Supporting Materials

Synthesis, Hydrolysis, and Protonation-Promoted Intramolecular Reductive Breakdown of Potential NRTIs: Stavudine α-*P*-Borano-γ-*P*-*N*-L-tryptophanyltriphosphates

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Detailed HPLC Chromatograms and NMR Spectra. Example LC-MS Analyses of Degradation Products

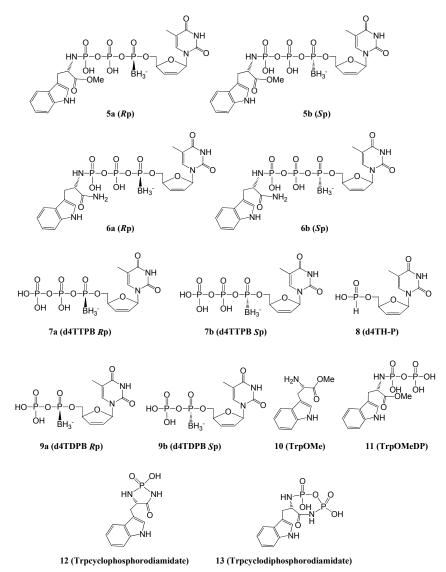


Chart S1. Structures of d4T α -*P*-borano- γ -*P*-*N*-L-tryptophanyltriphosphates and major degradation products in Tris buffer.

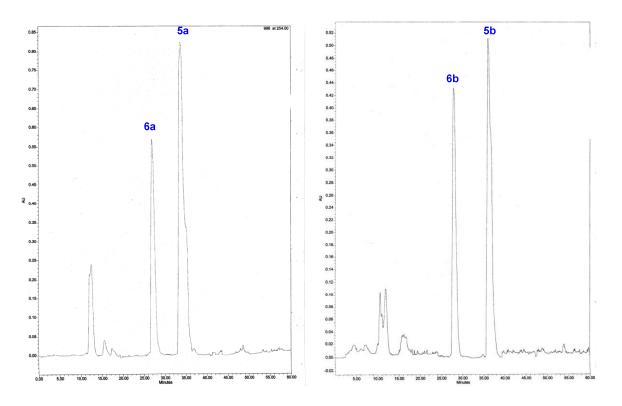


Figure S1. Isolation of **6a** and **6b** using HPLC after 17% NH₄OH treatment of the corresponding isomer of **5** at rt for 6 h. WatersTM (Milford, MA, USA) Delta 600E system (XTerra Prep RP-18 Column, 5 μ m, 10 mm × 150 mm, UV detection), 0%–20% B in 40 min, 20%–90% B for an additional 10 min at 3 mL/min (solvent A: 1% acetonitrile (ACN) in 20 mM TEAA, solvent B: 100% ACN).

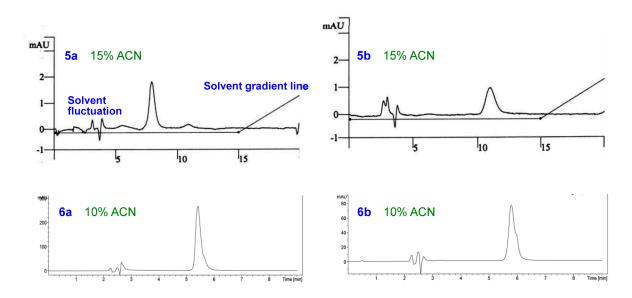


Figure S2. HPLC chromatograms of purified compounds **5a**, **5b**, **6a**, and **6b** at t = 0. Varian (Palo Alto, CA, USA) Star #1, UV detection at 254 nM. Column: Waters Delta-PakTM C18, 15 µm, 3.9 mm × 300 mm. Tested at 1 mL/min with isocratic condition of 15% or 10% ACN in 10 mM TEAA (pH 7) before column cleaning.

