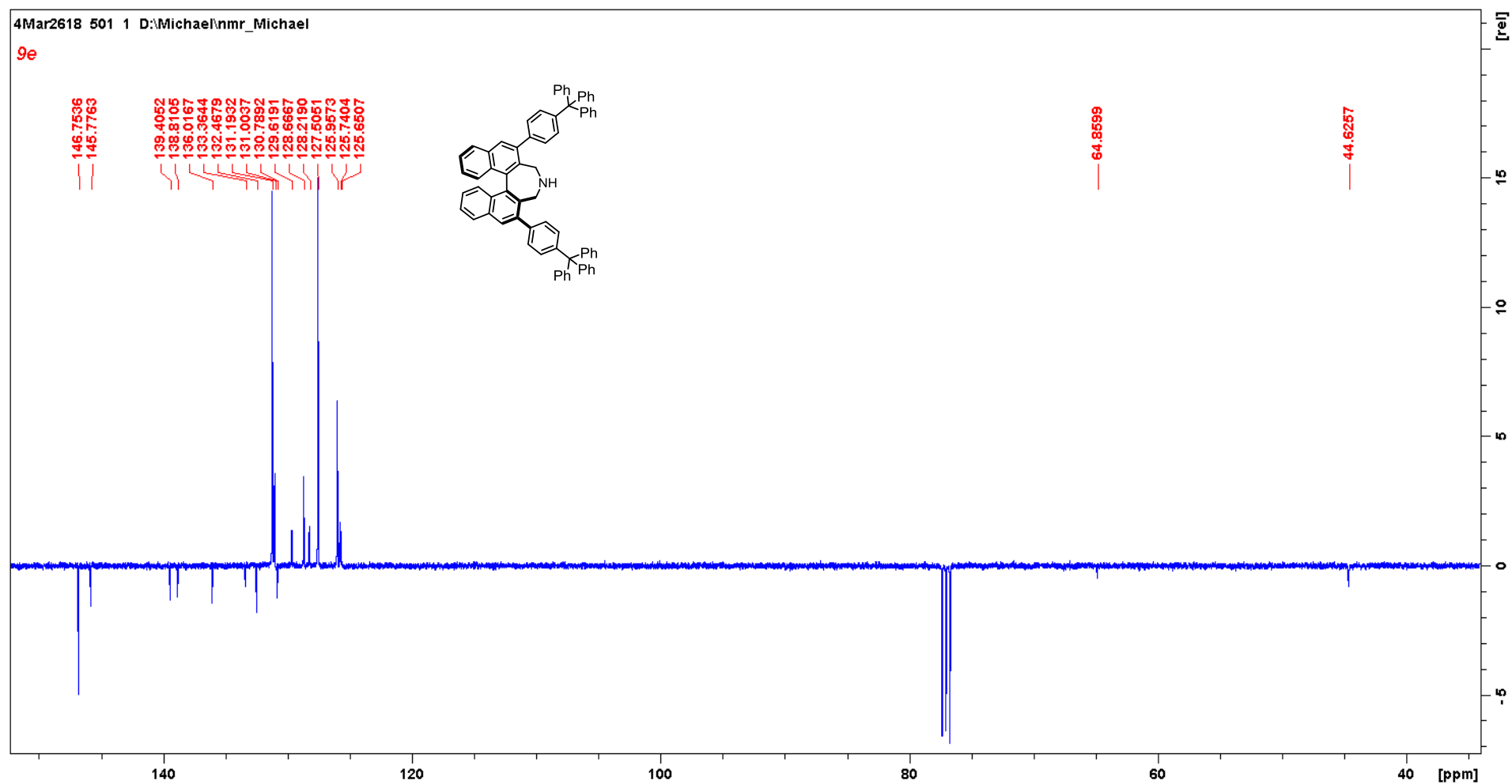


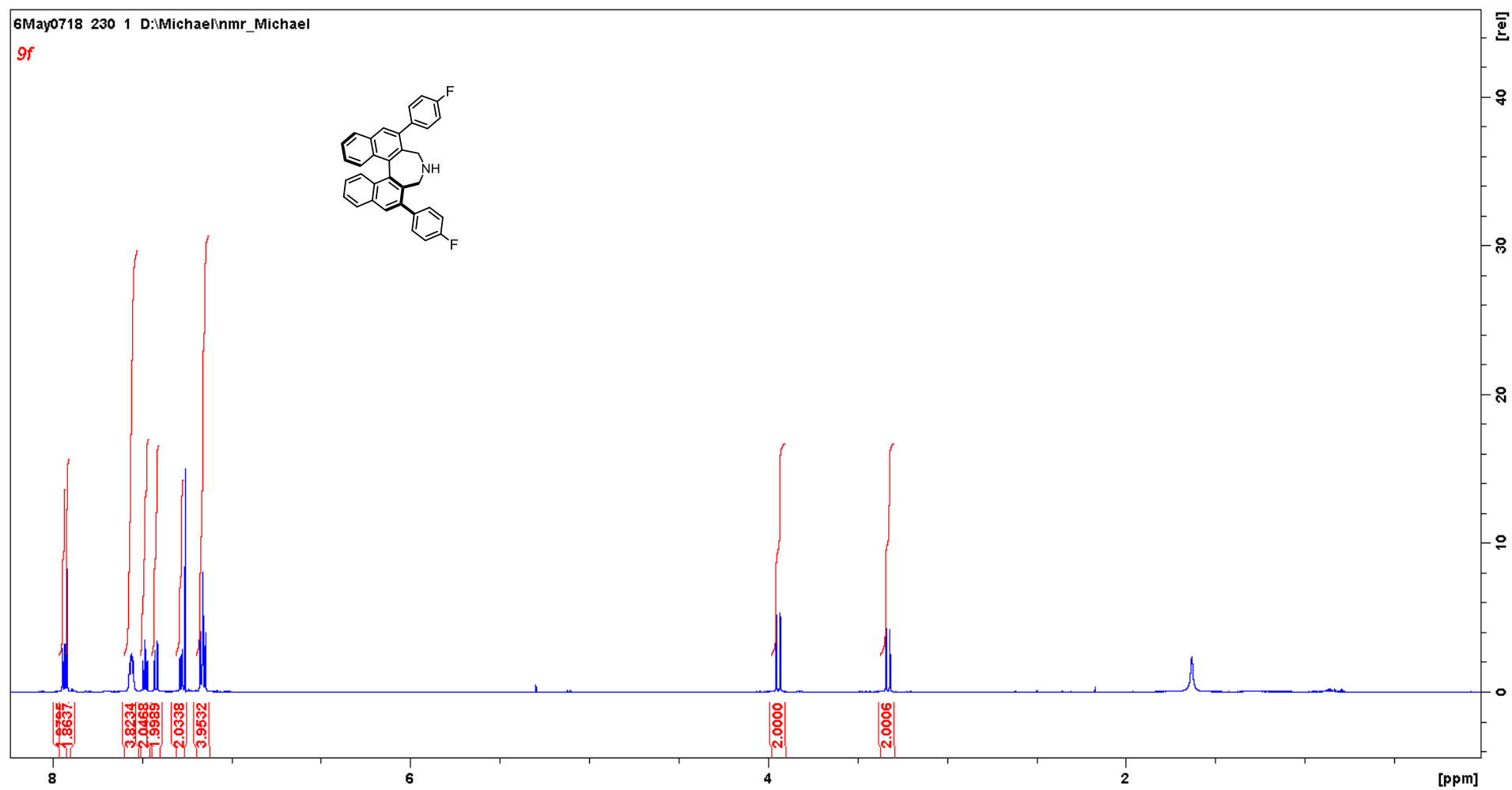


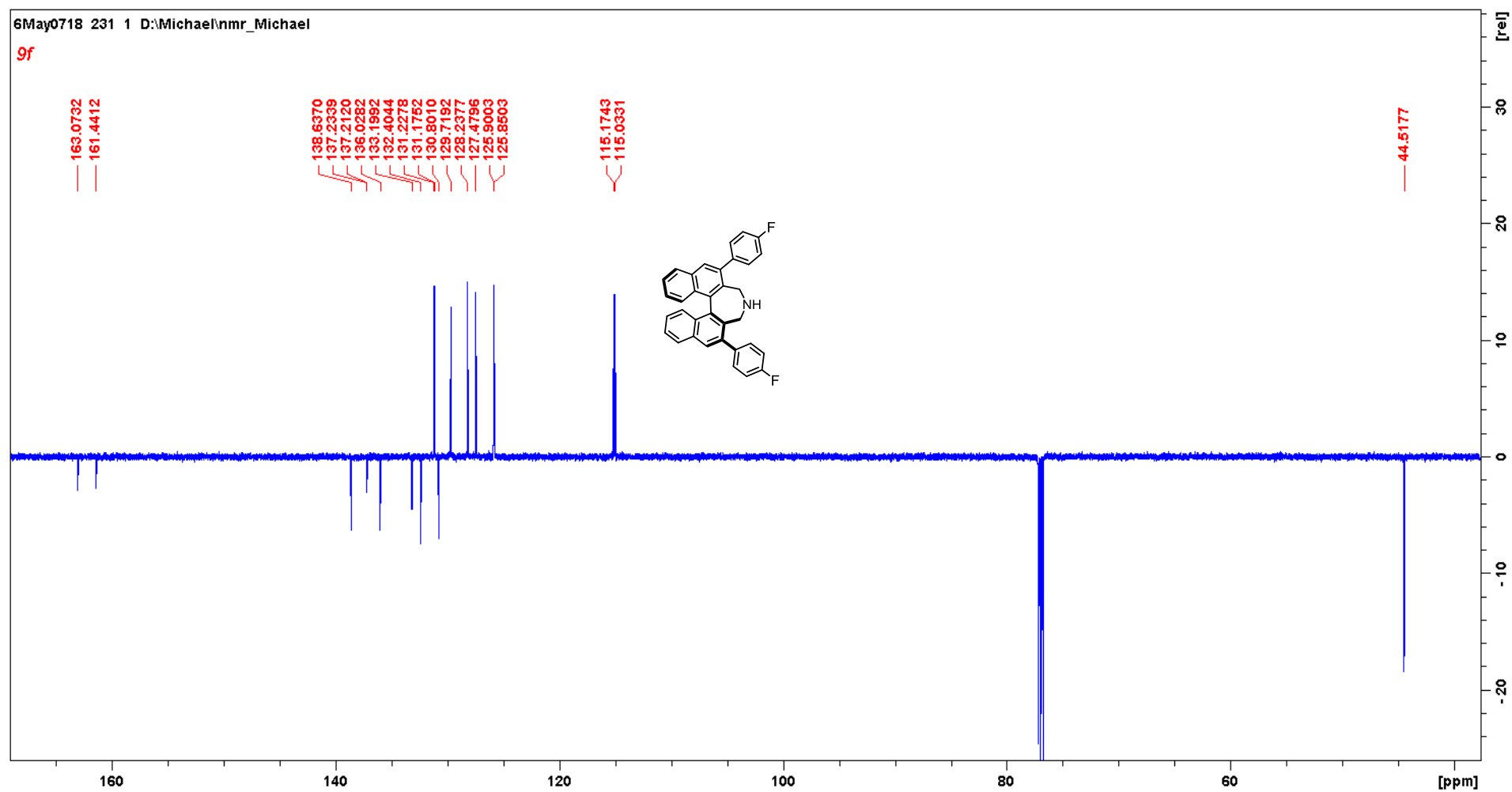


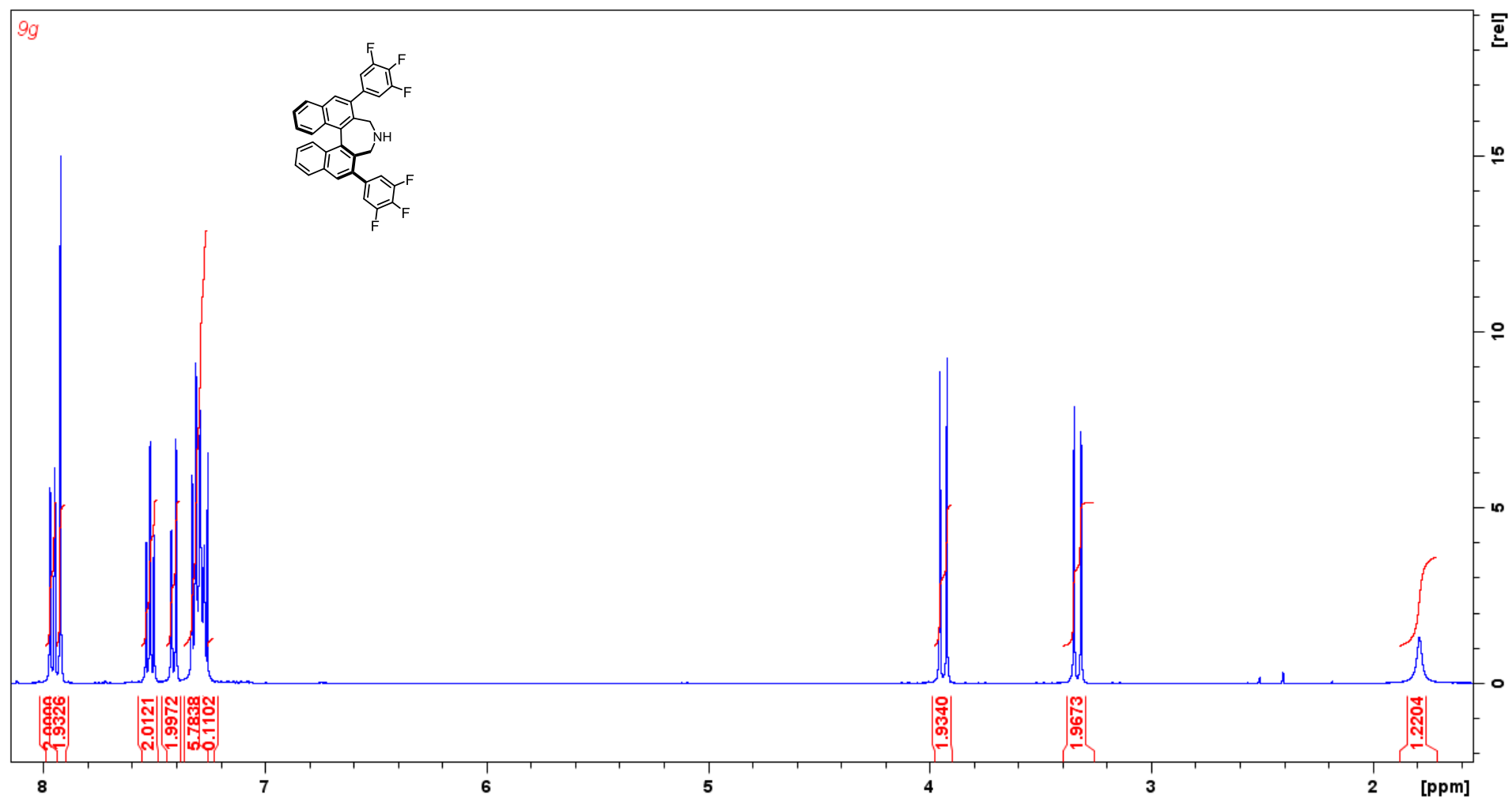


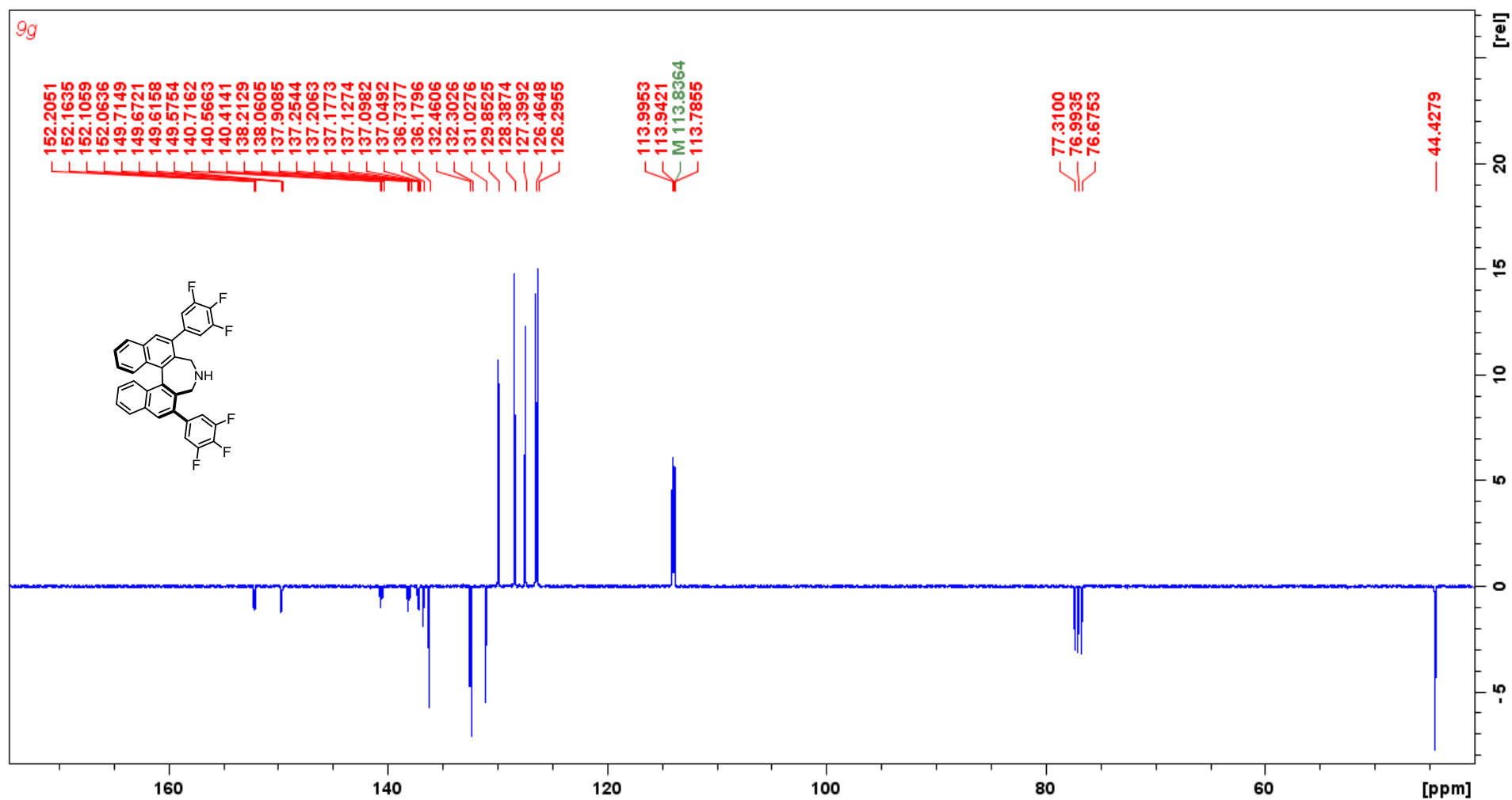


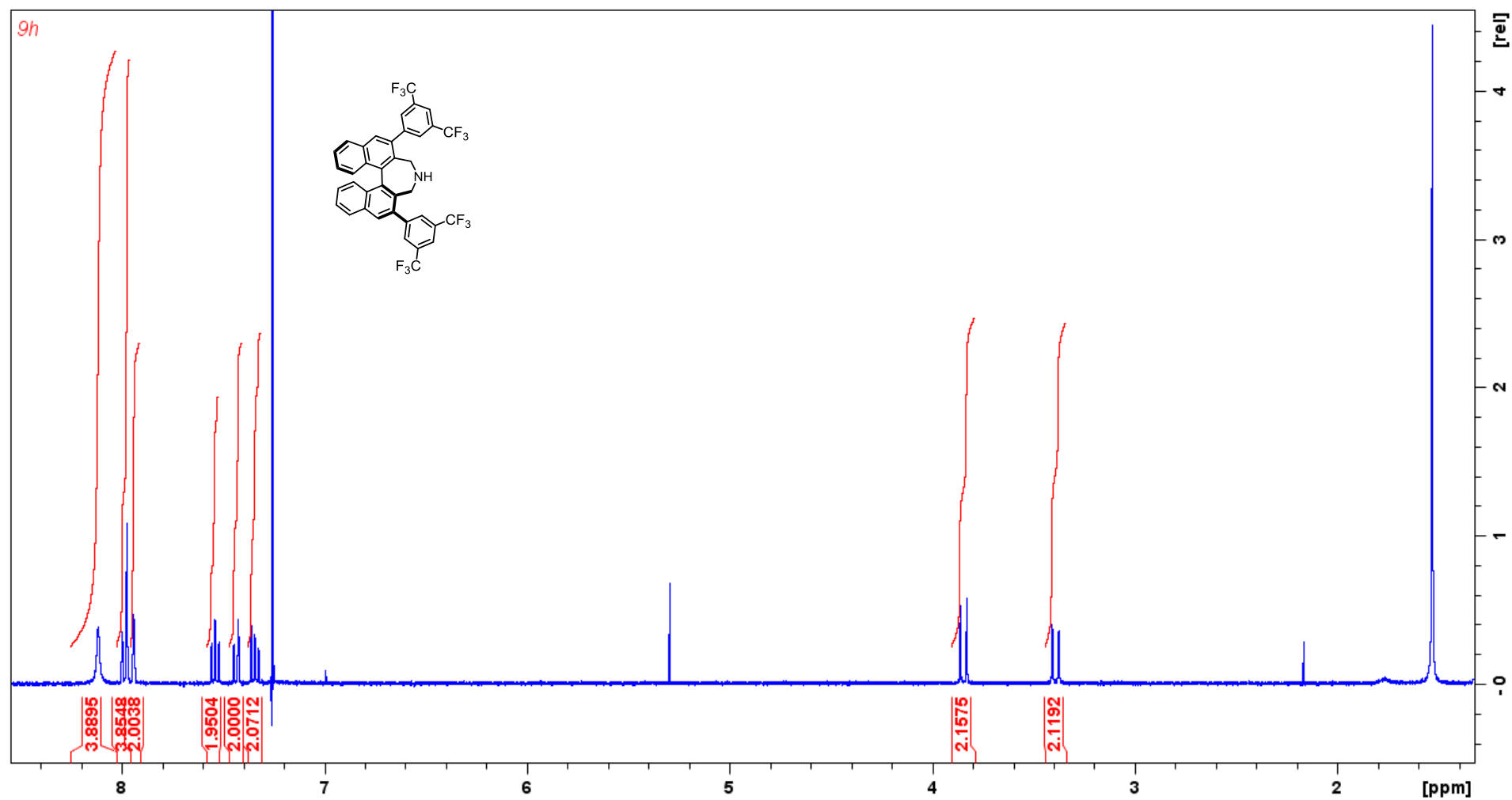


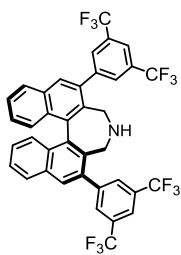


