SUPPLEMENTARY MATERIAL

16-Hydroxy-Lycopersene, a Polyisoprenoid Alcohol Isolated from *Tournefortia hirsutissima*, Inhibits Nitric Oxide Production in RAW 264.7 Cells and Induces Apoptosis in Hep3B Cells

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Figure S2. Inhibition of the NO production in LPS-stimulated RAW 264.7 cells by Th-H, Th-D, Th-HA, F4, F4-1, F4-2, and F4-4 to indicated doses.

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Figure S15. The cell cycle of (A) Hep3B, (B) HepG2, (C) PC3 and (D) HeLa cells by flow cytometry treated with **1** to its Cl₅₀ and PTX 10 nM.



Figure S1. Effect of extracts of n-hexane (Th-H), dicloromethane (Th-D) and hydroalcoholic (Th-D) from leaves of *Tournefortia hirsutissima* at 1 mg/ear dose on the edema in mice ear induced by TPA. Indomethacin (1 mg/ear) was used as control. * P < 0.05, ** P < 0.01, *** P < 0.001, **** P < 0.0001 compared to the TPA control.



Figure S2. Inhibition of the NO production in LPS-stimulated RAW 264.7 cells by Th-H, Th-D, Th-HA, F4, F4-1, F4-2, and F4-4 to indicated concentrations. Cells were treated with the investigated samples, DMSO (0.4%, v/v) or indomethacin ($30 \mu g/mL$) 2 h before stimulation with LPS ($1.0 \mu g/mL$). The nitrite concentration was determined by Griess

method and is expressed in percentage. All data represent the mean \pm standard deviation of at least three independent experiments performed by triplicate. Statistical significance was determined by one-way ANOVA followed by Dunnett's test. * *P* < 0.05, ** *P* < 0.01, *** *P* < 0.001 and **** *P* < 0.0001 compared with LPS group.



Figure S3. Effect of Th-H, Th-D, Th-HA, F4, F5, F4-1, F4-2, and F4-4 to indicated concentrations on cell viability of RAW 264.7 cells by using MTS assay. Cell viability is expressed in percentage. All data represent the mean ± standard deviation of at least three independent experiments performed by triplicate. Statistical significance was

determined by one-way ANOVA followed by Dunnett's test. * P < 0.05, ** P < 0.01, *** P < 0.001 and **** P < 0.0001 compared with control cells group (without treatment).



Figure S4. Analysis of F4-2-1 by using GC/MS. The most abundant component in this fraction was bis (2-ethylhexyl) phthalate with a percentage of 55.84%.



Figure S5. Chromatogram of F4-2-2 by using analytical HPLC, monitored at 210 nm using Merck column (Performance RP-18e, 100 x 4.6 mm). The mobile phase consisted of a gradient of iPrOH : MeOH : H₂O (30 : 67 : 03 to 30 : 70 : 00, *v*/*v*). Flow rate was 2.0 mL/min and sample injection of 7 μ L (1.5 mg/mL). The peaks with retention times at 1.90, 3.20 and 4.14 min correspond to **1**, **2** and **3**, respectively.



Figure S6. ¹H NMR spectrum (500 MHz, benzene-*d*₆) of compound 1.



Figure S7. ¹H NMR spectrum expansions (500 MHz, benzene-*d*₆) of compound **1**.







Figure S9. ¹H - ¹H COSY spectrum (benzene-*d*₆) of compound **1**.



Figure S10. TOCSY spectrum (benzene-*d*₆) of compound **1**.



Figure S11. HSQC spectrum (benzene- d_6) of compound 1.



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Figure S14. ¹H NMR spectrum (500 MHz, pyridine-*d*₅) of (*R*)-MTPA ester of 1 (1b).



Figure S15. The cell cycle of (A) Hep3B, (B) HepG2, (C) PC3 and (D) HeLa cells by flow cytometry treated with **1** to its IC₅₀ and PTX 10 nM. The data are the means \pm D.E. of three independent experiments. Statistical significance was determined by one-way ANOVA followed by Dunnett's test. * *P* < 0.05 compared to the non-treated control.