# **Supplementary Materials**

## **1. General Information**

All chemicals, precatalysts **2–6**, sulfocalixarene **1** and substrate Diethyl diallylmalonate **9** were purchased from Acros<sup>®</sup>, Aldrich<sup>®</sup>, Merck<sup>®</sup> or VWR<sup>®</sup> and used without further purification, unless otherwise specified. All solvents were distilled on the rotary evaporator before appliance. Substrate *N*-Tosyldiallylamine **7** was synthesized according to literature procedure [1]. All NMR spectra were measured at room temperature (298 K) on a BRUKER Avance 400 spectrometer. Chemical shifts ( $\delta$ ) are expressed in ppm and either refers to not-deuterated amount of used solvents [ $\delta_{\text{H}}$  (CDCl<sub>3</sub>) = 7.26,  $\delta_{\text{H}}$  (D<sub>2</sub>O) = 4.79,  $\delta_{\text{H}}$  (MeOD-*d*<sub>4</sub>) = 3.31,  $\delta_{\text{H}}$  (DMSO-*d*<sub>6</sub>) = 2.50] [2]. UV/vis spectra were detected via Cary Varian 60 spectrometer. Catalysis reactions were stirred via microplate shaker IKA<sup>®</sup> MS 3 basic.

## 2. <sup>1</sup>H-NMR Spectra of RCM Substrates and Products



Figure S1. Extract of <sup>1</sup>H-NMR spectra (rt, 400.13 MHz, D<sub>2</sub>O/MeOD- $d_4$ ) of RCM substrate 9. \*1 = MeOH; \*2 = H<sub>2</sub>O.



**Figure S2.** Extract of <sup>1</sup>H-NMR spectra (rt, 400.13 MHz,  $D_2O/MeOD-d_4$ ) of complete conversion of substrate **9** to product **10** using 5 mol % precatalyst **5a**. \*1 = MeOH; \*2 = H<sub>2</sub>O.



Figure S3. Extract of <sup>1</sup>H-NMR spectra (rt, 400.13 MHz, CDCl<sub>3</sub>) of RCM substrate 7.  $*1 = MeOH; *2 = H_2O.$ 



**Figure S4.** Extract of <sup>1</sup>H-NMR spectra (rt, 400.13 MHz, D<sub>2</sub>O/MeOD- $d_4$ ) of RCM of substrate 7 to 91% product **8** using 5 mol % catalyst **4**. \*1 = MeOH; \*2 = H<sub>2</sub>O.

## 3. Spectra of Solubilisation Experiments

Procedure: A mixture of the catalyst **2** (5 mg, 6.08  $\mu$ mol), and a supramolecular additive **1** (7.27 mg, 6.08  $\mu$ mol), when applicable in D<sub>2</sub>O or MeOD (1 mL) as the solvent was stirred at room temperature and a constant stirring rate of 1000 rpm with exclusion of light. After stirring for a certain time, the mixture was analyzed by <sup>1</sup>H-NMR spectroscopy.



Figure S5.  ${}^{31}P{}^{1}H$  NMR measured in D<sub>2</sub>O.



Figure S6.  ${}^{31}P{}^{1}H$  NMR measured in MeOD.



Figure S7. UV/vis spectra of precatalyst 2 and additive 1 in MeOH.



Figure S8. UV/vis spectra of precatalyst 2 and additive 1 in H<sub>2</sub>O.

# 4. Binding Studies

4.1.  $[Cy_3PH]^+[BF_4]^-$  with Sulfocalixarenes 1



 $c_{\text{Rezeptor}}/c_{\text{Substrat}}$ 

**Figure S9.** Binding isotherm of receptor **1a**: Plot  $\Delta \delta_0$  against c<sub>receptor</sub>/c<sub>substrate</sub> (Dots: experimental measured chemical shift; solid line: fit of the experimental data using a 1:1-binding model;  $\Delta \delta = \delta_{\text{receptor/substrate}} - \delta_{\text{substrate}}$ ).



 $c_{\text{receptor}}/c_{\text{substrate}}$ 

**Figure S10.** Binding isotherm of receptor **1b**: Plot  $\Delta \delta_0$  against c<sub>receptor</sub>/c<sub>substrate</sub> (Dots: experimental measured chemical shift; solid line: fit of the experimental data using a 1:1-binding model;  $\Delta \delta = \delta_{\text{receptor/substrate}} - \delta_{\text{substrate}}$ ).



**Figure S11.** Binding isotherm of receptor **1c**: Plot  $\Delta \delta_0$  against creceptor/c<sub>substrate</sub> (Dots: experimental measured chemical shift; solid line: fit of the experimental data using a 1:2-binding model;  $\Delta \delta = \delta_{\text{receptor/substrate}} - \delta_{\text{substrate}}$ ).