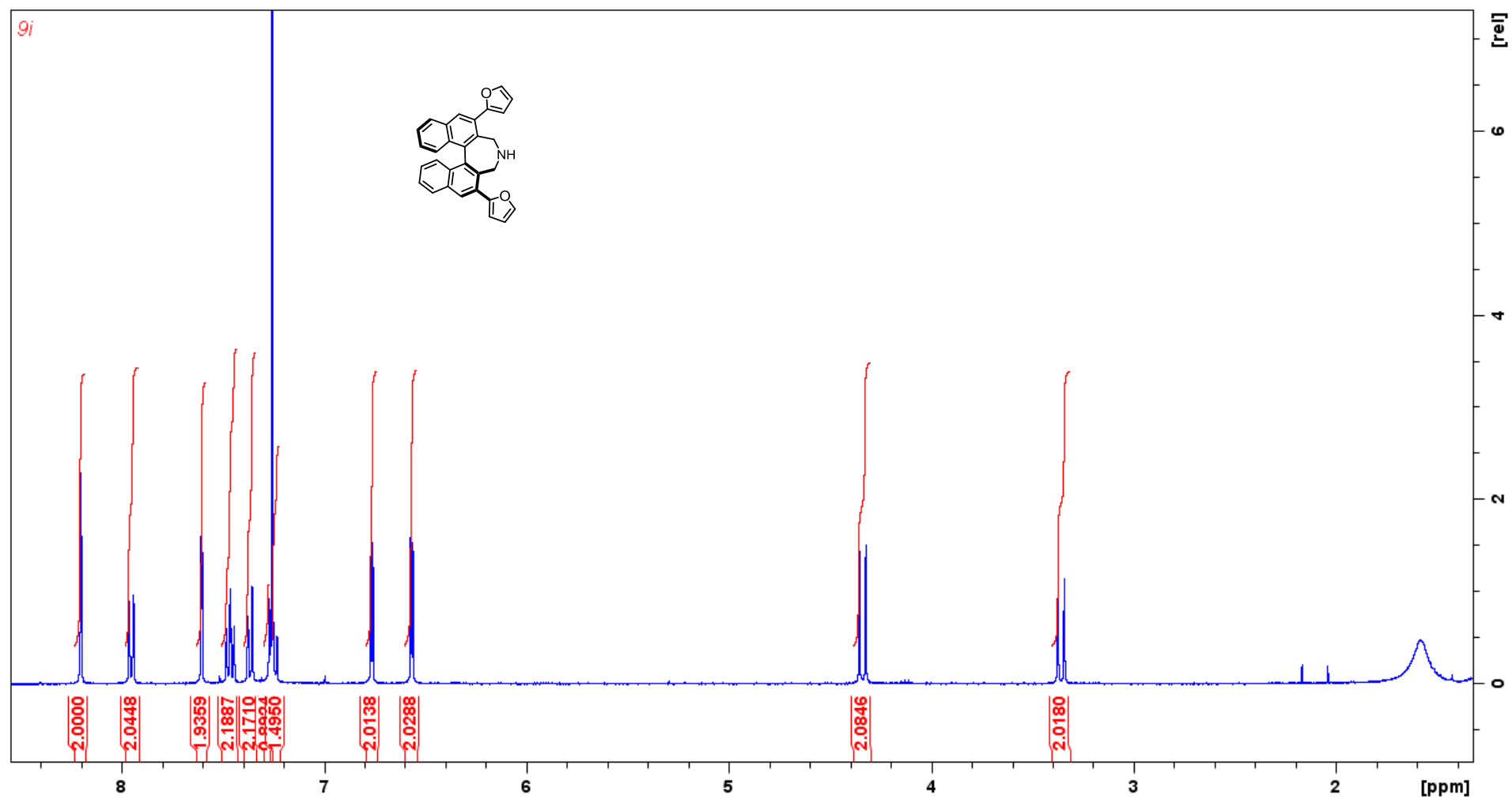




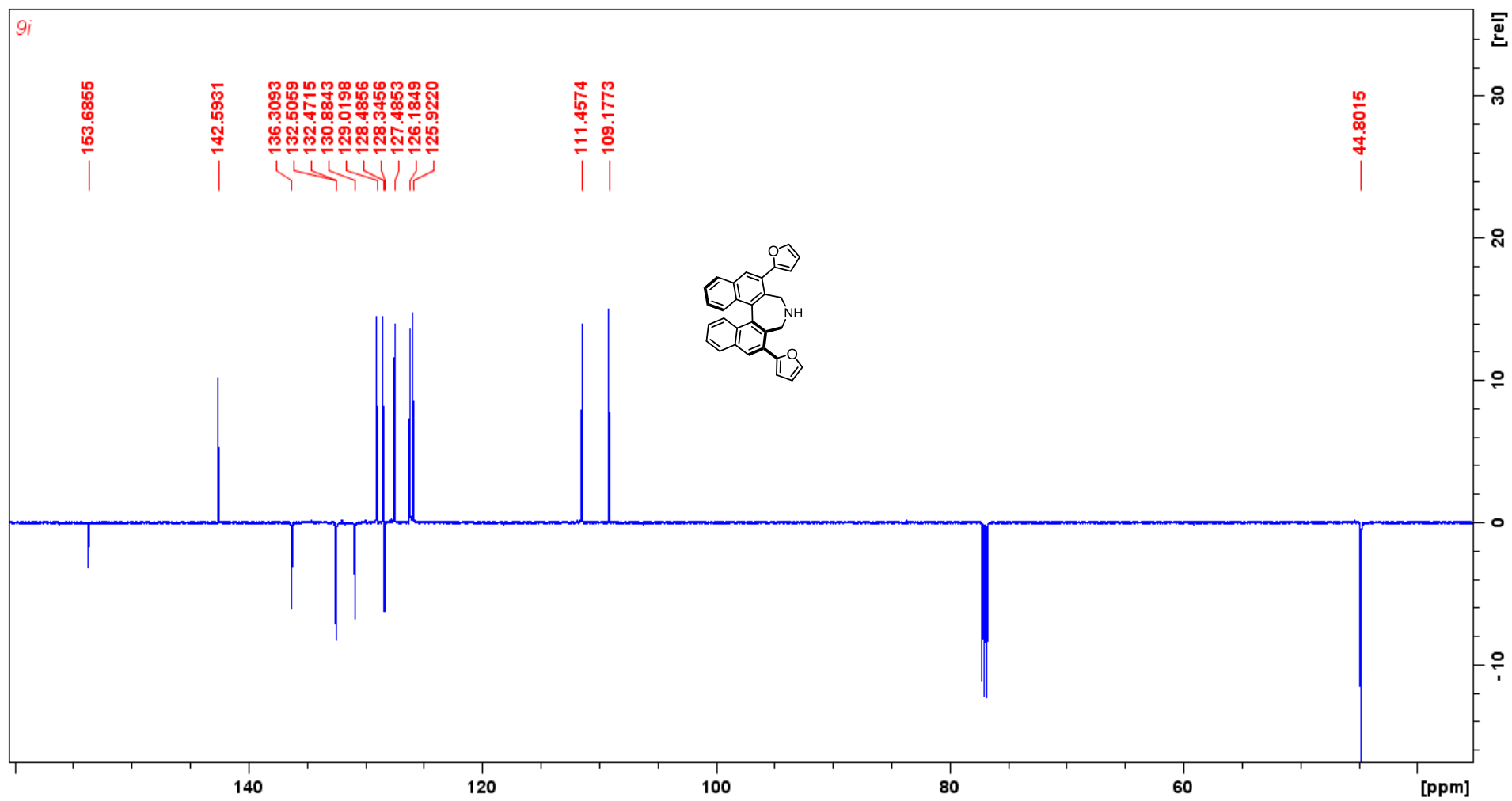


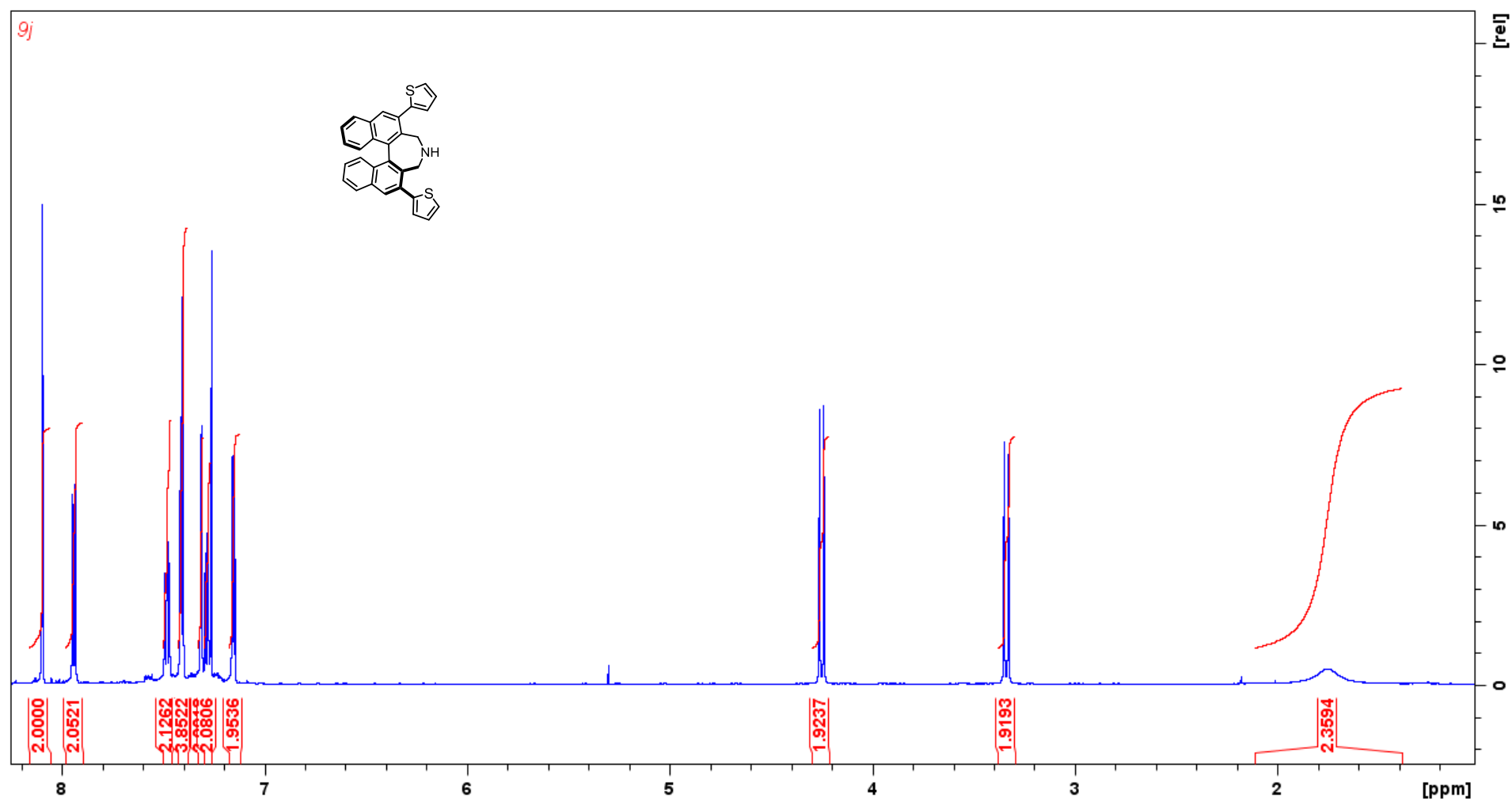


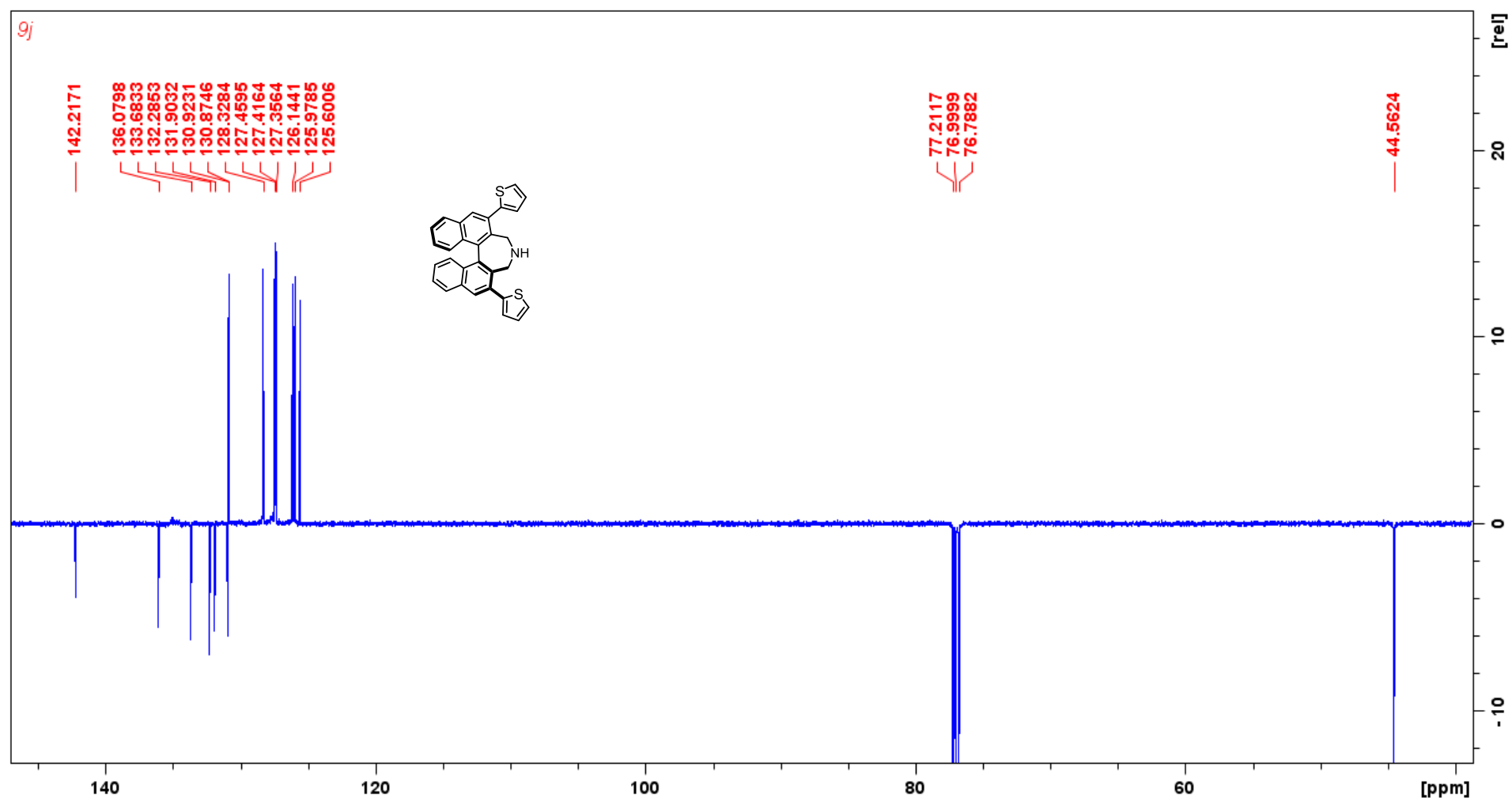


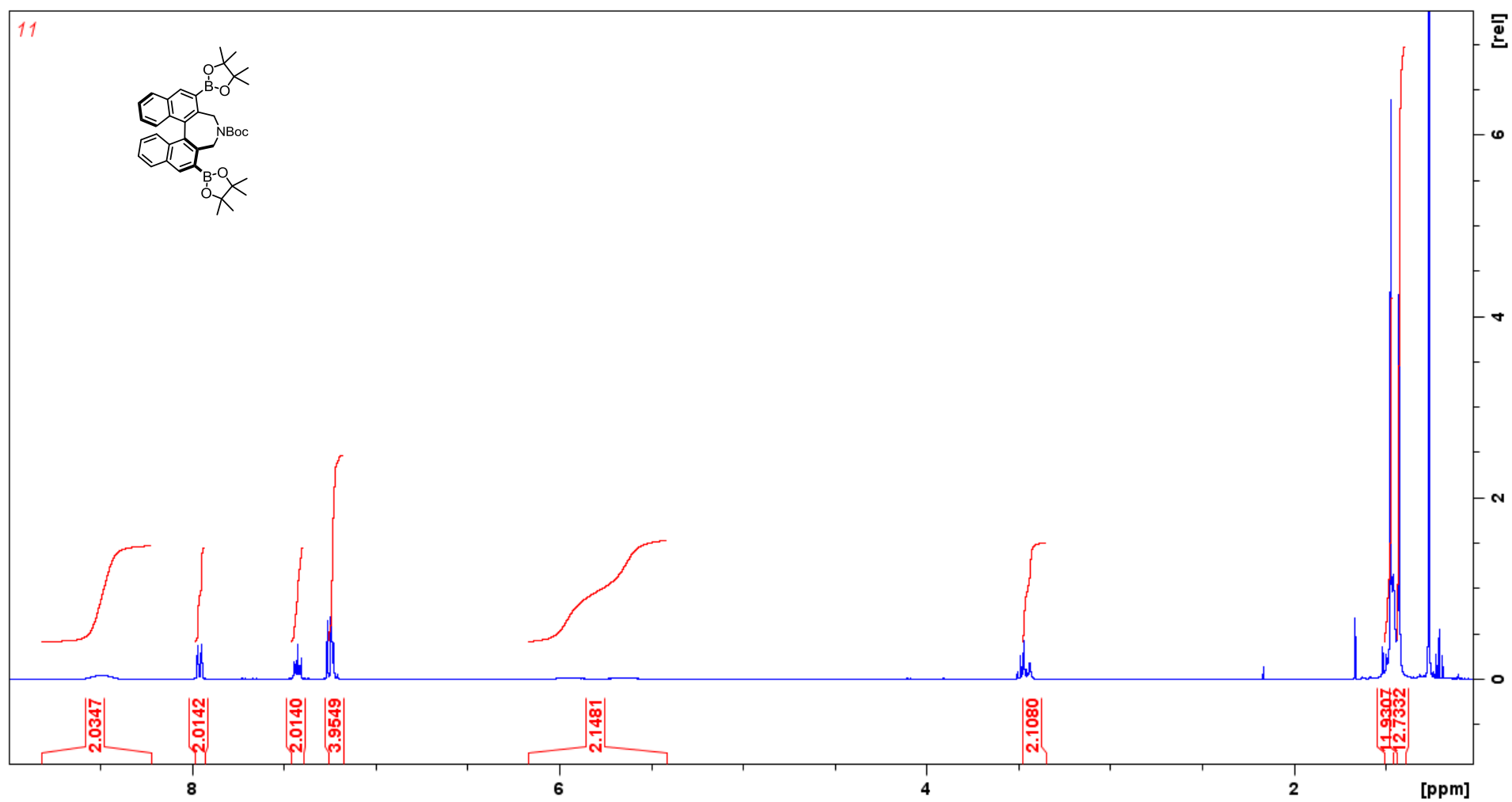


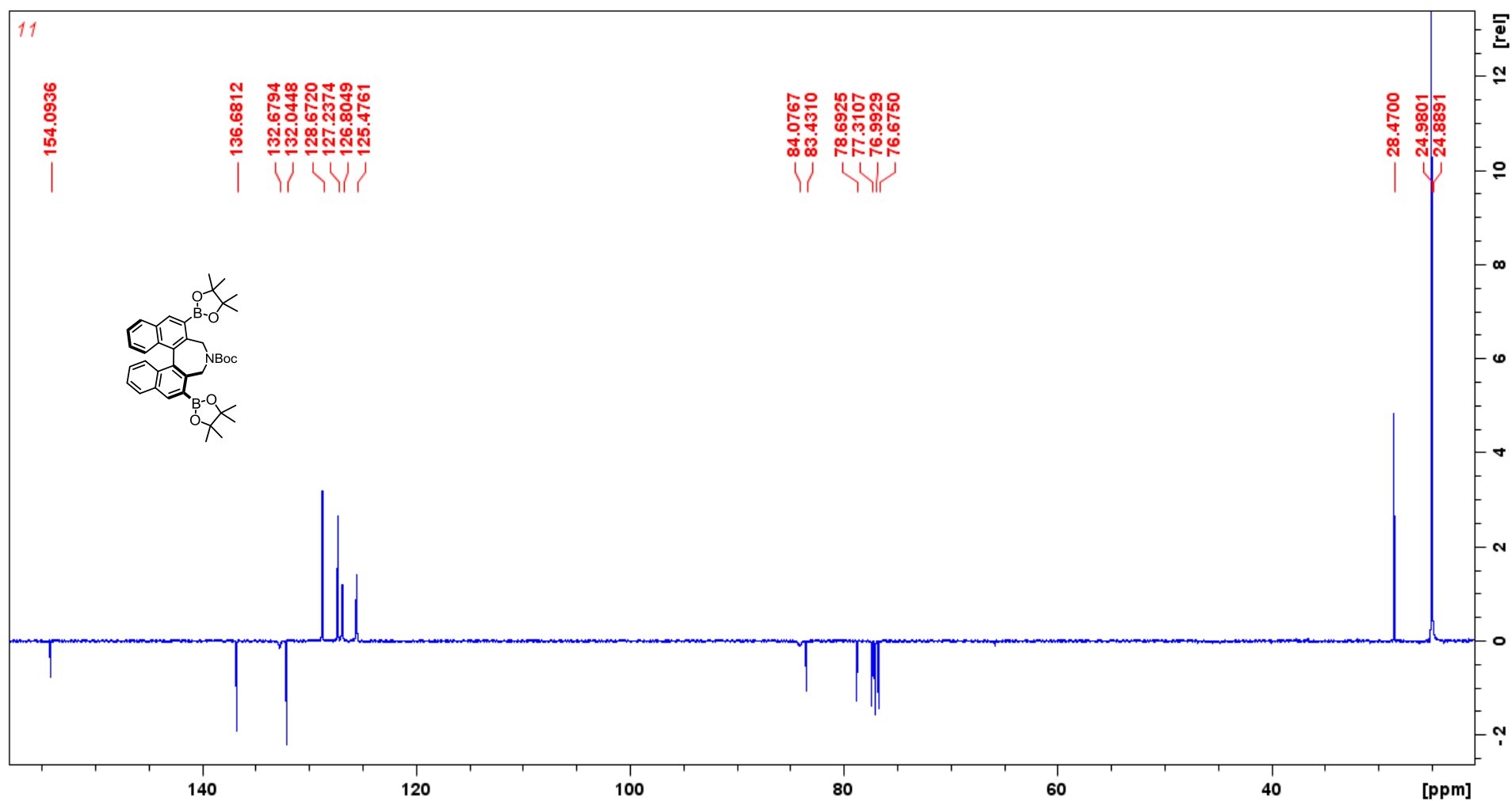


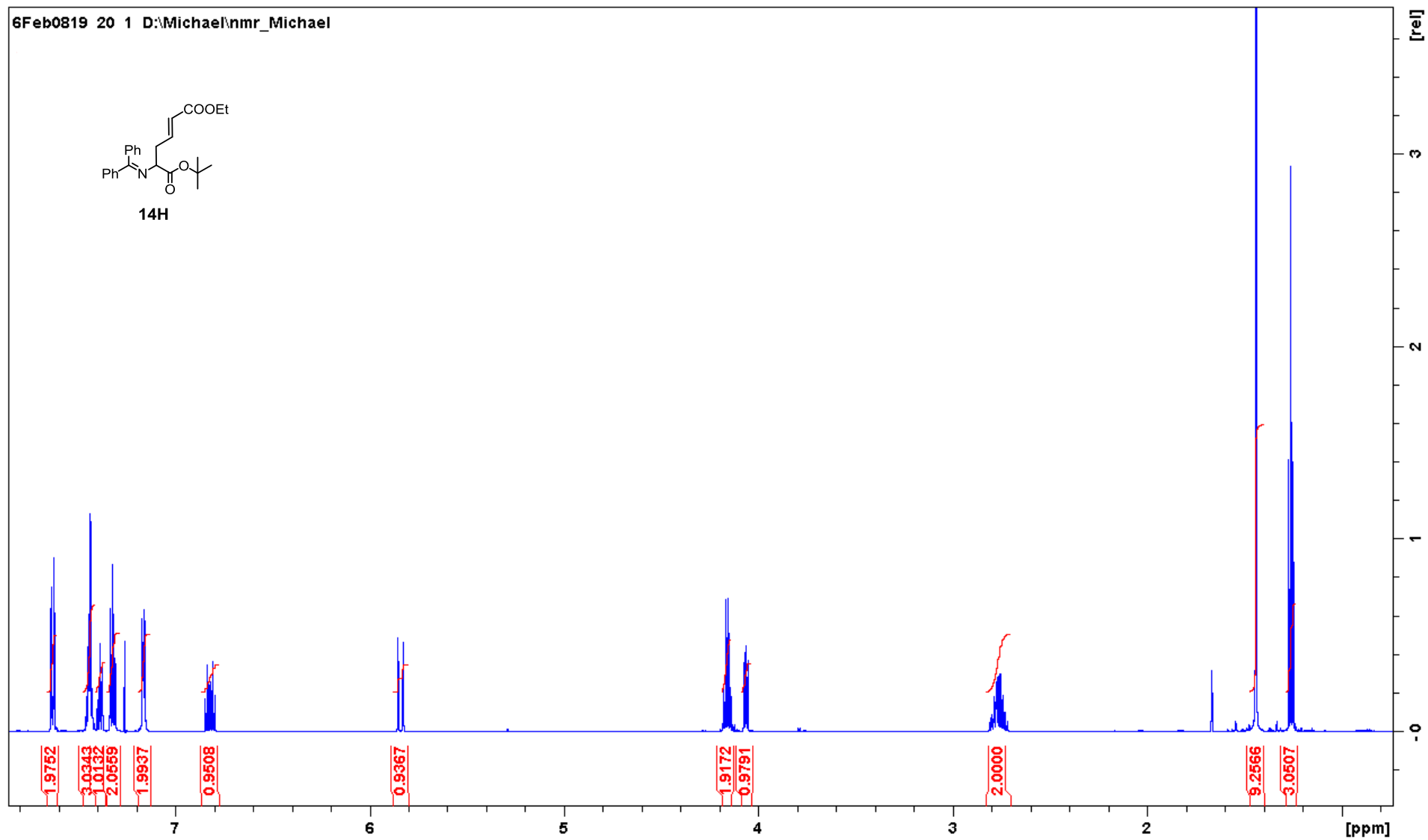