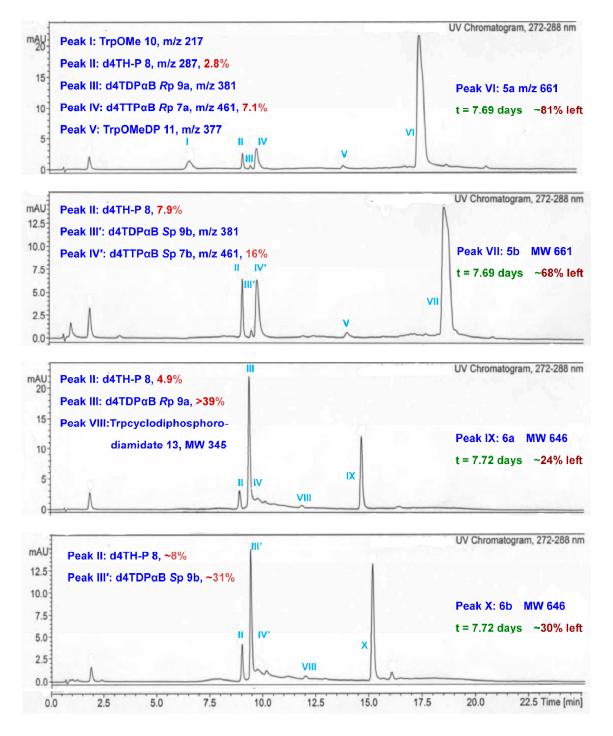


Figure S3. LC-MS analyses of 0.03 mM d4T boranotriphosphate analogs **5a**, **5b**, **6a** and **6b** in 4 mM, pH 7.4 Tris buffer at 37 ± 0.5 °C, after ~7.7 day incubation. LC-MS (Agilent, Santa Clara, CA, USA) condition: Eluted at 0.3 mL/min with 0%–25% B in 25 min (Solvent A: 10 mM pH 7 TEAA; Solvent B: ACN). Column: Eclipse XDB C-18, 2.1 × 50 mm ZORBAX, 3.5 µm. UV detection at 272–288 nm. MS negative ion detection.

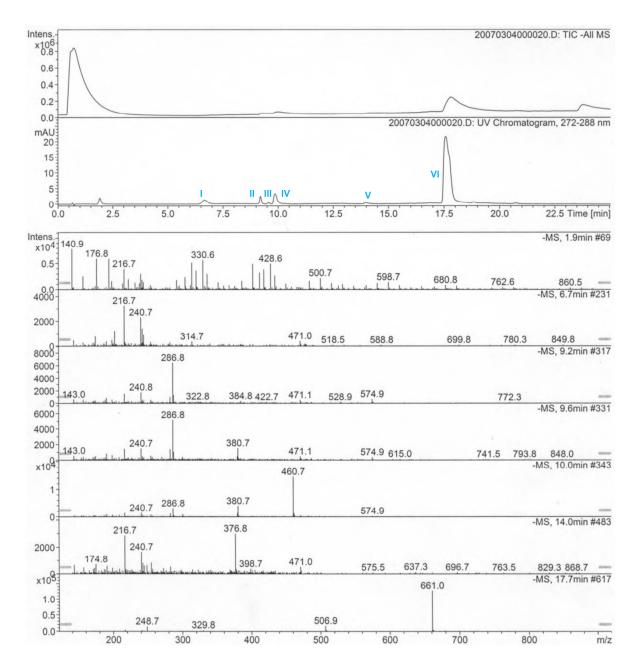


Figure S4. LC-MS analysis of 0.03 mM d4T boranotriphosphate analog **5a** in 4 mM, pH 7.4 Tris buffer at 37 ± 0.5 °C, after ~7.7 day incubation. LC-MS condition: as described in Figure S3.