#### 4.2. Cy<sub>3</sub>P=O with Sulfocalixarenes **1b**



**Figure S12.** Binding isotherm of receptor **1b** in D<sub>2</sub>O: Plot  $\Delta\delta_0$  against creceptor/c<sub>substrate</sub> (Dots: experimental measured chemical shift; solid line: fit of the experimental data using a 1:1-binding model;  $\Delta\delta = \delta_{\text{receptor/substrate}} - \delta_{\text{substrate}}$ ).

**Table S1.** Association constant, observed chemical shifts and complexation induced chemical shift (CIS) for the binding process of receptor **1b** with  $Cy_3PO$  as guest in  $D_2O$ .

$K_{Ass} = 28.8 \pm 1.15 \text{ M}^{-1}$	$\mathrm{H}^{1}$	$H^2$	H <sup>3</sup>	$H^4$	H <sup>5</sup>	$H^6$
$\delta_0$ (ppm)	1.2688	1.3840	1.7218	1.8109	1.8692	2.0092
$\Delta\delta_0$ (ppm)	-0.1649	-0.1172	-0.2852	-0.1974	-0.1178	-0.0850
$\Delta\delta_{calc}$ (ppm)	-0.5458	-0.3924	-0.9371	-0.6679	-0.4035	-0.2941

 $\Delta \delta_0 = \delta_{RS} - \delta_0$  determined by NMR Titration experiments measured in D<sub>2</sub>O and MeOD as internal standard;  $\Delta \delta_{calc}$  complexation induced shift, calculated by extrapolation with HypNMR.

## 4.3. Substrate 7 with Sulfocalixarenes 1



**Figure S13.** Binding isotherm of receptor **1a** in D<sub>2</sub>O: Plot  $\Delta \delta_0$  against c<sub>receptor</sub>/c<sub>substrate</sub> (Dots: experimental measured chemical shift; solid line: fit of the experimental data using a 1:1-binding model;  $\Delta \delta = \delta_{\text{receptor/substrate}} - \delta_{\text{substrate}}$ ).

$K_{Ass} = 22.4 M^{-1}$	$\mathbf{H}^{1}$	$H^2$	<b>H</b> <sup>3</sup>	$H^4$	H <sup>5</sup>	$H^6$
δ <sub>0</sub> (ppm)	2.254	3.6607	7.2923	7.5918	5.0662	5.5182
$\Delta\delta_0 (ppm)$	-0.1608	0.0673	-0.0223	0.0566	0.0713	0.0802
$\Delta \delta_{\text{calc}}$ (ppm)	-0.6564	0.2466	-0.1071	0.1949	0.2605	0.2989

**Table S2.** Association constant, observed chemical shifts and complexation induced chemical shift (CIS) for the binding process of receptor 1a with 7 as guest in D<sub>2</sub>O.

 $\Delta \delta_0 = \delta_{RS} - \delta_0$  determined by NMR Titration experiments measured in D<sub>2</sub>O and DMSO as internal standard;  $\Delta \delta_{calc}$  complexation induced shift, calculated by extrapolation with HypNMR.



**Figure S14.** Binding isotherm of receptor **1b** in D<sub>2</sub>O: Plot  $\Delta\delta_0$  against creceptor/c<sub>substrate</sub> (Dots: experimental measured chemical shift; solid line: fit of the experimental data using a 1:1-binding model;  $\Delta\delta = \delta_{receptor/substrate} - \delta_{substrate}$ ).

**Table S3.** Association constant, observed chemical shifts, and complexation induced chemical shift (CIS) for the binding process of receptor **1b** with **7** as guest in  $D_2O$ .

$K_{Ass} = 36.3 M^{-1}$	$\mathrm{H}^{1}$	$H^2$	H <sup>3</sup>	$\mathbf{H}^{4}$	H <sup>5</sup>	$H^6$
$\delta_0$ (ppm)	2.2536	3.6531	7.2939	7.5710	5.0664	5.5180
$\Delta\delta_0$ (ppm)	-0.0772	-0.0214	-0.0424	-0.0229	-0.0332	-0.0211
$\Delta \delta_{calc} (ppm)$	-0.3951	-0.1146	-0.2171	-0.0189	-0.1708	-0.1101

 $\Delta \delta_0 = \delta_{RS} - \delta_0$  determined by NMR Titration experiments measured in D<sub>2</sub>O and DMSO as internal standard;  $\Delta \delta_{calc}$  complexation induced shift, calculated by extrapolation with HypNMR.

#### References

- 1. So, C.M.; Kume, S.; Hayashi, T. Rhodium-Catalyzed Asymmetric Hydroarylation of 3-Pyrrolines Giving 3-Arylpyrrolidines: Protonation as a Key Step. J. Am. Chem. Soc. 2013, 135, 10990–10993.
- Fulmer, G.R.; Miller, A.J.M.; Sherden, N.H.; Gottlieb, H.E.; Nudelman, A.; Stoltz, B.M.; Bercaw, J.E.; Goldberg, K.I. NMR chemical shifts of trace impurities: Common laboratory solvents, organics, and gases in deuterated solvents relevant to the organometallic chemist. *Organometallics* 2010, 29, 2176–2179.