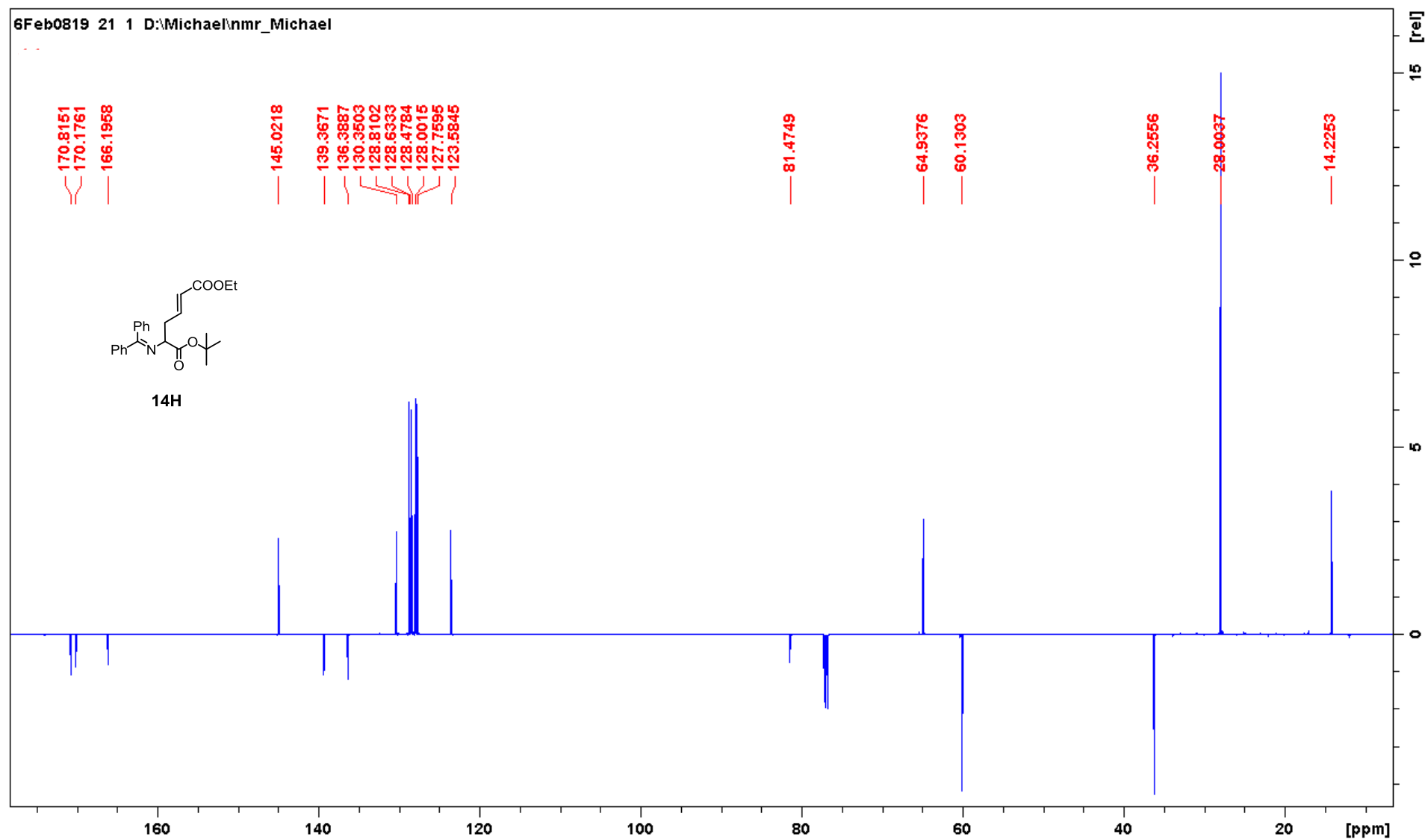




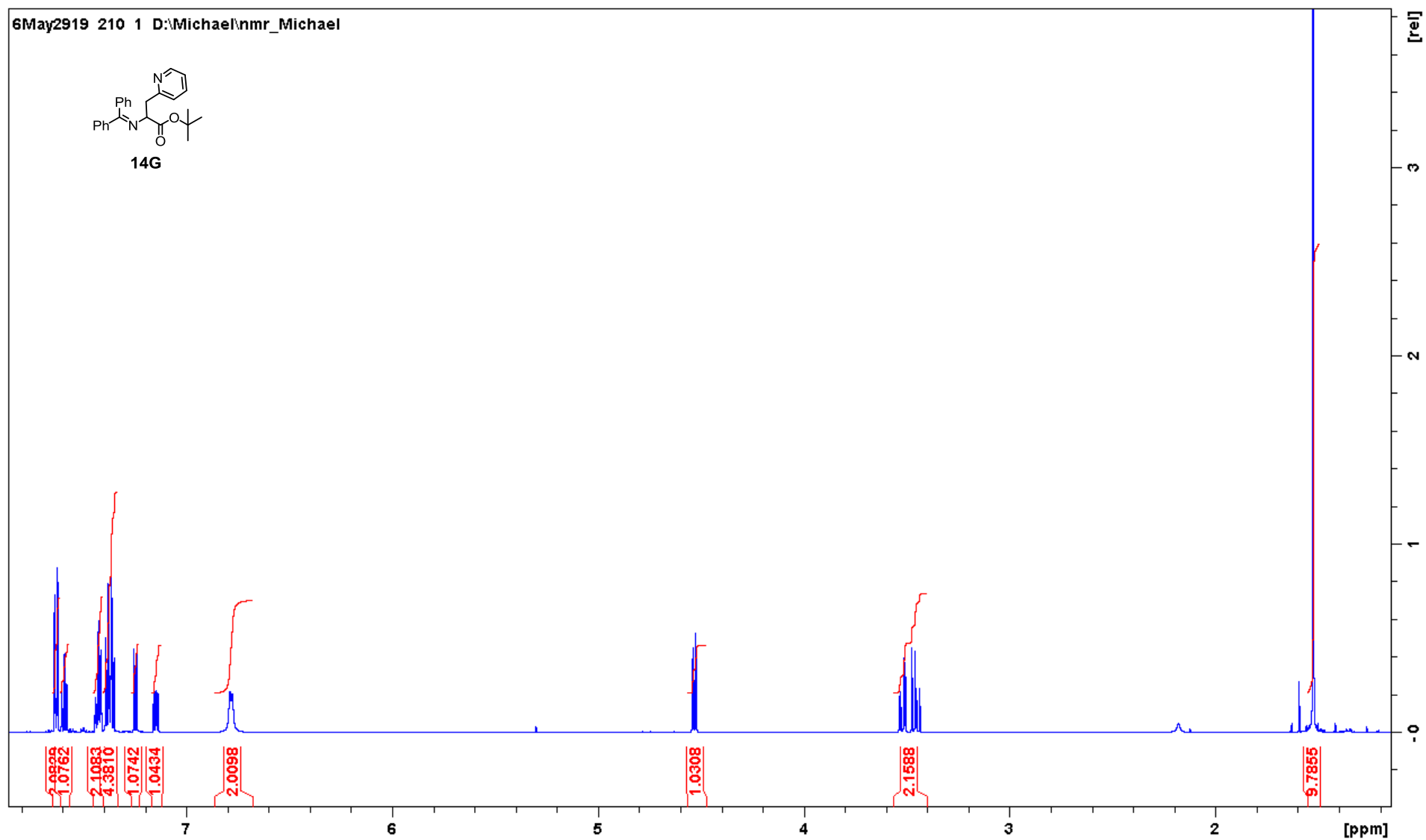
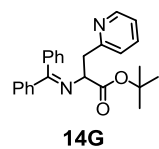




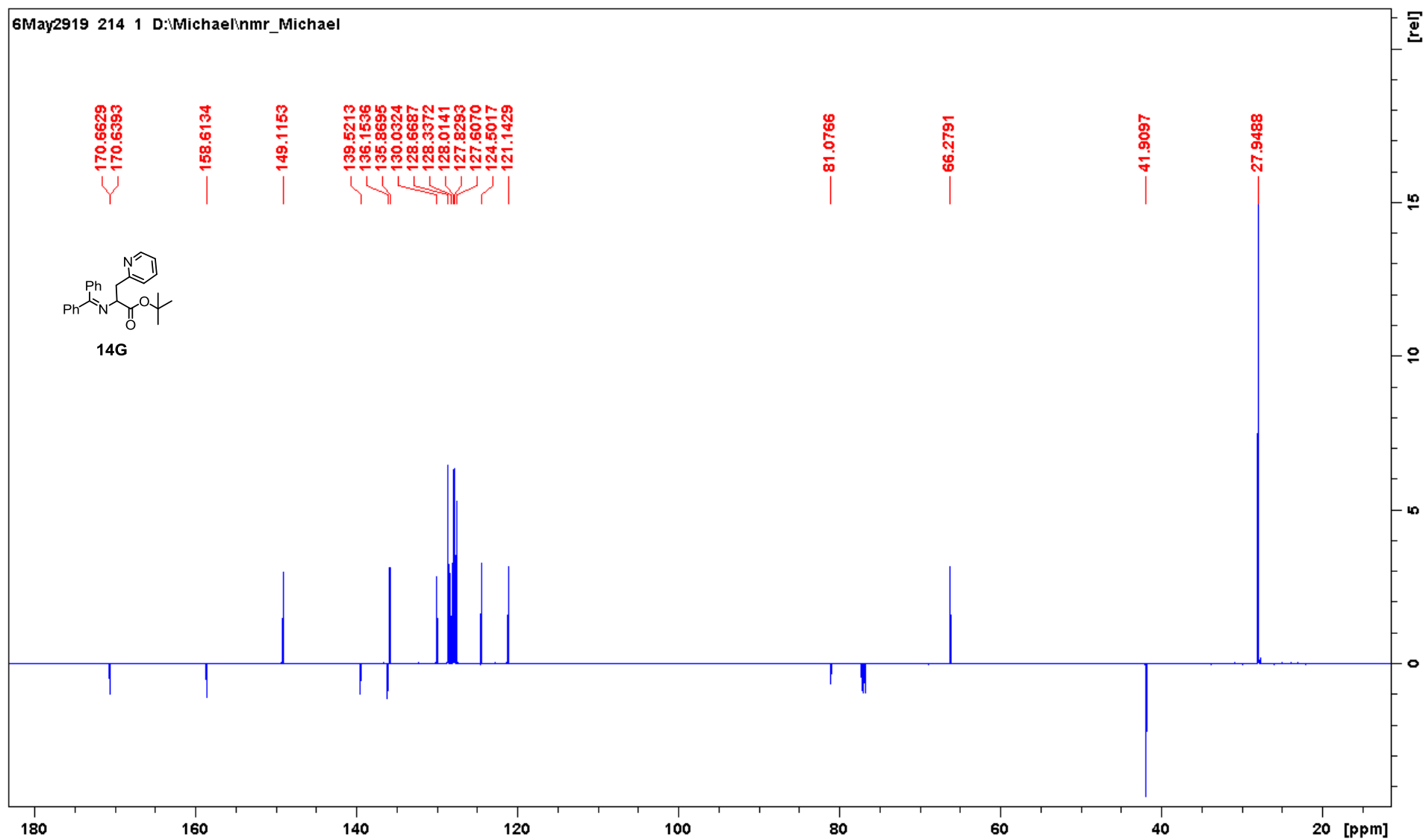




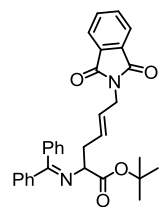
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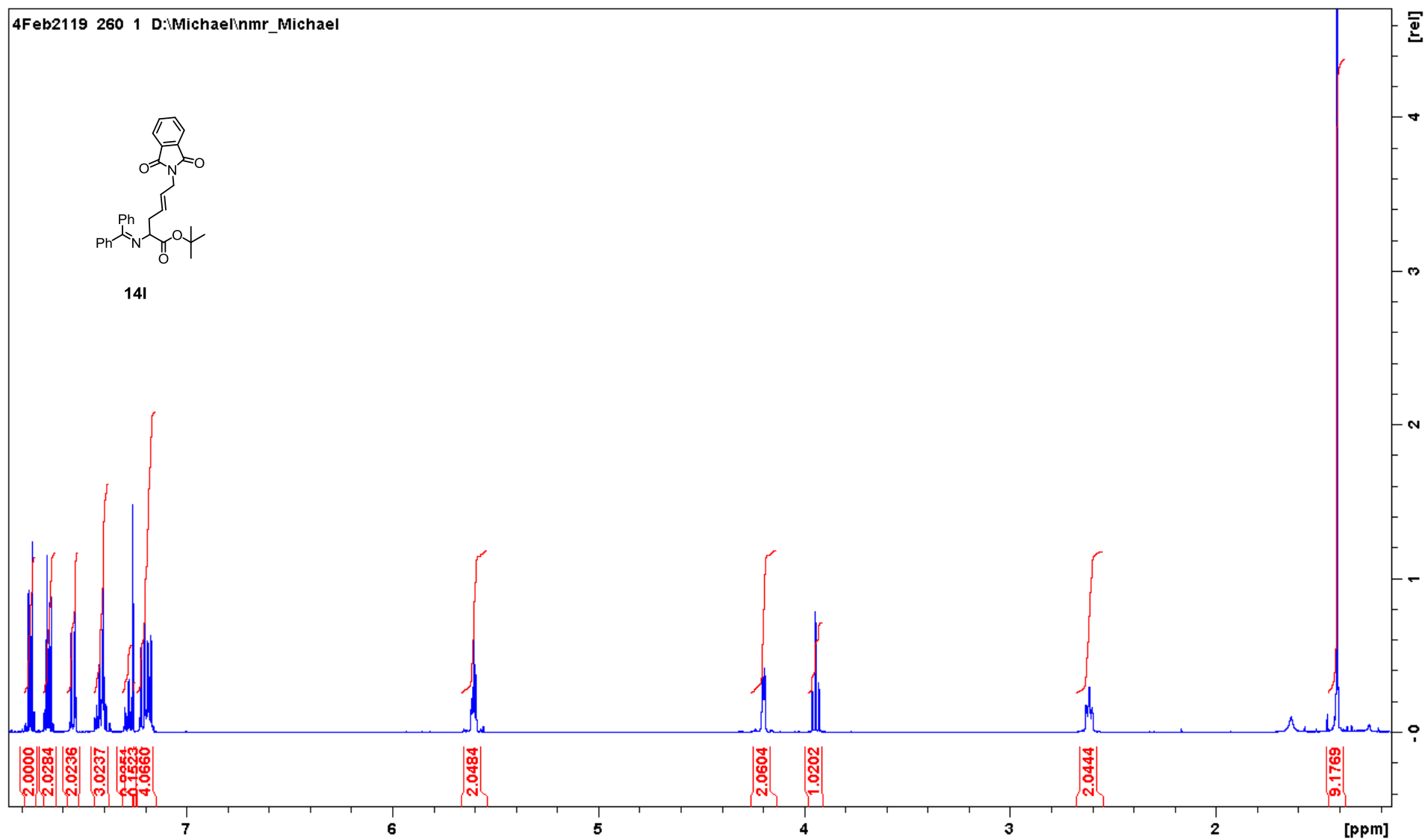