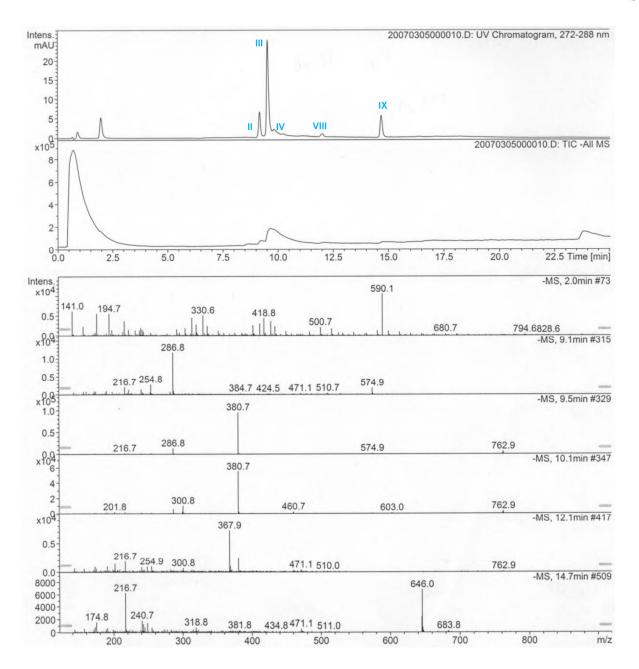


Figure S5. LC-MS analysis of 0.03 mM d4T boranotriphosphate analog **6a** in 4 mM, pH 7.4 Tris buffer at 37 ± 0.5 °C, after 16 day incubation. LC-MS condition: as described in Figure S3.

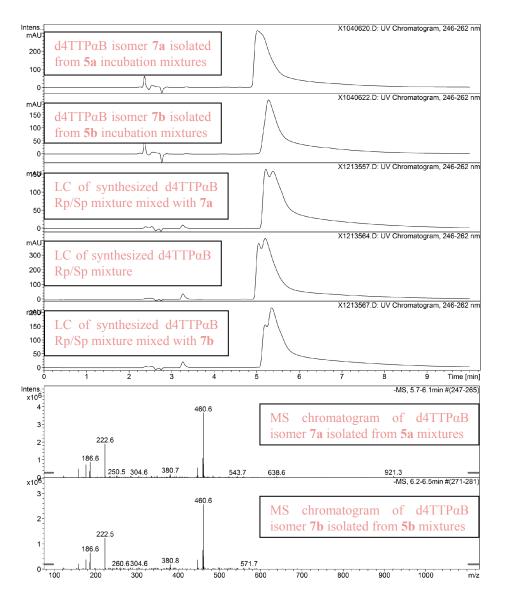


Figure S6. LC-MS verification of stereo specificity of the d4TTPαB isomers degraded from **5a** and **5b** incubation mixtures.

Incubation mixtures were the pools of the respective incubation solutions over 0.5-16 days at 37 ± 0.5 °C. LC-MS condition: Eluted at 40 µL/min with isocratic 8% B (solvent A: 5% ACN in 20 mM TEAA, solvent B: 90% ACN in H₂O). Column: Polaris 3 C-18 A 150×1.0 mm. UV detection at 190–400 nm. MS negative ion detection. The retention times of the two isolated isomers **7a** and **7b** were very close. As the addition of **7a** to the synthesized d4TTP α B *R*p/*S*p mixture (wherein the first and the second eluted peaks were assigned as *R*p and *S*p isomers, respectively) increased the *R*p isomer peak area, whereas **7b** increased the Sp isomer peak area, **7a** was considered to be the *R*p isomer and **7b** would be the *S*p isomer.

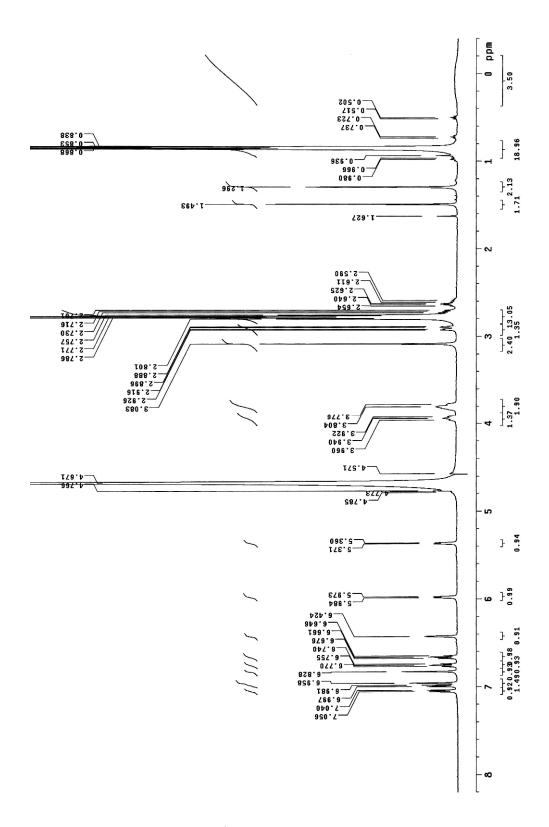


Figure S7. 5a ¹H-NMR in D₂O 500 MHz 2 °C.