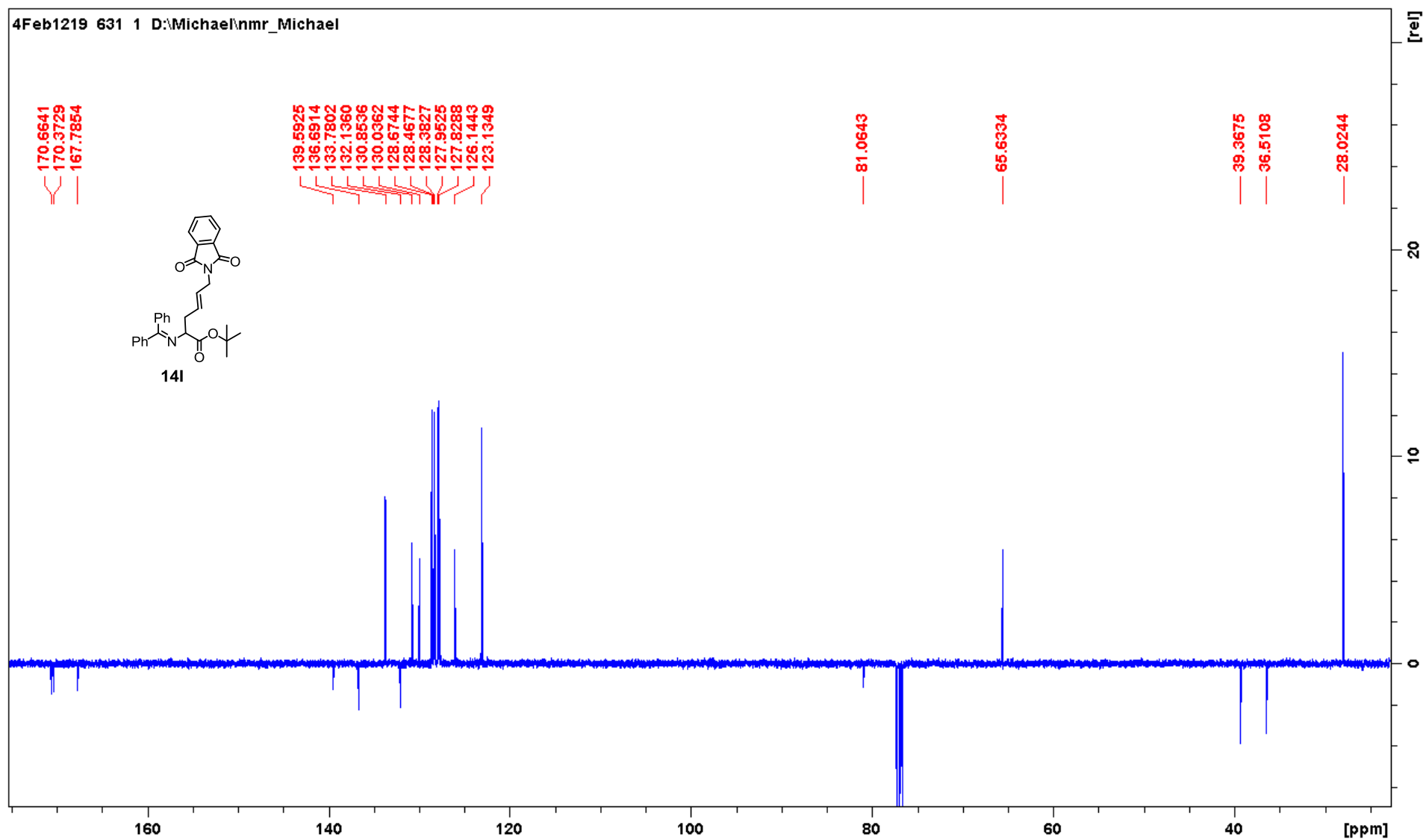


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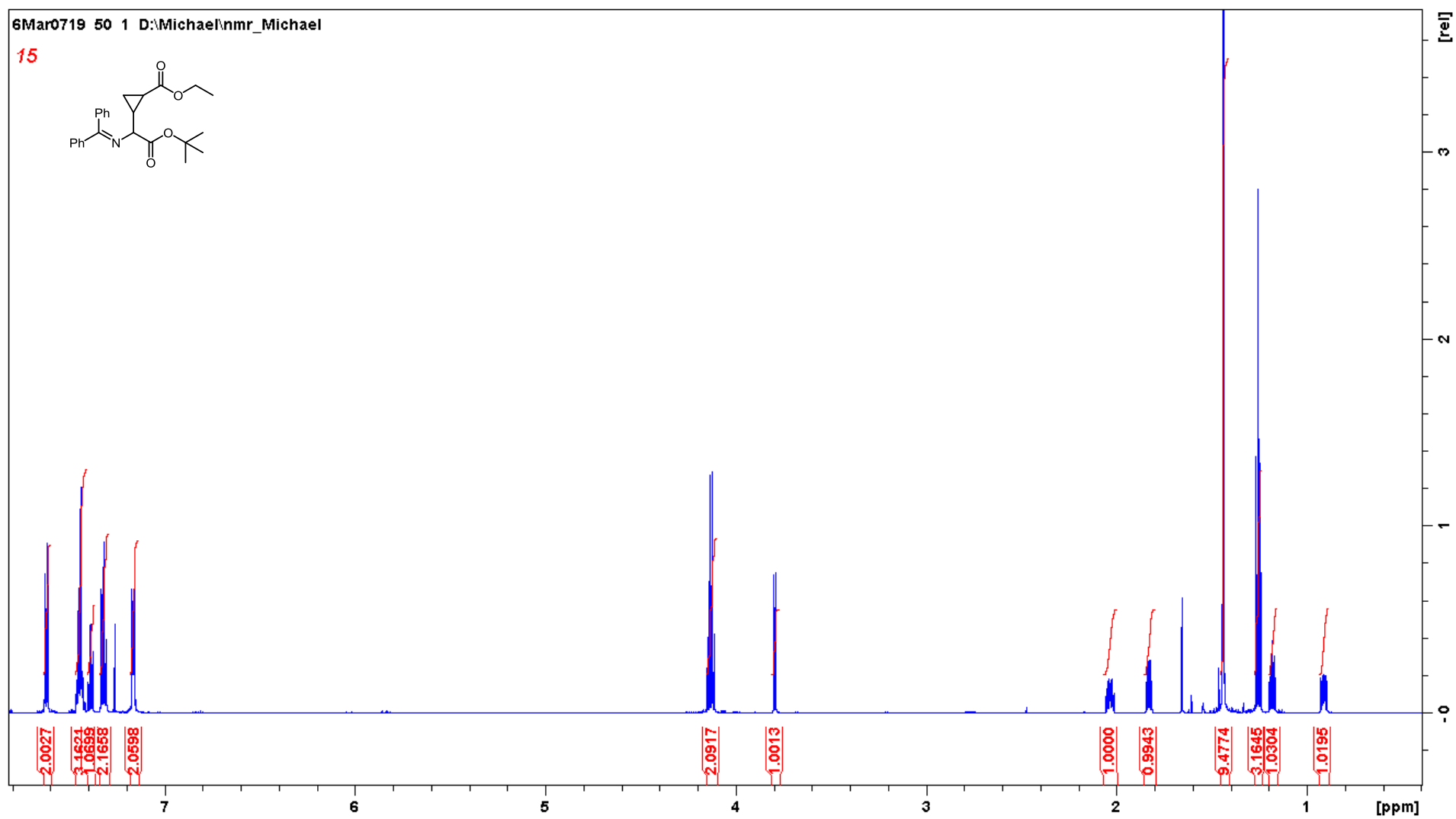
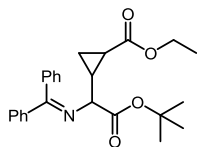
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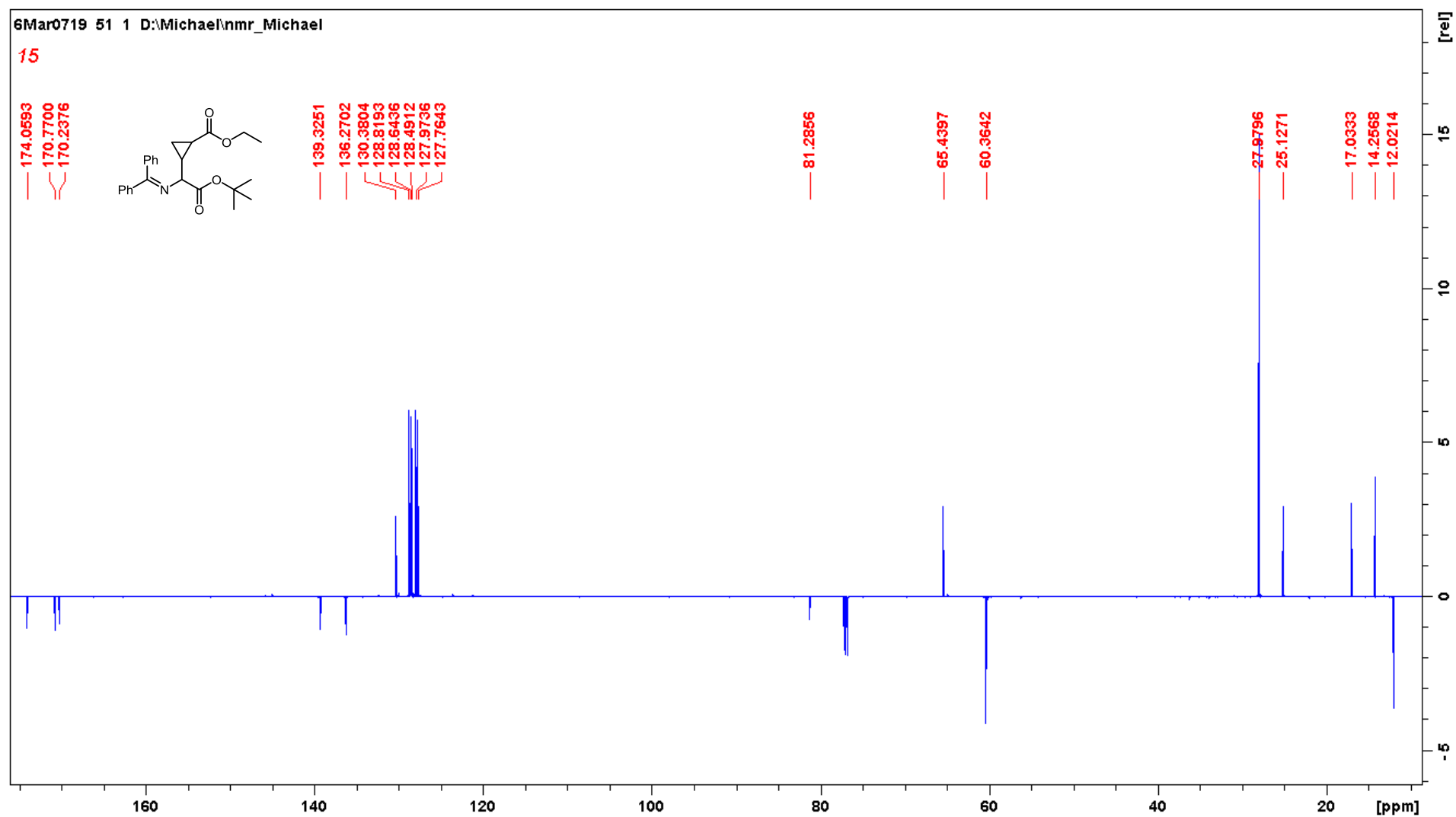




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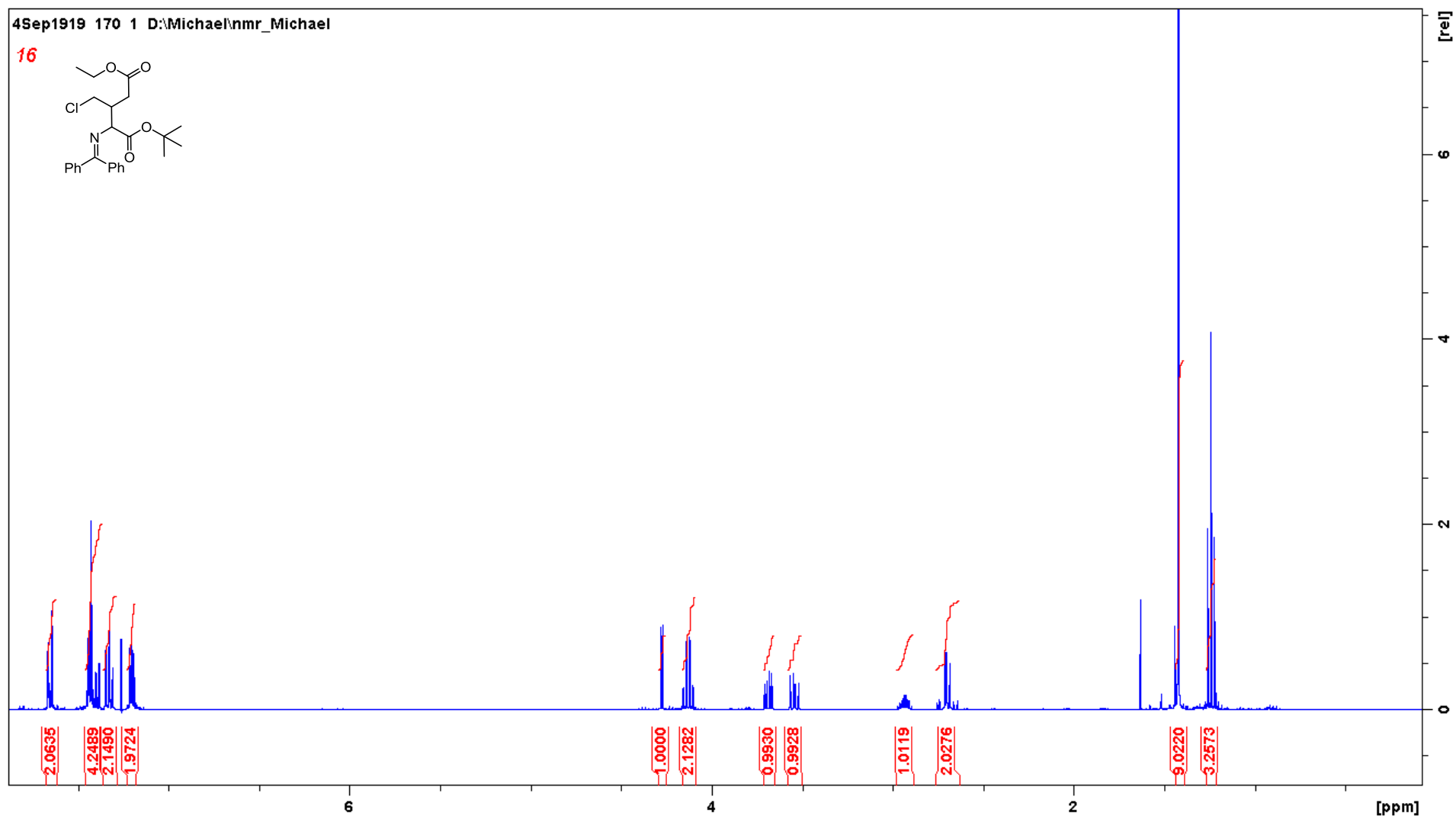
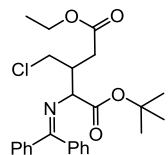
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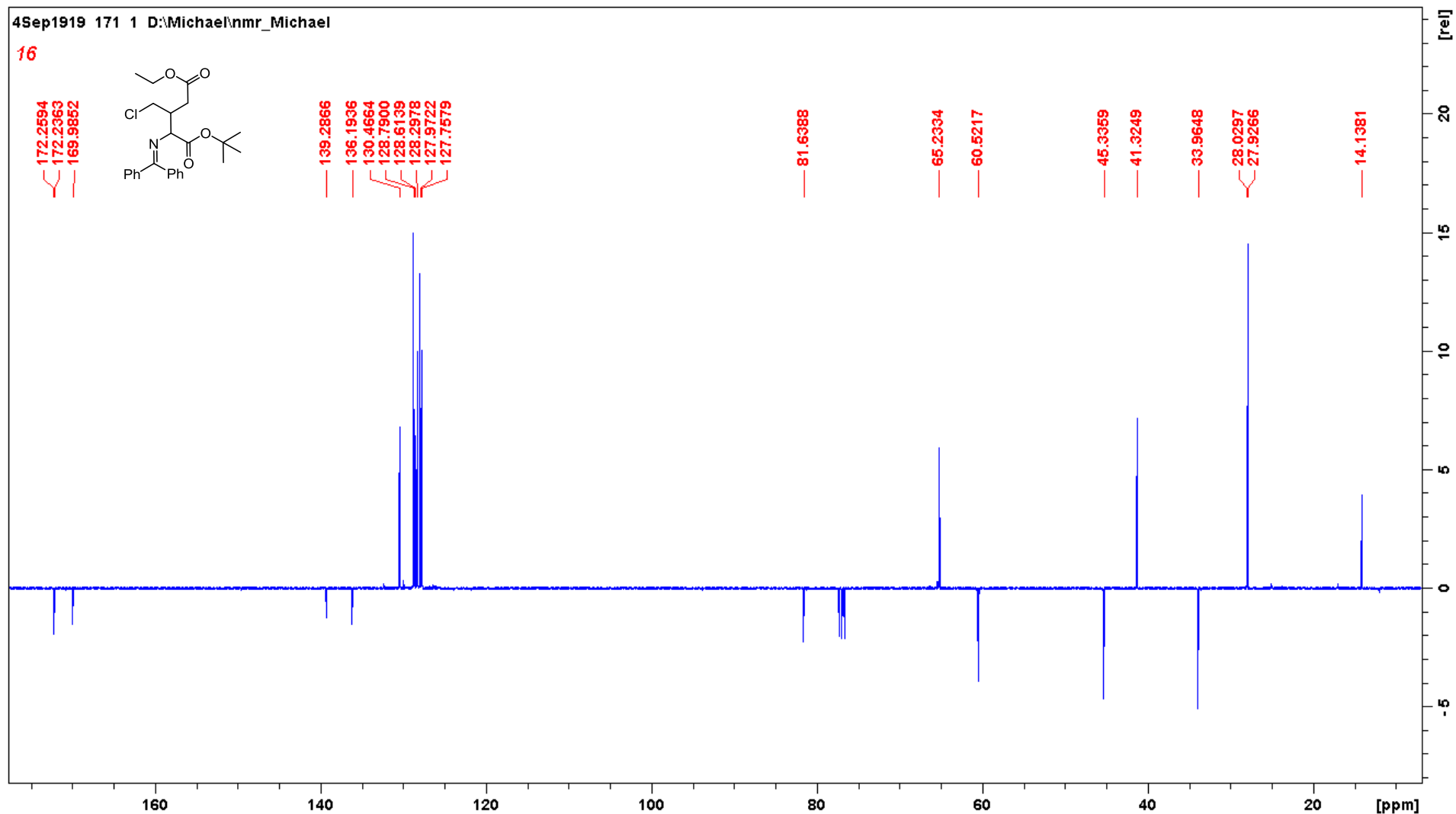


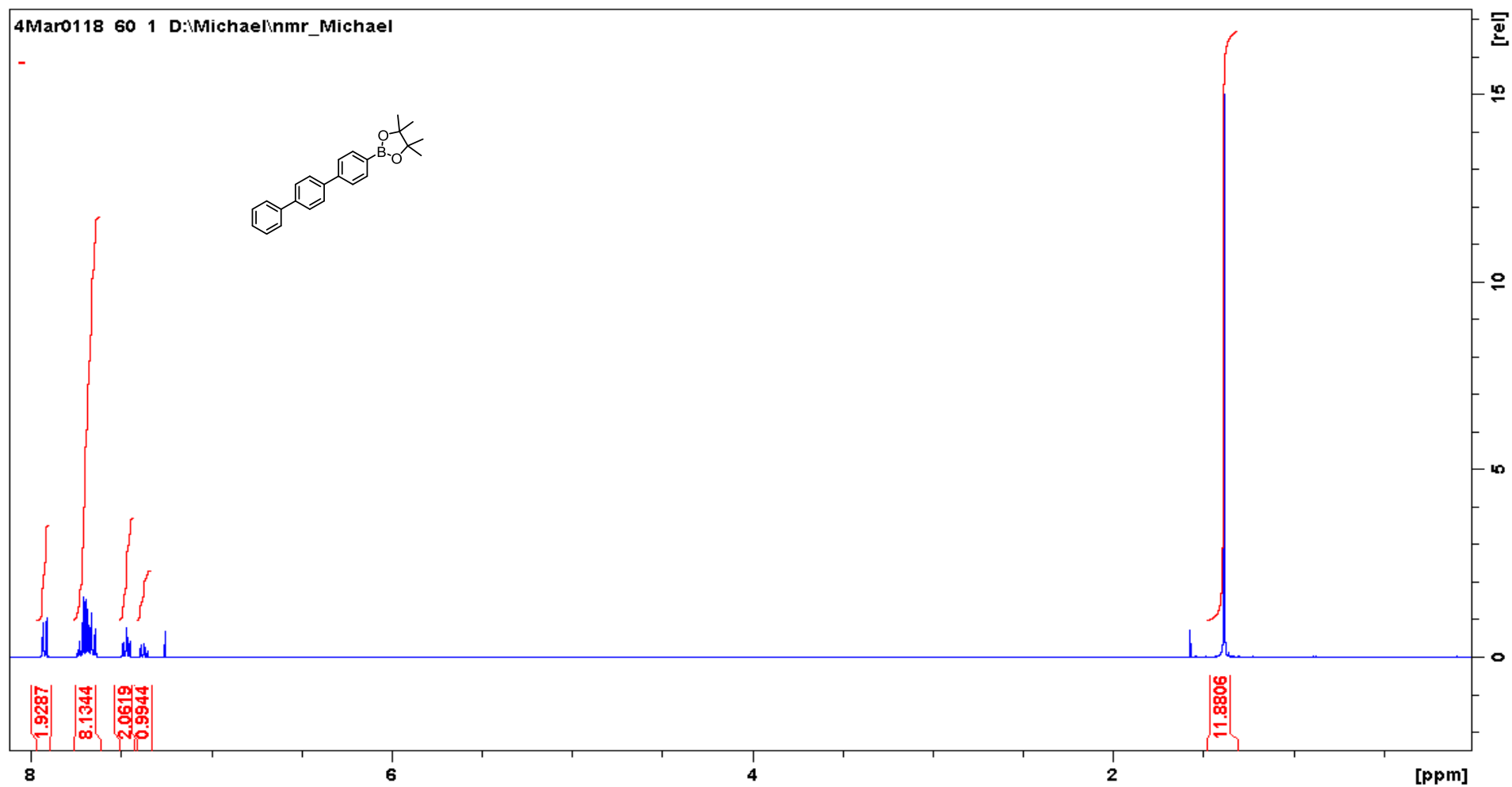


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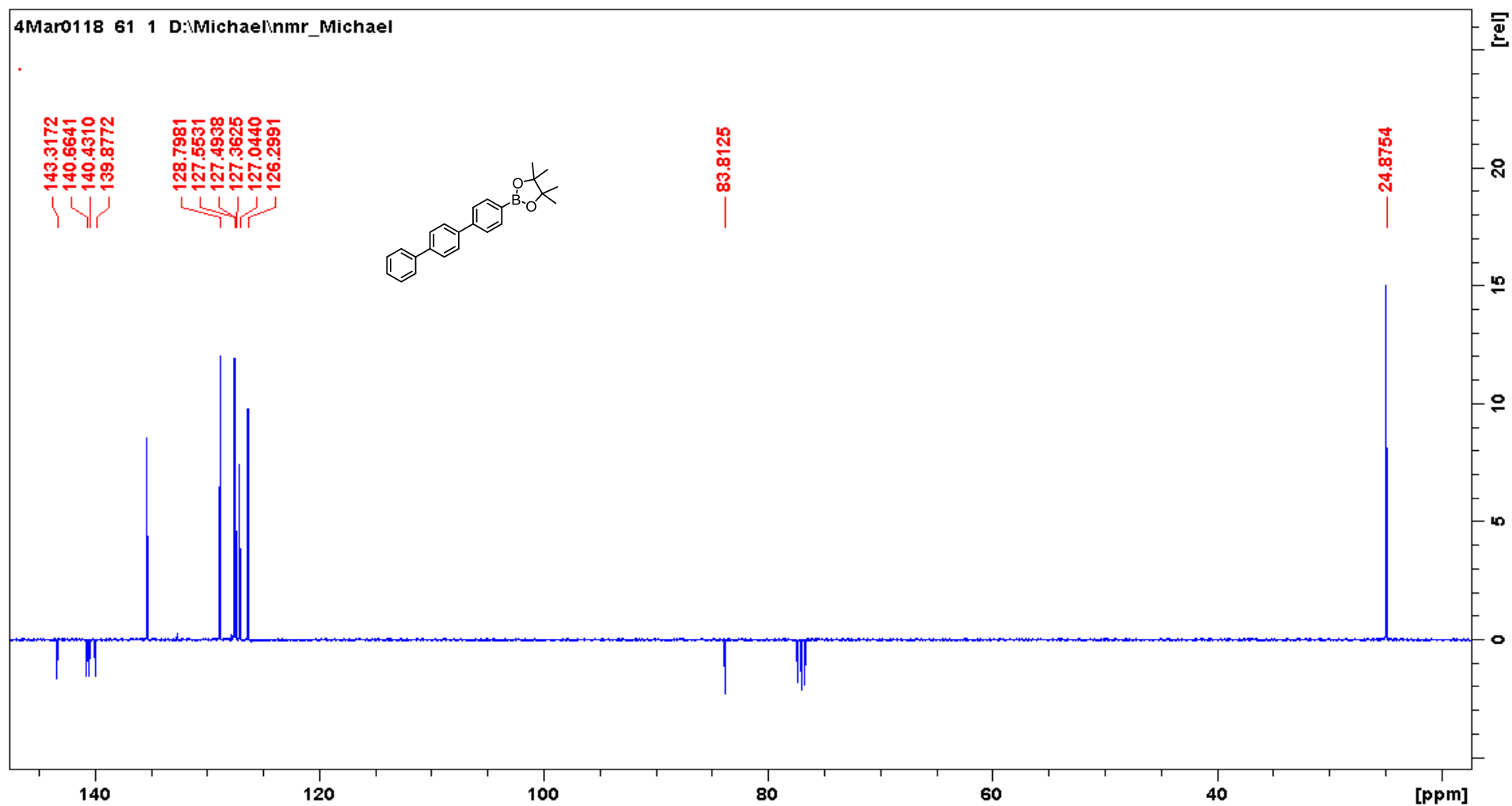
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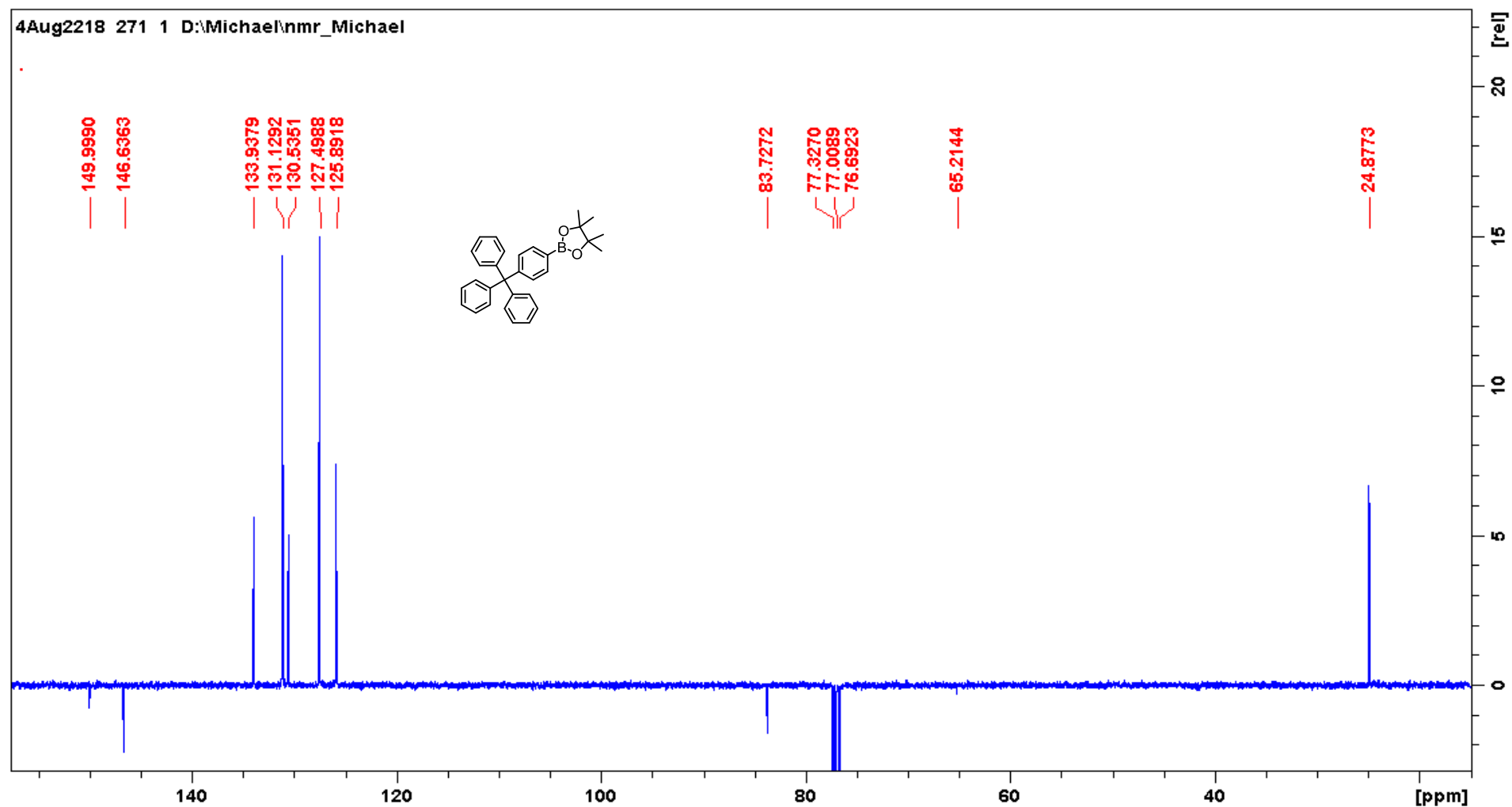


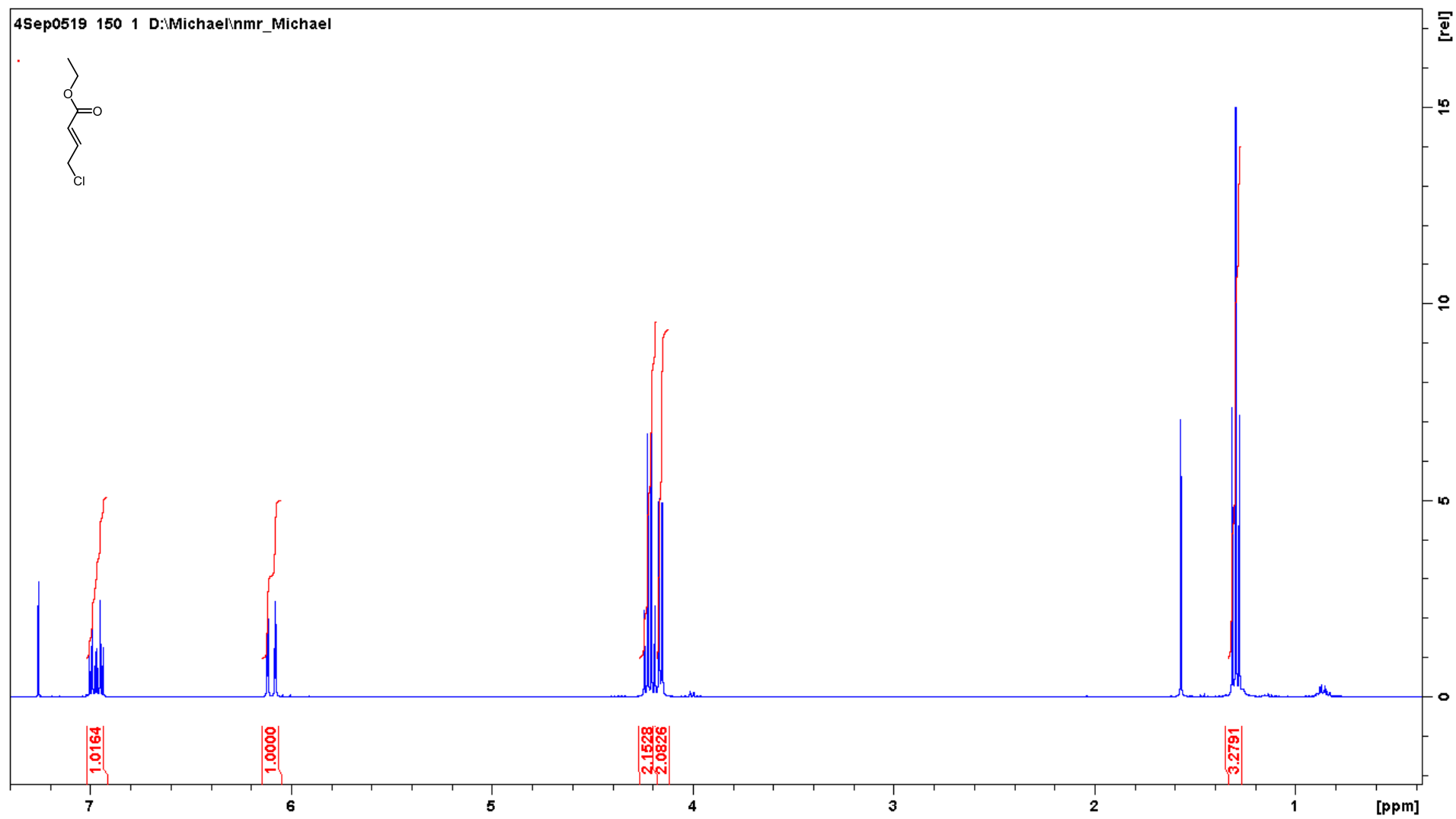


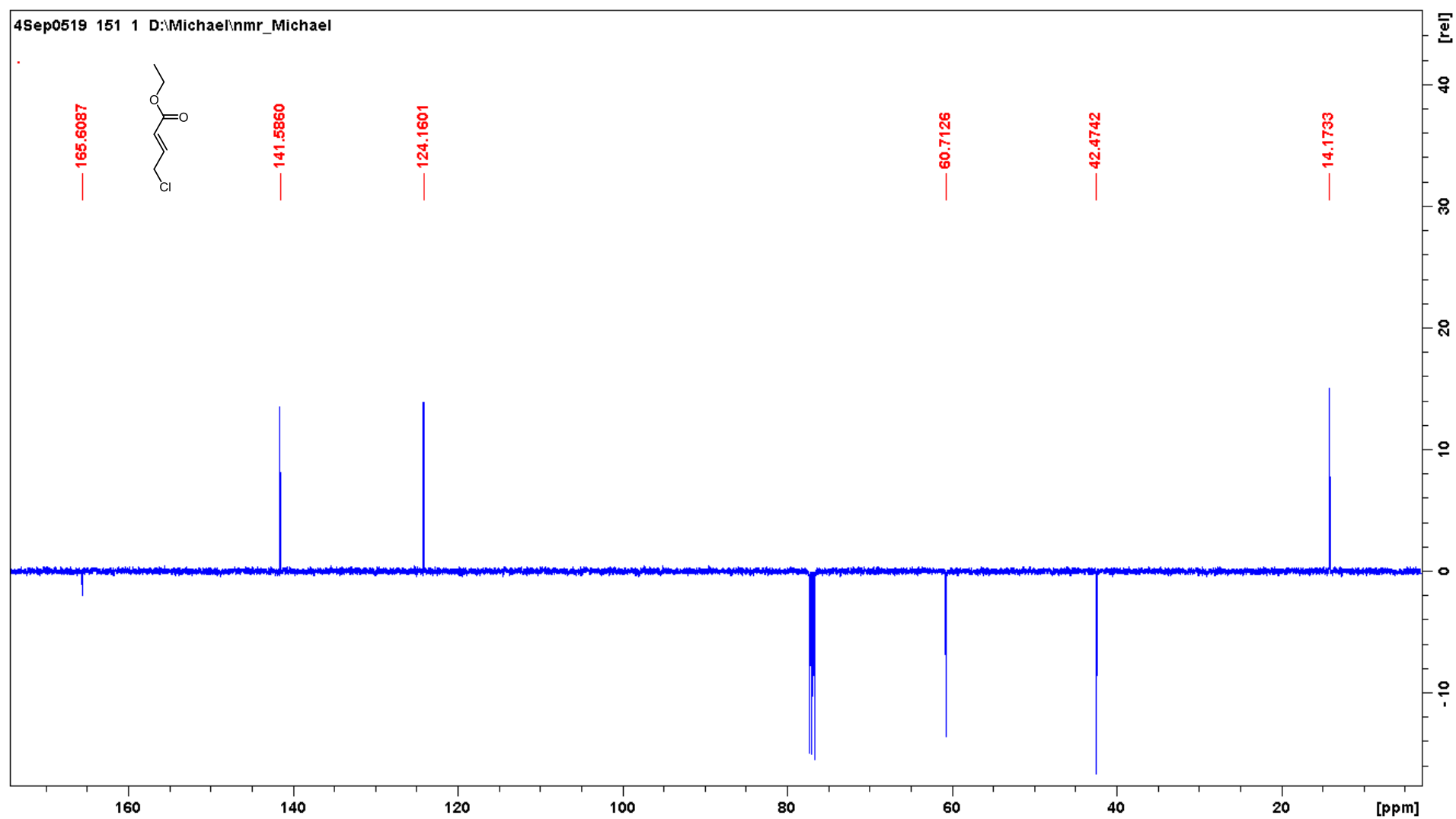


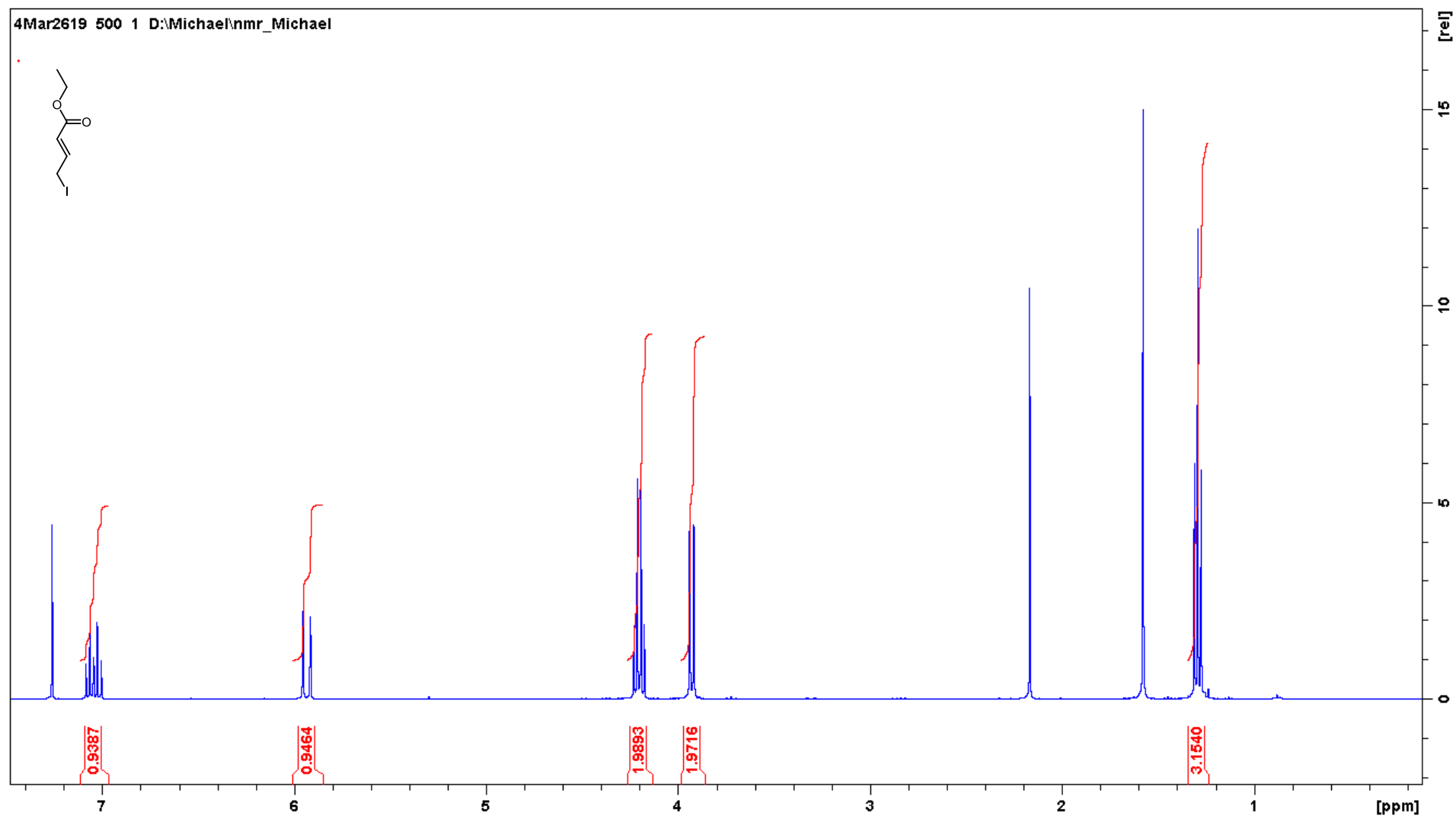


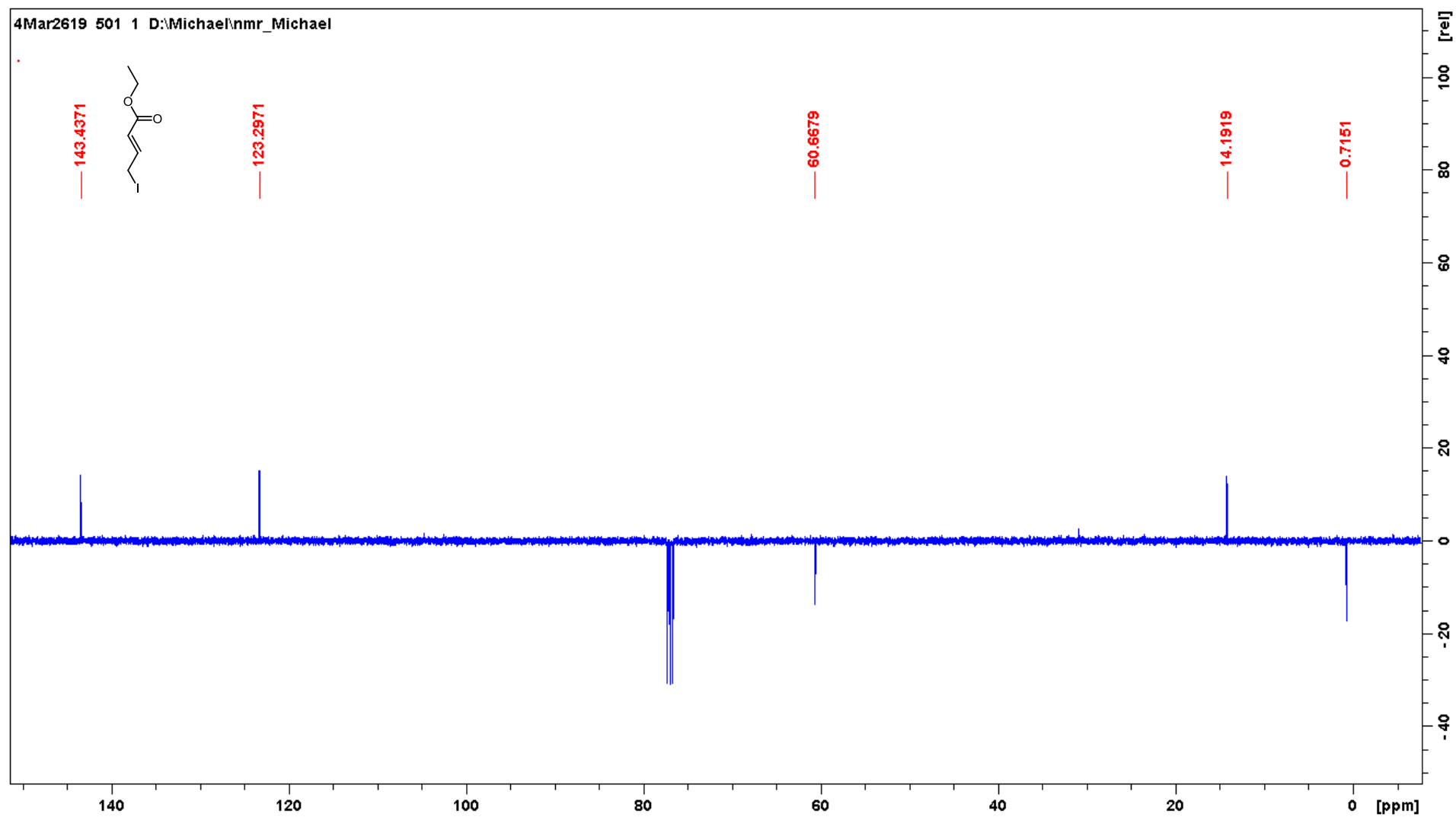


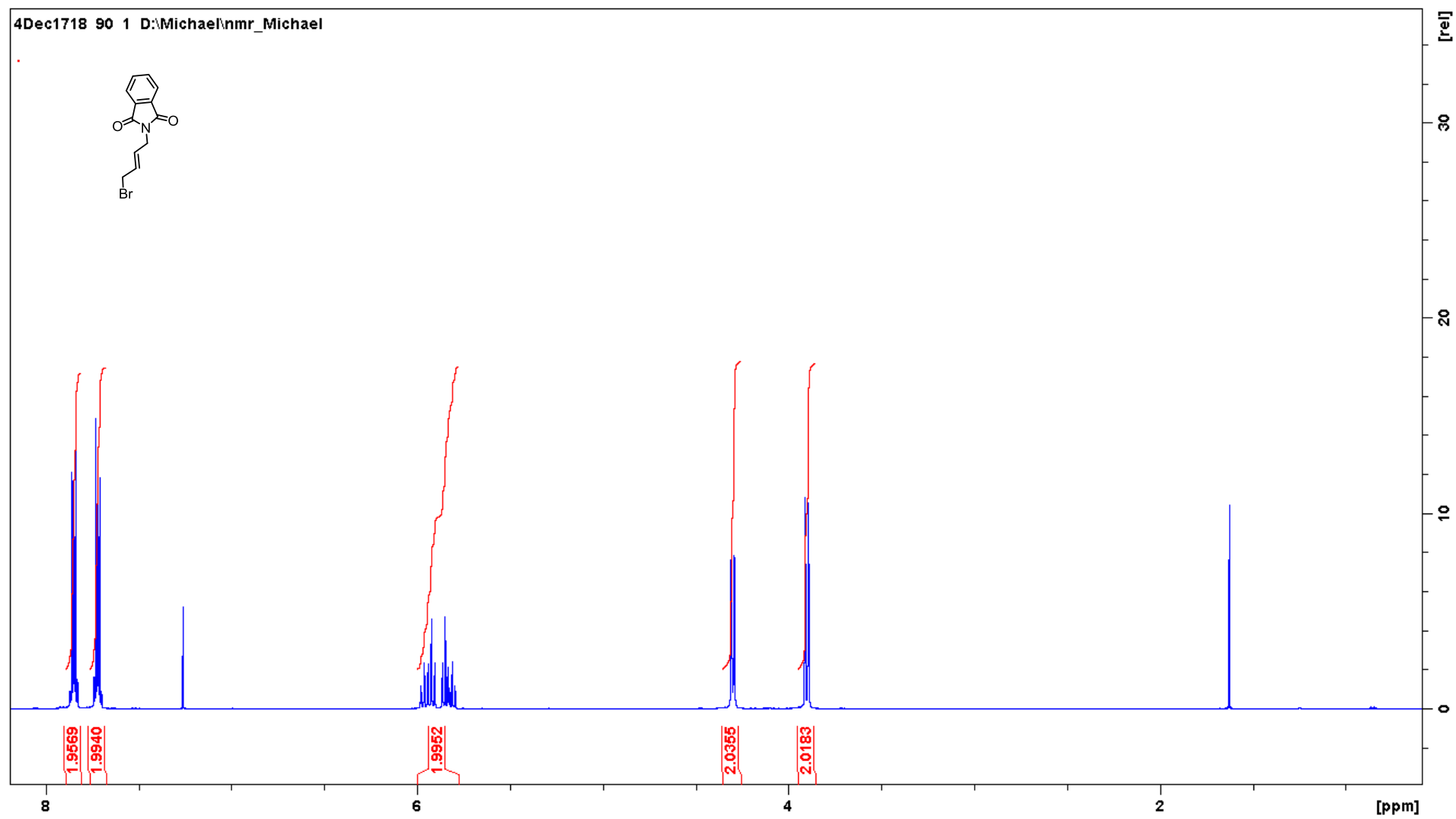




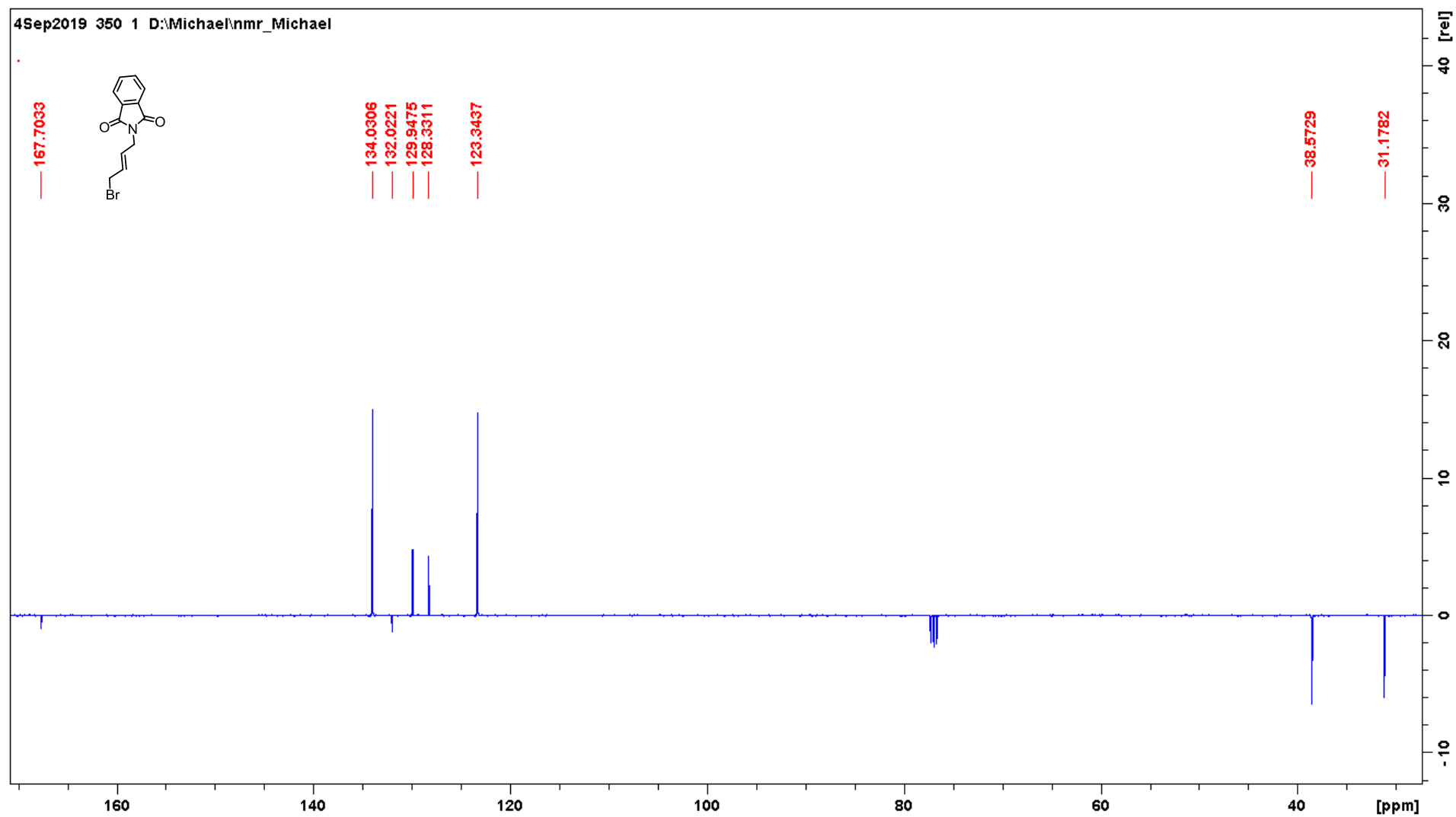


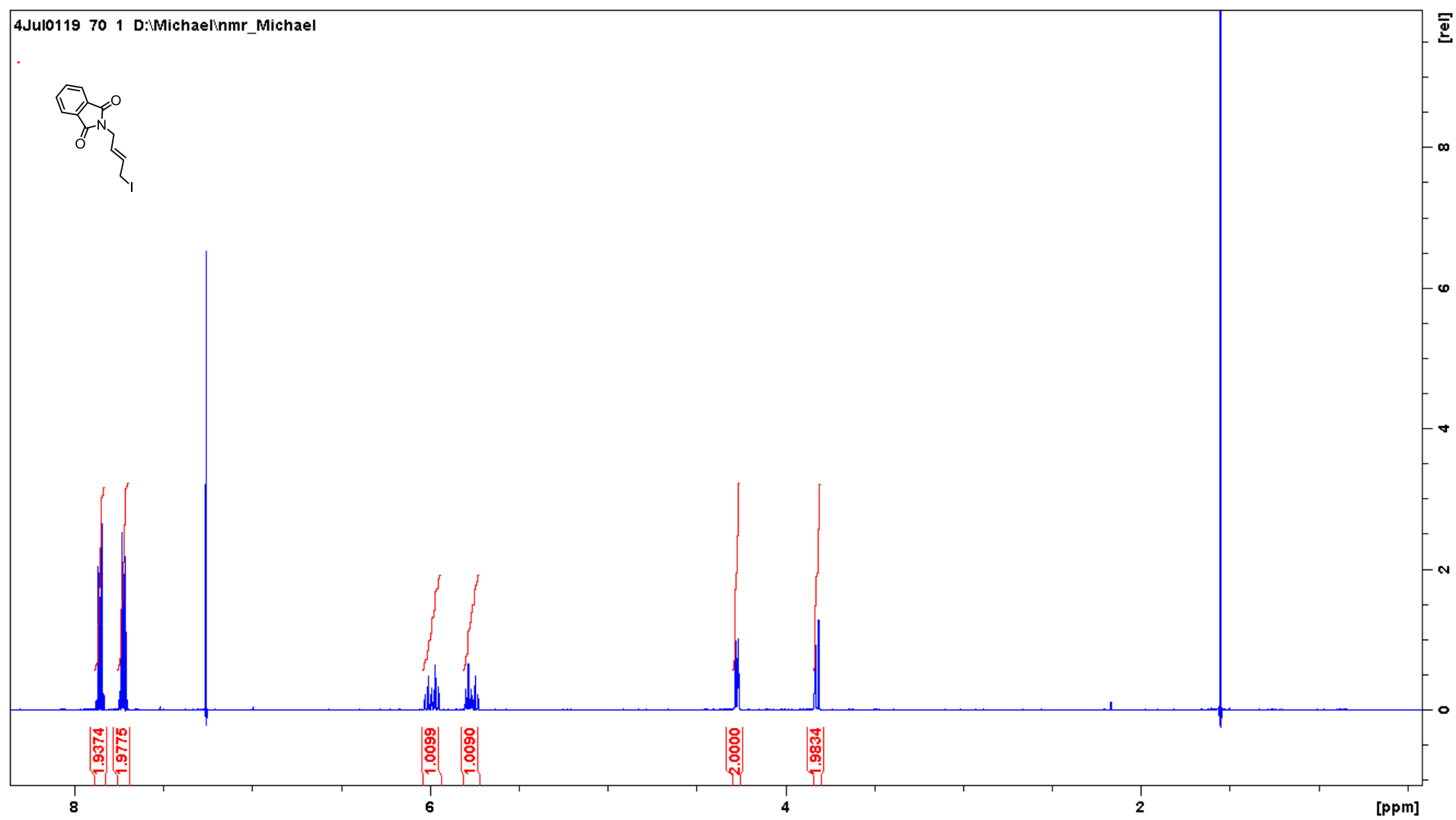


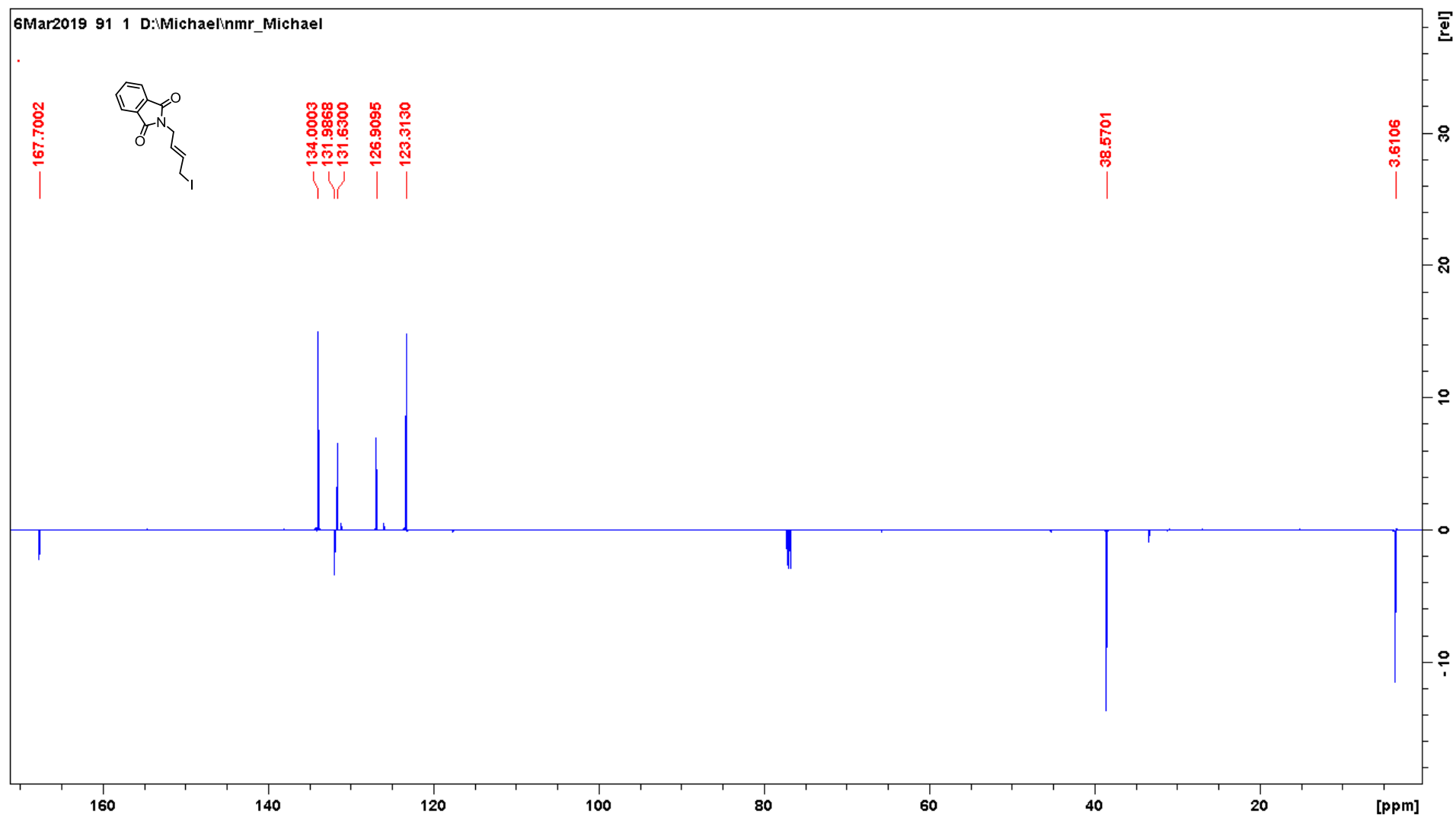












### Comments on screening experiments in PTC – Practical aspects

For screening experiments it is essential to establish a standardized, reproducible procedure with minimum expenditure of time and resources. Typical key data of interest are conversion, regioselectivity, expressed in relative yields of regioisomers, formation of side and sequence products and dia- and enantioselectivity. Depending on complexity of the reaction mixture a column chromatography is usually performed to obtain “isolated yields”. This step will give reproducible and reliable results if conducted on >100 mg scale but becomes increasingly error-prone on small scale. Moreover, this is the most time consuming part of the test reaction runs.

We therefore set out to develop a small scale screening procedure for our ligands, tested first in a prototypical reaction (**12** + **13A**, Scheme 2) which gives information on a) conversion (relativ to starting material), b) amount of product formed and isolated after extraction, and c) determination of enantiomeric excess, preferably without an extra step for purification or chromatographic separation. While a) can be often done by <sup>1</sup>H-NMR integration of proper, non-overlapping monitor protons, b) and c) might be more difficult to achieve.

We investigated 1,4-di(*tert*-butyl)benzene, 3,3'-dimethyl-1,1'-biphenyl and bibenzyl as internal standard (IS), added to the crude product mixture after extractive work-up, and recorded the <sup>1</sup>H-NMR spectrum in CDCl<sub>3</sub>. Best results were obtained with bibenzyl. Integration of the well separated singlet of methylene protons (4H) relative to dxd of the methine (1H) or methylene group (2H) of the product gave %-conversion, amount of product formed and eventually side products (if extracted in the organic phase). After evaporation an aliquot was subjected to chiral HPLC measurement (ca. 5 mg dissolved in 1 mL of 2-PrOH/heptane, 5 µL injected). For reaction (1) using reported conditions (Chiralcel ODH, 250 × 4.6 mm, 2-PrOH/heptane, 1:99, 0.5 mL min<sup>-1</sup>, 25 °C) well resolved peaks in the order of elution of internal standard, (*R*)-product, benzophenone, starting material, (*S*)-product were obtained.

#### Screening experiments (Details):

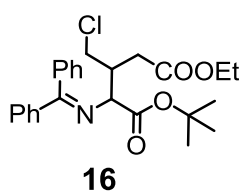
**Table S1.** Asymmetric substitution of *tert*-butyl 2-((diphenylmethylene)amino)acetate (**12**) with electrophiles **13B-13I** under PT conditions.<sup>a</sup>

entry	cat.	t/h	base	product	yield/% <sup>b</sup>	ee/% <sup>c</sup>	note
1	<b>1c</b>	20	KOH	<b>14B</b>	89	88	
2	<b>1d</b>	20	KOH	<b>14B</b>	<b>90</b>	88	
3	<b>1g</b>	20	KOH	<b>14B</b>	89	<b>98</b>	d
4	<b>1h</b>	20	KOH	<b>14B</b>	70	<b>98</b>	
5	<b>1c</b>	20	KOH	<b>14C</b>	80	91	
6	<b>1d</b>	20	KOH	<b>14C</b>	<b>86</b>	91	
7	<b>1g</b>	20	KOH	<b>14C</b>	71	<b>95</b>	e
8	<b>1h</b>	20	KOH	<b>14C</b>	45	52	
9	<b>1c</b>	20	KOH	<b>14D</b>	<b>90</b>	75	
10	<b>1d</b>	20	KOH	<b>14D</b>	<b>90</b>	74	
11	<b>1g</b>	20	KOH	<b>14D</b>	87	<b>93</b>	
12	<b>1h</b>	20	KOH	<b>14D</b>	61	84	

entry	cat.	t/h	base	product	yield/% <sup>b</sup>	ee/% <sup>c</sup>	note
13	<b>1c</b>	20	KOH	<b>14E</b>	51	83	
14	<b>1d</b>	20	KOH	<b>14E</b>	55	<b>88</b>	
15	<b>1g</b>	20	KOH	<b>14E</b>	<b>72</b>	68	
16	<b>1h</b>	20	KOH	<b>14E</b>	66	14	
17	<b>1c</b>	20	KOH	<b>14F</b>	<b>92</b>	89	
18	<b>1d</b>	20	KOH	<b>14F</b>	91	88	
19	<b>1g</b>	20	KOH	<b>14F</b>	90	<b>90</b>	f
20	<b>1h</b>	20	KOH	<b>14F</b>	30	90	
21	<b>1c</b>	20	KOH	<b>14G</b>	88	72	
22	<b>1d</b>	20	KOH	<b>14G</b>	<b>91</b>	71	
23	<b>1g</b>	20	KOH	<b>14G</b>	81	<b>83</b>	
24	<b>1h</b>	20	KOH	<b>14G</b>	83	73	
25	<b>1c</b>	20	CsOH	<b>14H</b>	62	51	g
26	<b>1d</b>	20	CsOH	<b>14H</b>	56	41	h