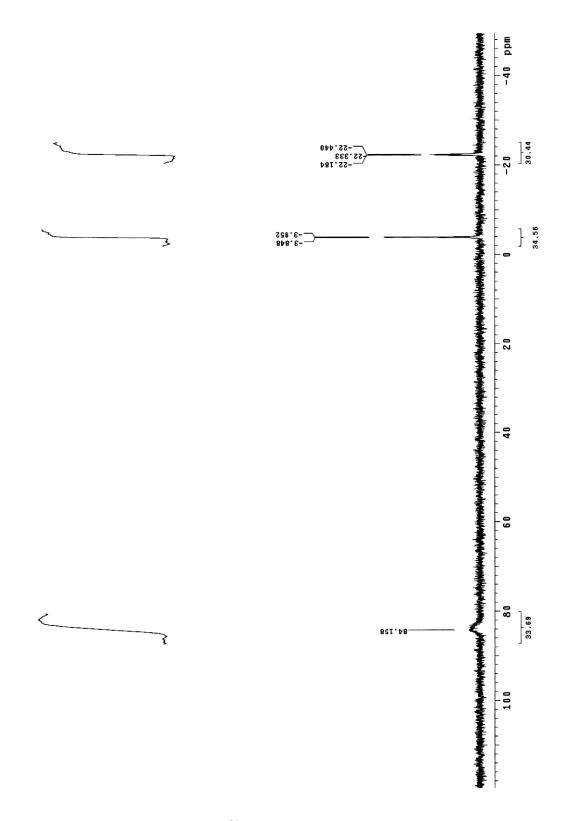


Figure S8. 5a ³¹P-NMR in D₂O 202 MHz 2 °C.

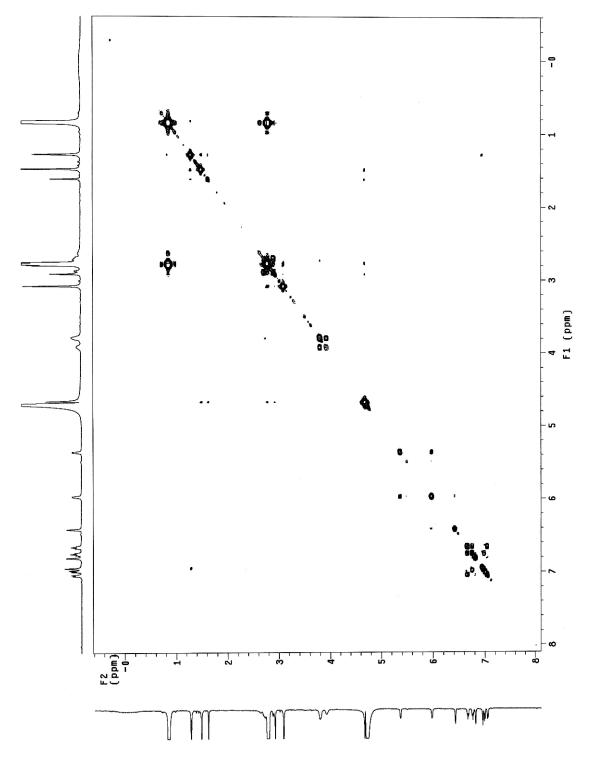


Figure S9. 5a COSY in D₂O 500 MHz 2 °C.