27	<b>1g</b>	4	KOH	<b>14H</b>	13	69	i
28	<b>1g</b>	4	KOH	<b>14H</b>	39	62	j
29	<b>1g</b>	4	CsOH	<b>14H</b>	54	74	
30	<b>1g</b>	20	CsOH	<b>14H</b>	<b>61</b>	79	k
31	<b>1h</b>	20	CsOH	<b>14H</b>	60	<b>80</b>	
32	<b>1c</b>	20	CsOH	<b>14H</b>	22	35	l
33	<b>1d</b>	20	CsOH	<b>14H</b>	<10	n.d.	l
34	<b>1g</b>	20	CsOH	<b>14H</b>	52	68	l
35	<b>1h</b>	20	CsOH	<b>14H</b>	51	67	l
36	<b>1g</b>	20	CsOH	<b>14H</b>	0	n.d.	m
37	<b>1c</b>	20	Cs <sub>2</sub> CO <sub>3</sub>	<b>14I</b>	<5	n.d.	
38	<b>1g</b>	20	Cs <sub>2</sub> CO <sub>3</sub>	<b>14I</b>	33	<b>88</b>	
39	<b>1h</b>	20	Cs <sub>2</sub> CO <sub>3</sub>	<b>14I</b>	<5	n.d.	
40	<b>1c</b>	20	Cs <sub>2</sub> CO <sub>3</sub>	<b>14I</b>	<b>33</b>	57	n
41	<b>1d</b>	20	Cs <sub>2</sub> CO <sub>3</sub>	<b>14I</b>	26	51	n
42	<b>1g</b>	20	Cs <sub>2</sub> CO <sub>3</sub>	<b>14I</b>	23	83	n
43	<b>1h</b>	20	Cs <sub>2</sub> CO <sub>3</sub>	<b>14I</b>	20	83	n

<sup>a</sup> 0.25 mmol of substrate, 0.30 mmol of electrophile, 1 mol % of catalyst with (*S,S*)-configuration, 1.5 mL toluene, 0.5 mL KOH or CsOH (50%) or solid Cs<sub>2</sub>CO<sub>3</sub> (5 equiv.), vigorous stirring at 0 °C for the time indicated. <sup>b</sup> Yield after extractive work-up based on <sup>1</sup>H-NMR integration of signals of product and bibenzyl added as internal standard. <sup>c</sup> Determined by chiral HPLC (Chiralcel ODH or Chiralpak ADH), products with (*R*)-configuration predominating. <sup>d</sup> 99% ee, 80% isolated yield, 0 °C, 24 h [1]. <sup>e</sup> 99% ee 86% over two steps, 0 °C, 24 h [1]. <sup>f</sup> 99% ee 89% isolated yield, 0 °C, 15 h [1]. <sup>g</sup> 12% of **15** formed. <sup>h</sup> 13% of **15** formed. <sup>i</sup> 37% of **15** formed. <sup>j</sup> 10 equiv. of KI added, 27% of **15** formed. <sup>k</sup> 8% of **15** formed. <sup>l</sup> Instead of **13H** the corresponding 4-iodocrotonate was used. <sup>m</sup> Instead of **13H** the corresponding 4-chlorocrotonate was used, resulting in formation of 53% of **15** and 27% of intermediate **16** (see structure below). <sup>n</sup> Instead of **13I** the corresponding iodo compound was used.

1-(*tert*-Butyl) 5-ethyl 3-(chloromethyl)-2-((diphenylmethylene)amino)pentanedioate (**16**):  $^1\text{H-NMR}$   $\delta$ :



7.63–7.67 (m, 2H); 7.37–7.45 (m, 4H); 7.30–7.35 (m, 2H); 7.18–7.22 (m, 2H); 4.18 (d,  $J$  = 4.1 Hz, 1H); 4.12 (qm,  $J$  = 7.1 Hz, 2H); 3.69 (dd,  $J$  = 11.0, 5.0 Hz, 1H); 3.54 (dd,  $J$  = 10.9, 8.2 Hz, 1H); 2.90–2.97 (m, 1H); 2.73 (dd,  $J$  = 16.5, 4.8 Hz, 1H); 2.67 (dd,  $J$  = 16.5, 8.5 Hz, 1H); 1.42 (s, 9H); 1.24 (t,  $J$  = 7.1 Hz, 3H) ppm.  $^{13}\text{C-NMR}$   $\delta$ : 172.26 (C); 172.24 (C); 169.99 (C); 139.29 (C); 136.19 (C); 130.47 (CH); 128.79 (CH); 128.61 (CH); 128.30 (CH); 127.97 (CH); 127.76 (CH); 81.64 (C); 65.23 (CH);

60.52 ( $\text{CH}_2$ ); 45.34 ( $\text{CH}_2$ ); 41.32 (CH); 33.96 ( $\text{CH}_2$ ); 27.93 ( $\text{CH}_3$ ); 14.14 ( $\text{CH}_3$ ) ppm. HRMS (ESI) calcd for  $\text{C}_{25}\text{H}_{30}\text{ClNaNO}_4$  [ $\text{M} + \text{Na}$ ] $^+$ : 466.1761; found. 466.1746.

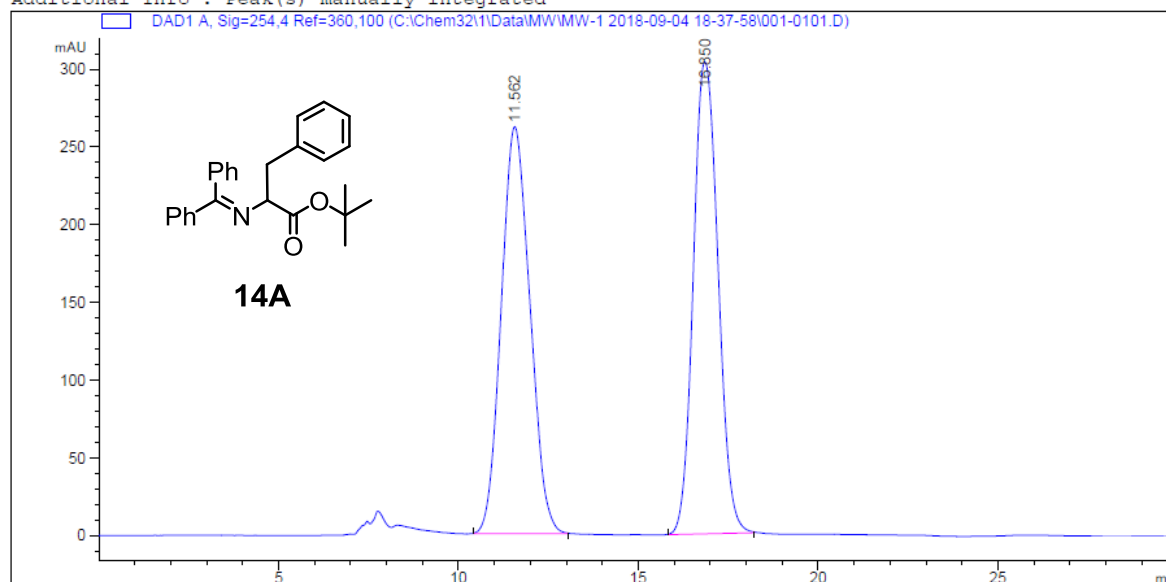
*Ethyl (E)-4-chlorobut-2-enoate*: To a stirred solution of ethyl 4-bromocrotonate (772 mg, 3.0 mmol 75% purity) in DMF (1 mL) was added dry LiCl (636 mg, 5 equiv.) at room temperature. After 24 h, the suspension was poured into water/ether (10 mL/30 mL) and the organic phase was washed with water (4  $\times$  10 mL) and dried ( $\text{MgSO}_4$ ). The crude product was purified by chromatography (EtOAc (0 $\rightarrow$ 15%)/heptane) to afford the product as a colorless oil; yield 80–90%.  $^1\text{H-NMR}$   $\delta$ : 6.97 (dt,  $J$  = 15.4, 6.1 Hz, 1H); 6.10 (dt,  $J$  = 15.3, 1.6 Hz, 1H); 4.22 (q,  $J$  = 7.1 Hz, 2H); 4.16 (dd,  $J$  = 6.1, 1.6 Hz, 2H); 1.30 (t,  $J$  = 7.1 Hz, 3H) ppm.  $^{13}\text{C-NMR}$   $\delta$ : 165.61 (C); 141.59 (CH); 124.16 (CH); 60.71 ( $\text{CH}_2$ ); 42.47 ( $\text{CH}_2$ ); 14.17 ( $\text{CH}_3$ ) ppm. HRMS (ESI) calculated for  $\text{C}_4\text{H}_4\text{ClO}$  [ $\text{M} - \text{OC}_2\text{H}_5$ ] $^+$ : 102.9951, found 102.9941.

# UV traces of chiral HPLC separations

Sample Name: W STD 14

```
=====
Acq. Operator   : SYSTEM                      Seq. Line :    1
Acq. Instrument : LC1200                     Location  :    1
Injection Date  : 04.09.2018 18:39:02        Inj       :    1
                                           Inj Volume: 5.000 µl
Method         : C:\Chem32\1\Data\MW\MW-1 2018-09-04 18-37-58\CHIR_ODH.M (Sequence Method)
Last changed    : 04.09.2018 18:37:59 by SYSTEM
Sample Info     : ODH, 1% iPrOH, 25°C, 0.5 mL min-1
=====
```

Additional Info : Peak(s) manually integrated



## Area Percent Report

```
=====
Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs
=====
```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.562	BB	0.8486	1.45102e4	261.82501	50.0590
2	16.850	BB	0.7523	1.44760e4	303.57013	49.9410

Totals :                      2.89863e4    565.39514

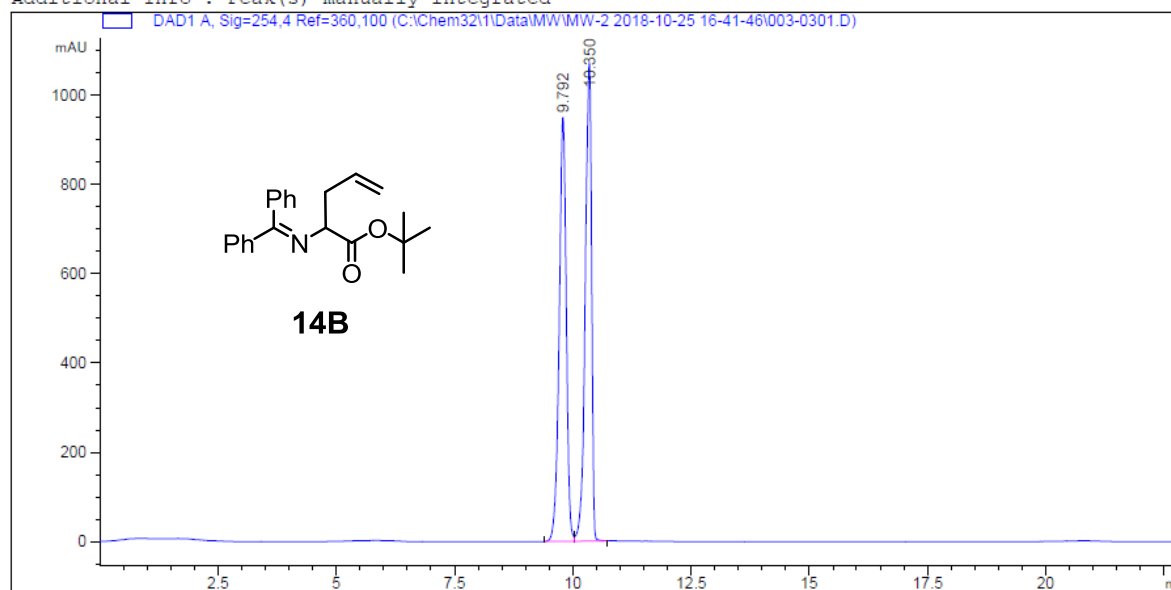
Sample Name: STD-D1

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : LCI200                      Location  :    3
Injection Date  : 25.10.2018 17:48:21         Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : C:\Chem32\1\Data\MW\MW-2 2018-10-25 16-41-46\CHIR_ADH.M
Last changed    : 25.10.2018 17:46:20 by SYSTEM
Analysis Method : C:\Chem32\1\Data\MW\MW-2 2018-10-25 16-41-46\CHIR_ADH.M (Sequence Method)
Last changed    : 25.10.2018 17:47:16 by SYSTEM
Sample Info     : W STD-D1 Chiralpak ADH 2-PrOH/heptane (0.5:99.5) 0.5 mL min-1, 25 °C

```

Additional Info : Peak(s) manually integrated



```

=====
                        Area Percent Report
=====

```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs

```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.792	BV	0.1596	9905.24414	949.96033	49.8037
2	10.350	VB	0.1506	9983.34277	1072.17236	50.1963

Totals : 1.98886e4 2022.13269

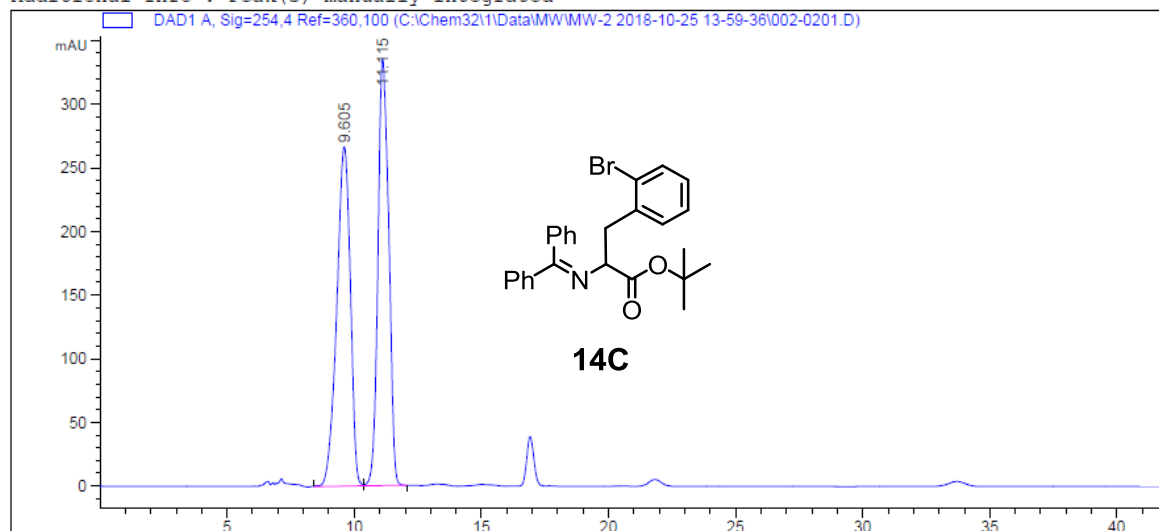


Sample Name: STD-C1

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Acq. Instrument : LC1200                     Location  :    2
Injection Date  : 25.10.2018 15:01:57        Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : C:\Chem32\1\Data\MW\MW-2 2018-10-25 13-59-36\CHIR_ADH.M
Last changed    : 25.10.2018 15:44:49 by SYSTEM
                  (modified after loading)
Analysis Method : C:\Chem32\1\Data\MW\MW-2 2018-10-25 13-59-36\CHIR_ADH.M (Sequence Method)
Last changed    : 20.08.2019 17:30:28 by SYSTEM
                  (modified after loading)
Sample Info     : W STD-C1 Chiralpak ADH 2-PrOH/heptane (1:99) 0.5 mL min-1, 25 °C
  
```

Additional Info : Peak(s) manually integrated



```

=====
                          Area Percent Report
=====
  
```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution      :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.605	BV	0.5879	9920.35547	266.72058	50.0998
2	11.115	VB	0.4896	9880.82813	335.50867	49.9002

```
Totals :                      1.98012e4  602.22925
```

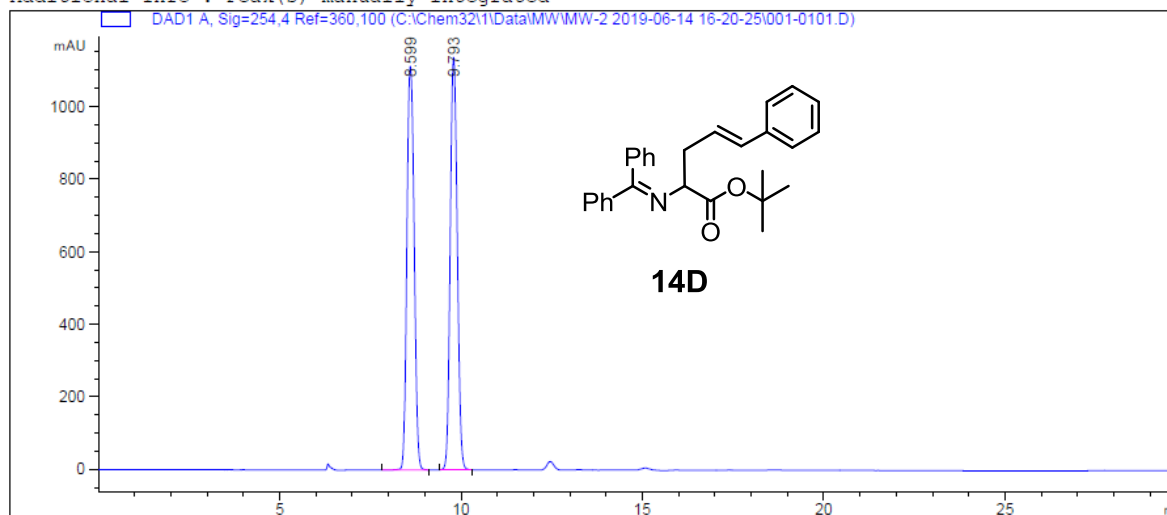
Sample Name: W STD K01

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    1
Acq. Instrument : LC1200                     Location  :    1
Injection Date  : 14.06.2019 16:21:31        Inj       :    1
                                           Inj Volume: 5.000 µl
Method         : C:\Chem32\1\Data\MW\MW-2 2019-06-14 16-20-25\CHIR_ADH.M (Sequence Method)
Last changed    : 14.06.2019 16:20:27 by SYSTEM
Sample Info     : W STD K01/5-6 (RACEMAT) Chiralpak ADH 2-PrOH/heptane (5:95) 0.5 mL min-1, 25 °C
=====

```

Additional Info : Peak(s) manually integrated



```

=====
Area Percent Report
=====

```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs

```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.599	BB	0.2245	1.54999e4	1112.48364	50.0858
2	9.793	BB	0.2165	1.54468e4	1136.89917	49.9142

```
Totals :                3.09466e4  2249.38281
```

```

=====
*** End of Report ***
=====

```

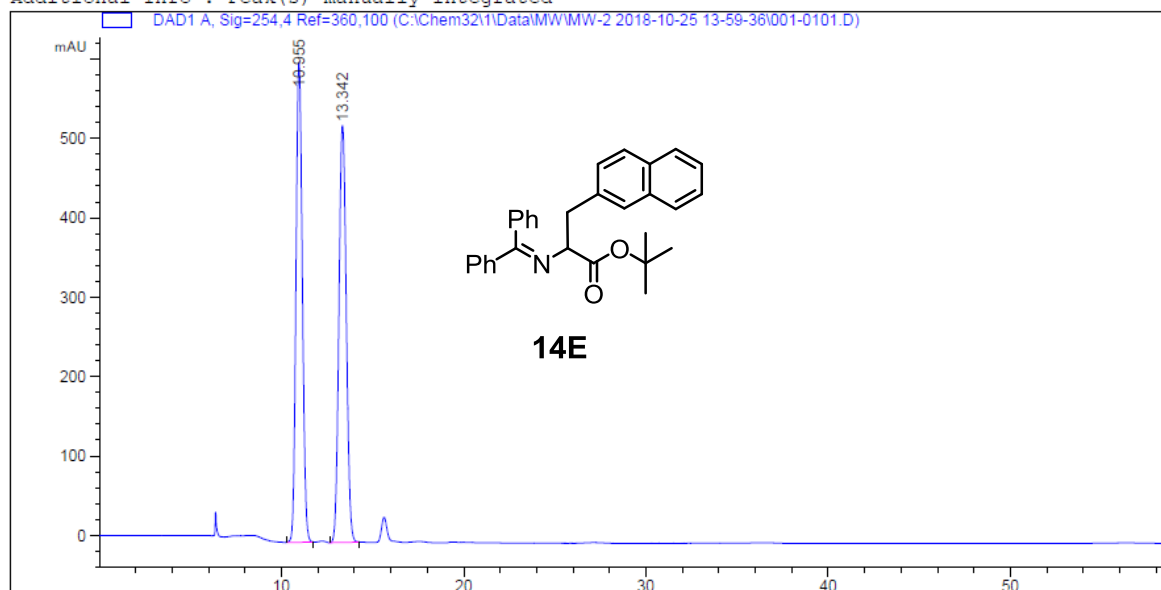
Sample Name: STD-B1

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    1
Acq. Instrument : LC1200                     Location  :    1
Injection Date  : 25.10.2018 14:00:43        Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : C:\Chem32\1\Data\MW\MW-2 2018-10-25 13-59-36\CHIR_ADH.M
Last changed    : 25.10.2018 14:01:05 by SYSTEM
                  (modified after loading)
Analysis Method : C:\Chem32\1\Data\MW\MW-2 2018-10-25 13-59-36\CHIR_ADH.M (Sequence Method)
Last changed    : 25.10.2018 15:00:53 by SYSTEM
Sample Info     : W STD-B1 Chiralpak ADH 2-PrOH/heptane (1:99) 0.5 mL min-1, 25 °C

```

Additional Info : Peak(s) manually integrated



```

=====
                          Area Percent Report
=====

```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs

```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	10.955	BB	0.3839	1.47045e4	604.30414	50.1185
2	13.342	VB	0.4392	1.46350e4	524.99420	49.8815

Totals :                      2.93395e4   1129.29834

Sample Name: CHSC3/2

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    2
Acq. Instrument : LC1200                     Location  :    2
Injection Date  : 15.02.2019 10:57:09        Inj       :    1
                                           Inj Volume: 5.000 µl

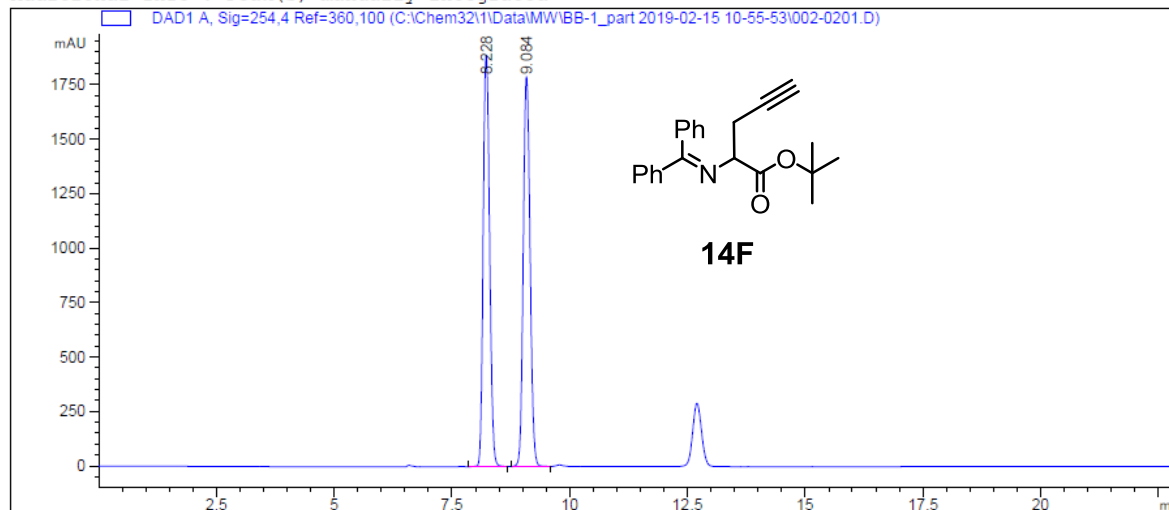
Acq. Method     : C:\Chem32\1\Data\MW\BB-1_part 2019-02-15 10-55-53\CHIR_ADH.M
Last changed    : 15.02.2019 11:19:19 by SYSTEM
                  (modified after loading)

Analysis Method : C:\Chem32\1\Data\MW\BB-1_part 2019-02-15 10-55-53\CHIR_ADH.M (Sequence Method)
Last changed    : 15.02.2019 11:23:11 by SYSTEM
                  (modified after loading)

Sample Info     : CHSC 03 racem Prod, Chiralpak ADH, 5% 2-PrOH, 95% Heptan, 0.5 mlmin-1, 25 °C

```

Additional Info : Peak(s) manually integrated



```

=====
                          Area Percent Report
=====

```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs

```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.228	VB	0.1505	1.78747e4	1887.15088	49.8157
2	9.084	BV	0.1574	1.80070e4	1790.12805	50.1843

```
Totals :                      3.58817e4  3677.27893
```

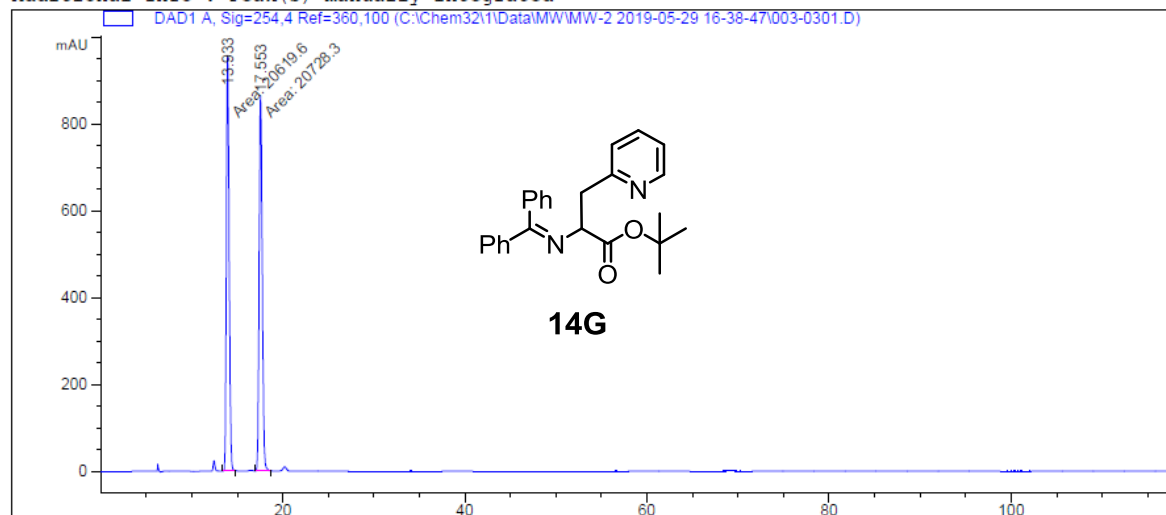
Sample Name: W STD H01

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : LC1200                     Location  :    3
Injection Date  : 29.05.2019 17:42:18        Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : C:\Chem32\1\Data\MW\MW-2 2019-05-29 16-38-47\CHIR_ADH.M
Last changed    : 29.05.2019 17:43:15 by SYSTEM
                  (modified after loading)
Analysis Method : C:\Chem32\1\Data\MW\MW-2 2019-05-29 16-38-47\CHIR_ADH.M (Sequence Method)
Last changed    : 29.05.2019 16:38:48 by SYSTEM
Sample Info     : W STD H01/8-9 RACEMAT Chiralpak ADH 2-PrOH/heptane (5:95) 0.5 L min-1, 25 °C

```

Additional Info : Peak(s) manually integrated



```

=====
                          Area Percent Report
=====

```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs

```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.933	MF	0.3603	2.06196e4	953.83917	49.8685
2	17.553	MF	0.4002	2.07283e4	863.33826	50.1315

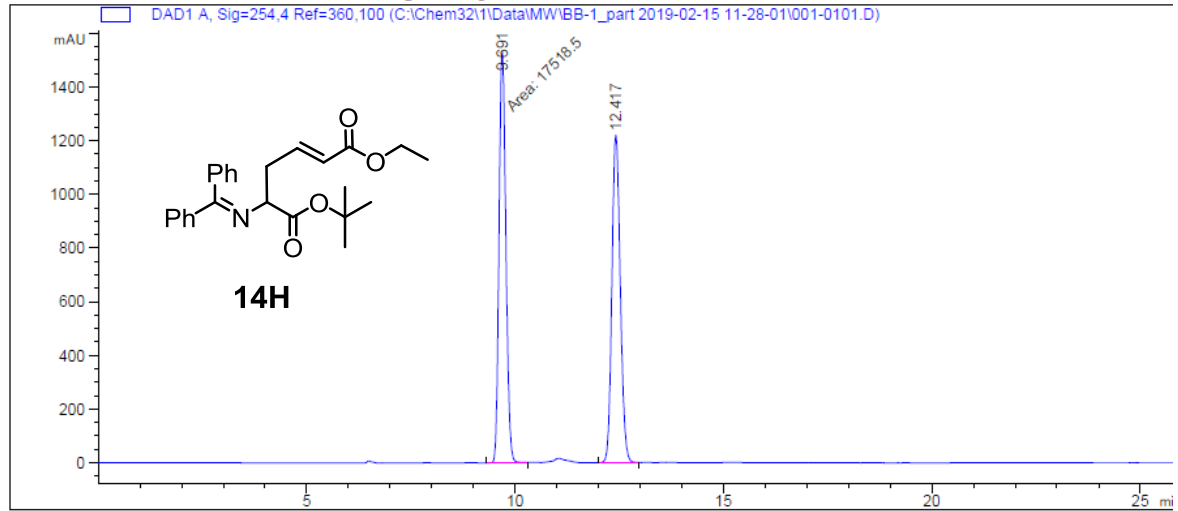
Totals :                      4.13479e4   1817.17743

Sample Name: CHSC 1/3C/8-10

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    1
Acq. Instrument : LC1200                     Location  :    1
Injection Date  : 15.02.2019 11:29:05        Inj       :    1
                                           Inj Volume: 5.000 µl
Acq. Method     : C:\Chem32\1\Data\MW\BB-1_part 2019-02-15 11-28-01\CHIR_ADH.M
Last changed    : 15.02.2019 11:54:50 by SYSTEM
                  (modified after loading)
Analysis Method : C:\Chem32\1\Data\MW\BB-1_part 2019-02-15 11-28-01\CHIR_ADH.M (Sequence Method)
Last changed    : 15.02.2019 12:07:25 by SYSTEM
                  (modified after loading)
Sample Info     : CHSC 01 racem Prod, Chiralpak ADH, 5% 2-PrOH, 95% Heptan, 0.5 mlmin-1, 25 °C
  
```

Additional Info : Peak(s) manually integrated



```

=====
                          Area Percent Report
=====
  
```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.691	MF	0.1906	1.75185e4	1531.91235	50.6581
2	12.417	BB	0.2192	1.70633e4	1220.04663	49.3419

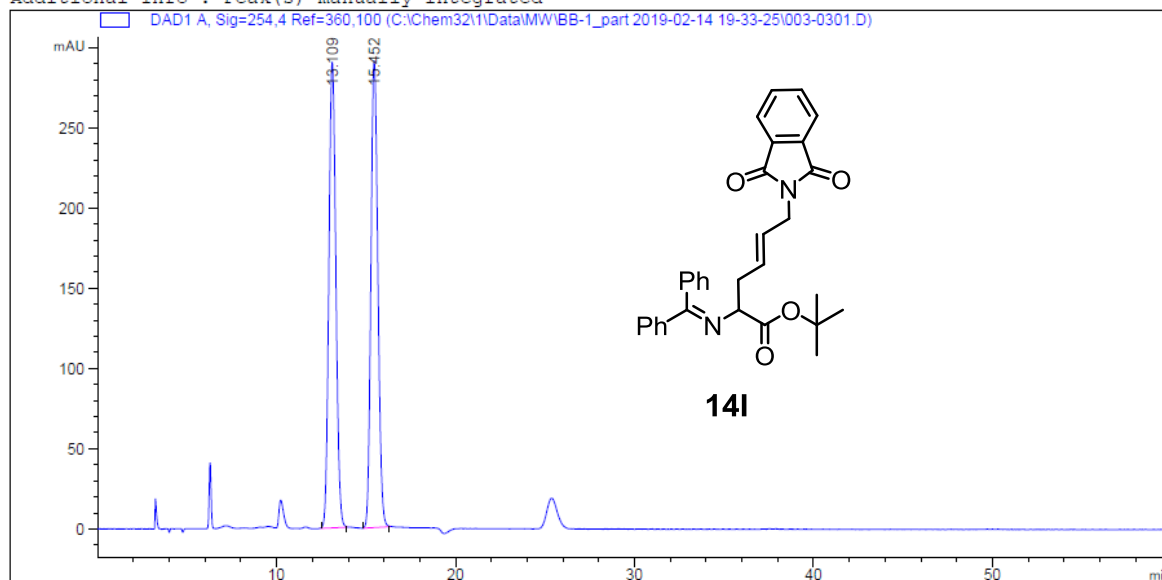
```
Totals :                      3.45818e4  2751.95898
```

Sample Name: CHSC4/8-9

```

=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : LC1200                     Location  :    3
Injection Date  : 14.02.2019 19:35:09        Inj       :    1
                                           Inj Volume: 20.000 µl
Method         : C:\Chem32\1\Data\MW\BB-1_part 2019-02-14 19-33-25\CHIR_ADH.M (Sequence Method)
Last changed    : 14.02.2019 19:33:26 by SYSTEM
Sample Info     : CHSC 04 racem Prod, Chiralpak ADH, 5% 2-PrOH, 95% Heptan, 0.5 mlmin-1, 25 °C
  
```

Additional Info : Peak(s) manually integrated



```

=====
                          Area Percent Report
=====
  
```

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Do not use Multiplier & Dilution Factor with ISTDs
  
```

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.109	BB	0.4288	7931.43066	290.25415	50.4199
2	15.452	BB	0.4187	7799.31787	290.97043	49.5801

Totals :                      1.57307e4    581.22458

## X-ray Structure Analyses

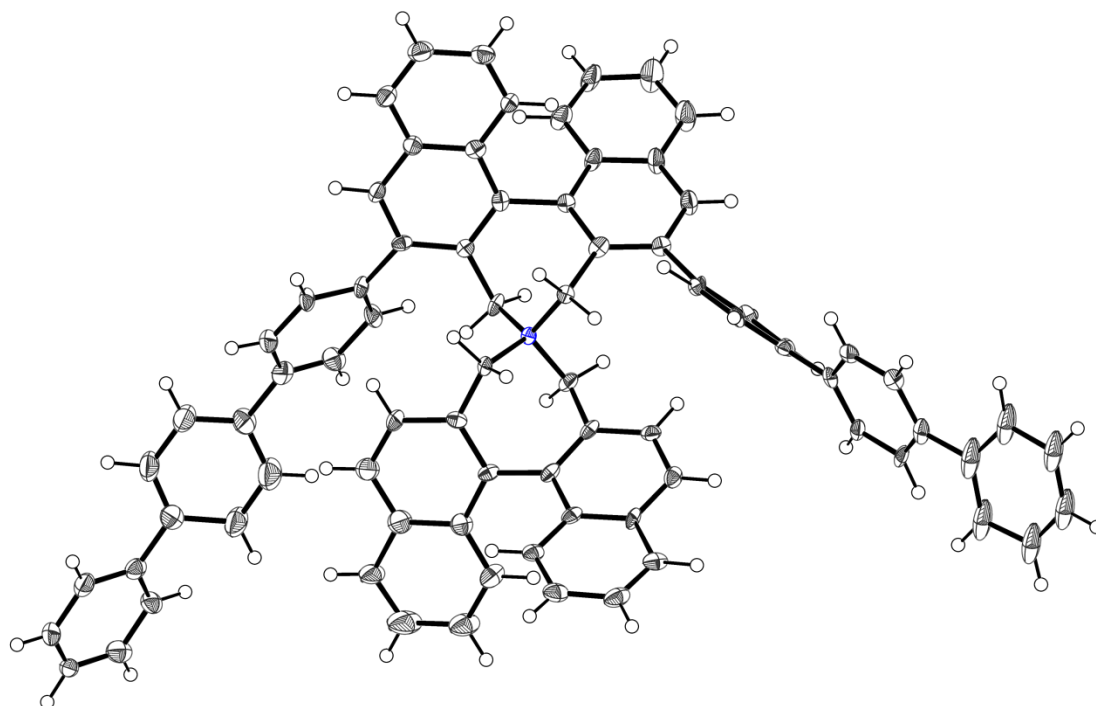
The X-ray intensity data were measured on Bruker D8 Venture and X8 APEX2 diffractometer equipped with multilayer monochromators, Mo K/ $\alpha$  INCOATEC micro focus sealed tubes and Oxford and Cryoflex2 cooling systems. The structure was solved by *direct methods*. Non-hydrogen atoms were refined with *anisotropic displacement parameters*. Hydrogen atoms were inserted at calculated positions and refined with riding model. The following software was used: *Bruker SAINT software package* [2] using a narrow-frame algorithm for frame integration, *SADABS* [3] for absorption correction, *OLEX2* [4] for structure solution, refinement, molecular diagrams and graphical user-interface, *Shelxle* [5] for refinement and graphical user-interface *SHELXS-2015* [6] for structure solution, *SHELXL-2015* [7] for refinement, *Platon* [8] for symmetry check. Experimental data and CCDC-Codes (Available online: <http://www.ccdc.cam.ac.uk/conts/retrieving.html>) can be found in Table S2. Crystal data, data collection parameters, and structure refinement details are given in Tables S3 to S8. Crystal structures are visualized in Figures S1 to S5.

**Table S2.** Experimental parameter and CCDC-Codes.

Sample	Machine	Source	Temp.	Detector Distance	Time/Frame	#Frames	Frame width	CCDC
	Bruker		[K]	[mm]	[s]		[°]	
(R)(R)- <b>1d</b>	X8	Mo	150	30	21	2236	0.700	1956062
(R) <sub>bina</sub> (S) <sub>biphe</sub> - <b>1e'</b>	X8	Mo	150	30	60	1049	0.700	1956063
[(R)- <b>7</b> ] <sub>2</sub> (S,S)-Dibenzoyltartrate	D8	Mo	100	30	30	1891	0.500	1956064



(11*bR*,11*b'R*)-2,6-Di([1,1':4',1''-terphenyl]-4-yl)-3,3',5,5'-tetrahydro-4,4'-spirobi[dinaphtho[2,1-*c*:1',2'-*e*]azepin]-4-ium bromide (*R*)(*R*)-**1d**



**Figure S1.** Crystal structure of (*R*)(*R*)-**1d**. C-C- Bond precision: 0.0121 Å. Solvent and counter ion omitted for clarity. Squeeze was used to cut the volume and the corresponding electron densities, because small electron densities in excluded volumes could not be matched. Details see cif-code.

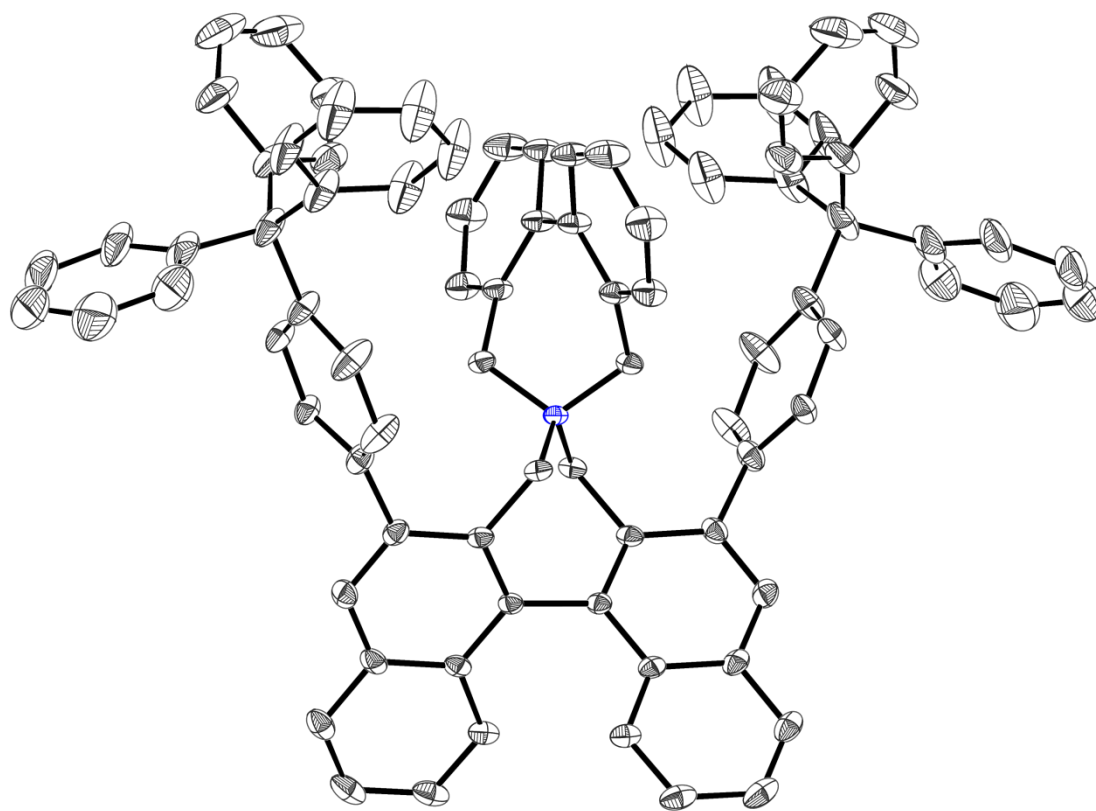
**Table S3.** Sample and crystal data.

Chemical formula	C <sub>82.5</sub> H <sub>61</sub> BrCl <sub>5</sub> N	Crystal system	orthorhombic	
Formula weight [g/mol]	1323.48	Space group	<i>P</i> 212121	
Temperature [K]	150	<i>Z</i>	4	
Measurement method	\f and \w scans	Volume [Å <sup>3</sup> ]	7229.8(11)	
Radiation (Wavelength [Å])	MoKα (λ = 0.71073)	Unit cell dimensions [Å] and [°]	9.1716(8)	90
Crystal size / [mm <sup>3</sup> ]	0.3 × 0.14 × 0.11		26.018(2)	90
Crystal habit	clear colourless block		30.298(3)	90
Density (calculated) / [g/cm <sup>3</sup> ]	1.216	Absorption coefficient / [mm <sup>-1</sup> ]	0.797	
Abs. correction Tmin	0.5297	Abs. correction Tmax	0.746	
Abs. correction type	multiscan	<i>F</i> (000) [e <sup>-</sup> ]	2732	

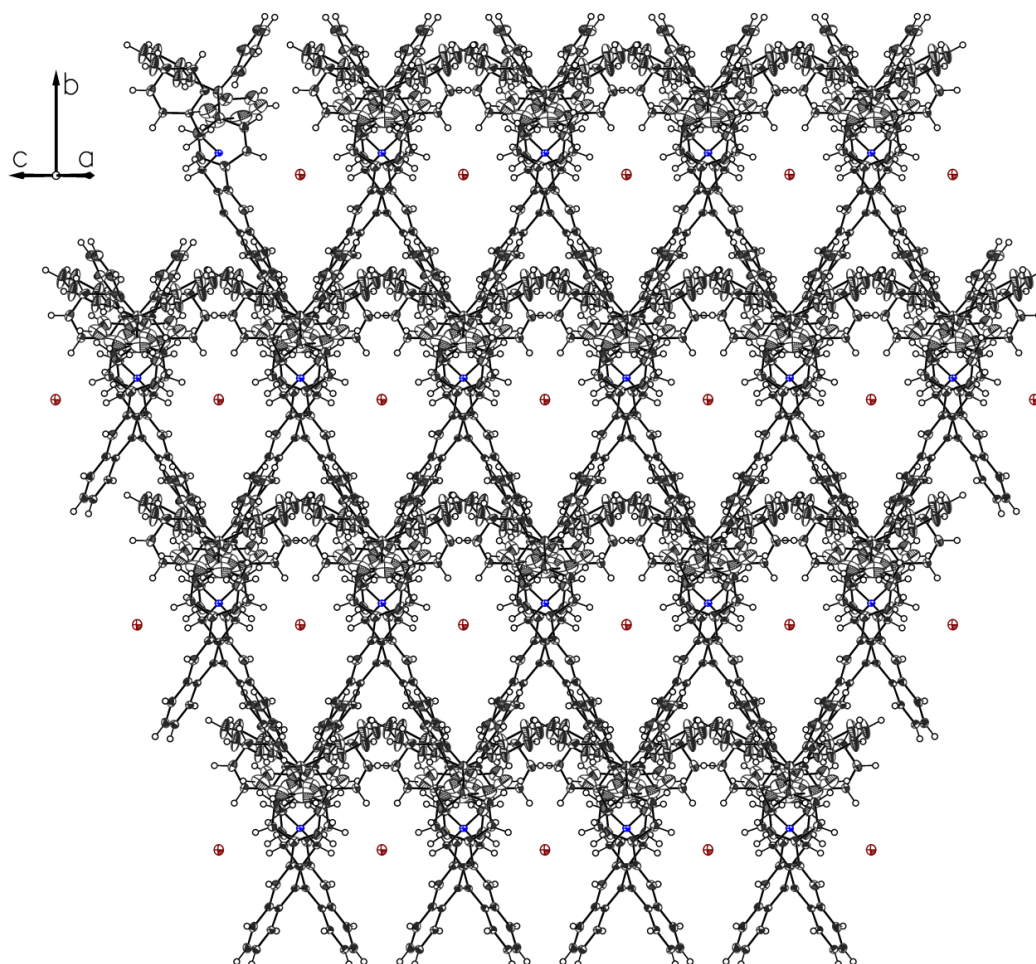
**Table S4.** Data collection and structure refinement.

<b>Index ranges</b>	$-11 \leq h \leq 11, -31 \leq k \leq 31, -36 \leq l \leq 36$	<b>Theta range for data collection [°]</b>	2.064 to 50.7	
<b>Reflections number</b>	217019	<b>Data / restraints / parameters</b>	13222/0/838	
<b>Refinement method</b>	Least squares	<b>Final R indices</b>	all data	R1 = 0.0955, wR2 = 0.2433
<b>Function minimized</b>	$\sum w(F_o^2 - F_c^2)^2$		$I > 2\sigma(I)$	R1 = 0.0812, wR2 = 0.2278
<b>Goodness-of-fit on <math>F^2</math></b>	1.064	<b>Weighting scheme</b>	$w = 1/[\sigma^2(F_o^2) + (0.1579P)^2 + 14.4721P]$	
<b>Largest diff. peak and hole [e Å<sup>-3</sup>]</b>	1.91/-1.02		where $P = (F_o^2 + 2F_c^2)/3$	

*(R)*-2',6'-Bis(4-tritylphenyl)-3',5,5',7-tetrahydrospiro[dibenzo[*c,e*]azepine-6,4'-dinaphtho[2,1-*c*:1',2'-*e*]azepin]-6-ium bromide (*R*)-**1e'**



**Figure S2.** Crystal structure of (*R*)-**1e'**. C-C- Bond precision: 0.0074 Å. Solvent, counter ion and hydrogens omitted for clarity.



**Figure S3.** Packing view. Solvent omitted for clarity.

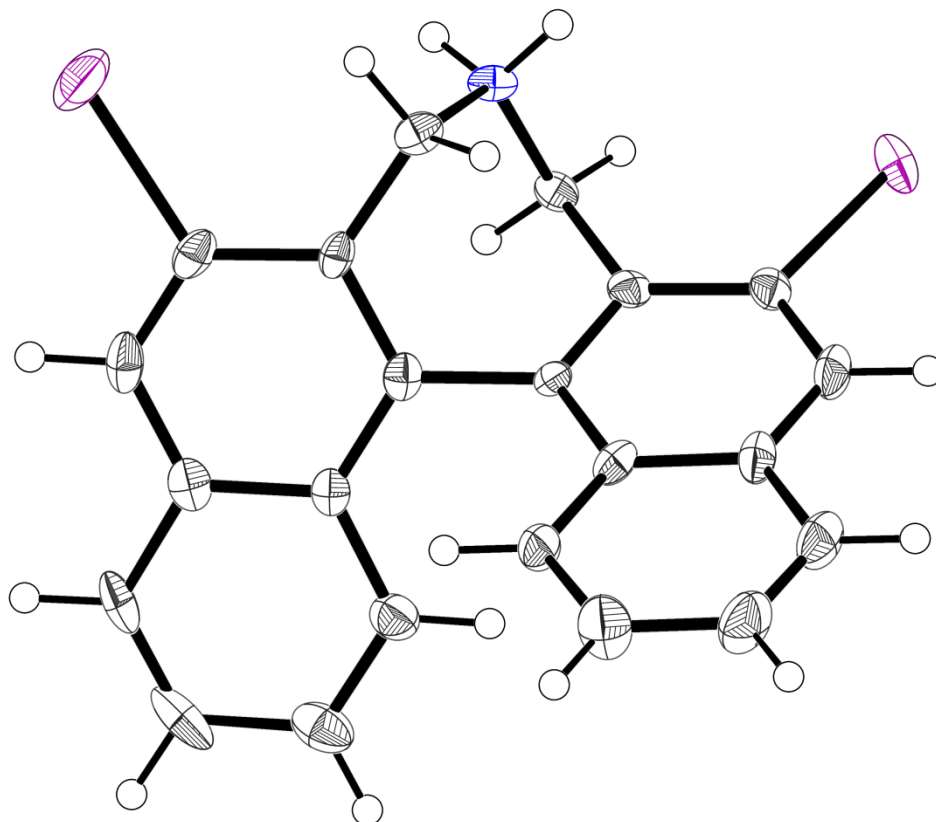
**Table S5.** Sample and crystal data.

<b>Chemical formula</b>	$C_{58}H_{48}Br_{0.5}N_{0.5}$	<b>Crystal system</b>	monoclinic	
<b>Formula weight [g/mol]</b>	791.92	<b>Space group</b>	$C2$	
<b>Temperature [K]</b>	150	<b>Z</b>	4	
<b>Measurement method</b>	\f and \w scans	<b>Volume [<math>\text{\AA}^3</math>]</b>	4478.3(5)	
<b>Radiation (Wavelength [<math>\text{\AA}</math>])</b>	MoK $\alpha$ ( $\lambda = 0.71073$ )	<b>Unit cell dimensions [<math>\text{\AA}</math>] and [<math>^\circ</math>]</b>	21.7990(14)	90
<b>Crystal size / [<math>\text{mm}^3</math>]</b>	$0.25 \times 0.23 \times 0.1$		23.2291(13)	94.251(2)
<b>Crystal habit</b>	clear colourless block		8.8684(5)	90
<b>Density (calculated) / [<math>\text{g}/\text{cm}^3</math>]</b>	1.175	<b>Absorption coefficient / [<math>\text{mm}^{-1}</math>]</b>	0.51	
<b>Abs. correction Tmin</b>	0.5749	<b>Abs. correction Tmax</b>	0.746	
<b>Abs. correction type</b>	multiscan	<b>F(000) [<math>e^-</math>]</b>	1668	

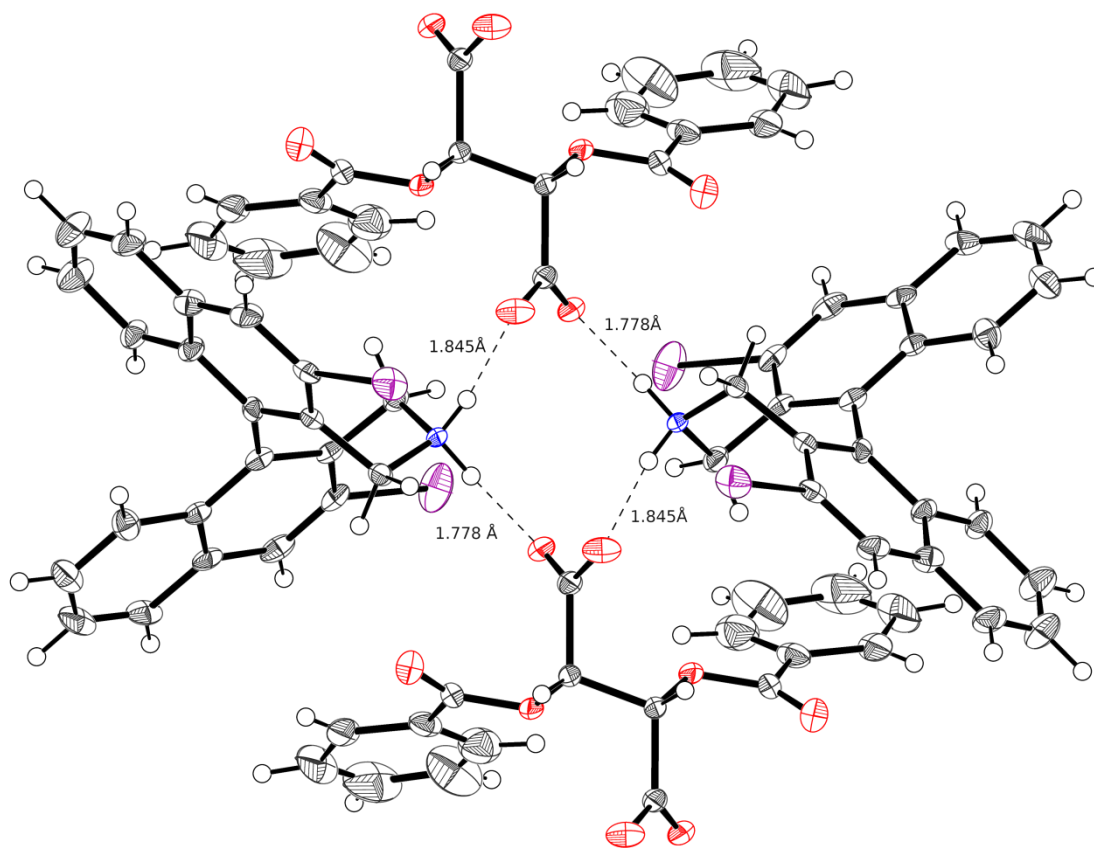
**Table S6.** Data collection and structure refinement.

<b>Index ranges</b>	$-30 \leq h \leq 29, -32 \leq k \leq 32, -12 \leq l \leq 12$	<b>Theta range for data collection [°]</b>	2.566 to 60.592	
<b>Reflections number</b>	43599	<b>Data / restraints / parameters</b>	13255/32/450	
<b>Refinement method</b>	Least squares	<b>Final R indices</b>	all data	R1 = 0.0876, wR2 = 0.2295
<b>Function minimized</b>	$\sum w(F_o^2 - F_c^2)^2$		$I > 2\sigma(I)$	R1 = 0.0802, wR2 = 0.2193
<b>Goodness-of-fit on <math>F^2</math></b>	1.048	<b>Weighting scheme</b>	$w = 1/[\sigma^2(F_o^2) + (0.1825P)^2 + 2.6917P]$	
<b>Largest diff. peak and hole [e Å<sup>-3</sup>]</b>	1.49/-0.65		where $P = (F_o^2 + 2F_c^2)/3$	

*(R)*-2,6-Diiodo-4,5-dihydro-3*H*-dinaphtho[2,1-*c*:1',2'-*e*]azepin-4-ium ion [*(R)*-**7**]<sup>+</sup>



**Figure S4.** Crystal structure of [*(R)*-**7**]<sub>2</sub>(*S,S*)-Dibenzoyltartrate. C-C- Bond precision: 0.0129 Å. Solvent and counter ion omitted for clarity.



**Figure S5.**  $[(R)-7]^+$  and counter ion form two independent hydrogen bonds because of symmetry for (two each because of symmetry reasons) construct a rhombus. Bond length and rhombus visualized.

**Table S7.** Sample and crystal data.

<b>Chemical formula</b>	$C_{32}H_{24}Cl_2I_2NO_4$	<b>Crystal system</b>	monoclinic	
<b>Formula weight [g/mol]</b>	811.22	<b>Space group</b>	$C2$	
<b>Temperature [K]</b>	100	<b>Z</b>	4	
<b>Measurement method</b>	\f and \w scans	<b>Volume [<math>\text{\AA}^3</math>]</b>	3028.14(18)	
<b>Radiation (Wavelength [<math>\text{\AA}</math>])</b>	MoK $\alpha$ ( $\lambda = 0.71073$ )	<b>Unit cell dimensions [<math>\text{\AA}</math>] and [<math>^\circ</math>]</b>	28.7403(11)	90
<b>Crystal size / [<math>\text{mm}^3</math>]</b>	$0.07 \times 0.07 \times 0.05$		12.0555(4)	101.095(2)
<b>Crystal habit</b>	clear colourless block		8.9062(3)	90
<b>Density (calculated) / [<math>\text{g}/\text{cm}^3</math>]</b>	1.779	<b>Absorption coefficient / [<math>\text{mm}^{-1}</math>]</b>	2.292	
<b>Abs. correction Tmin</b>	0.2263	<b>Abs. correction Tmax</b>	0.265	
<b>Abs. correction type</b>	multiscan	<b>F(000) [<math>e^-</math>]</b>	1580	

**Table S8.** Data collection and structure refinement.

Index ranges	$-34 \leq h \leq 34, -14 \leq k \leq 14, -10 \leq l \leq 10$	Theta range for data collection [°]	4.66 to 50.694	
Reflections number	47080	Data / restraints / parameters	5534/7/364	
Refinement method	Least squares	Final R indices	all data	R1 = 0.0367, wR2 = 0.0897
Function minimized	$\sum w(F_o^2 - F_c^2)^2$		$I > 2\sigma(I)$	R1 = 0.0362, wR2 = 0.0894
Goodness-of-fit on $F^2$	1.129	Weighting scheme	$w = 1/[\sigma^2(F_o^2) + (0.0383P)^2 + 21.9719P]$	
Largest diff. peak and hole [e Å <sup>-3</sup> ]	0.85/-1.24		where $P = (F_o^2 + 2F_c^2)/3$	

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