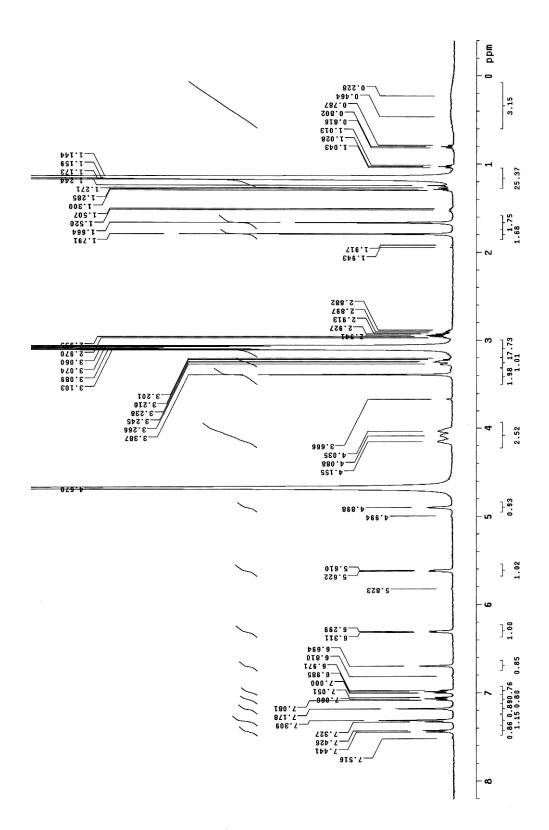


Figure S10. 5b ¹H-NMR in D₂O 500 MHz 25 °C.

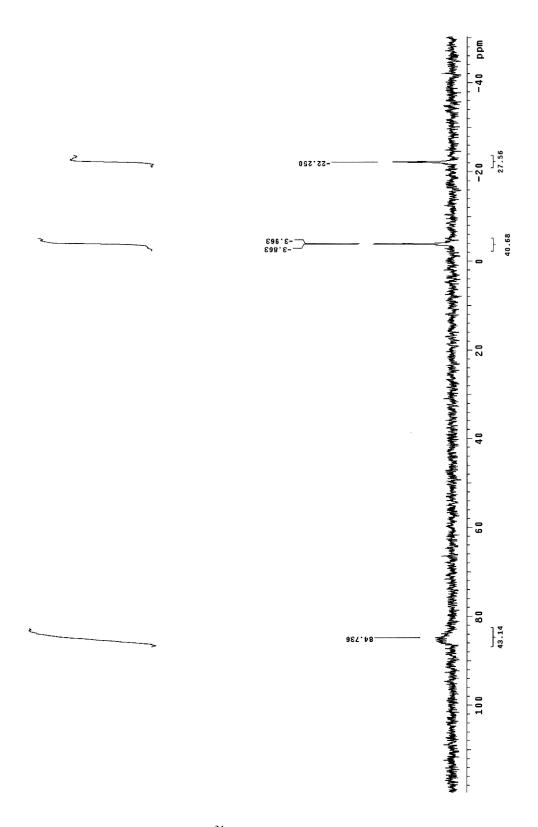


Figure S11. 5b ³¹P-NMR in D₂O 202 MHz 2 °C.

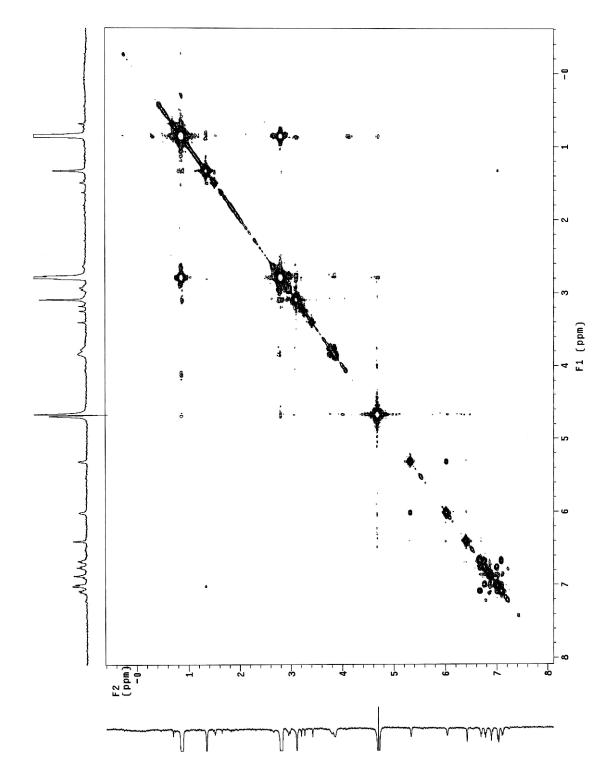


Figure S12. 5b COSY in D₂O 500 MHz 2 °C.

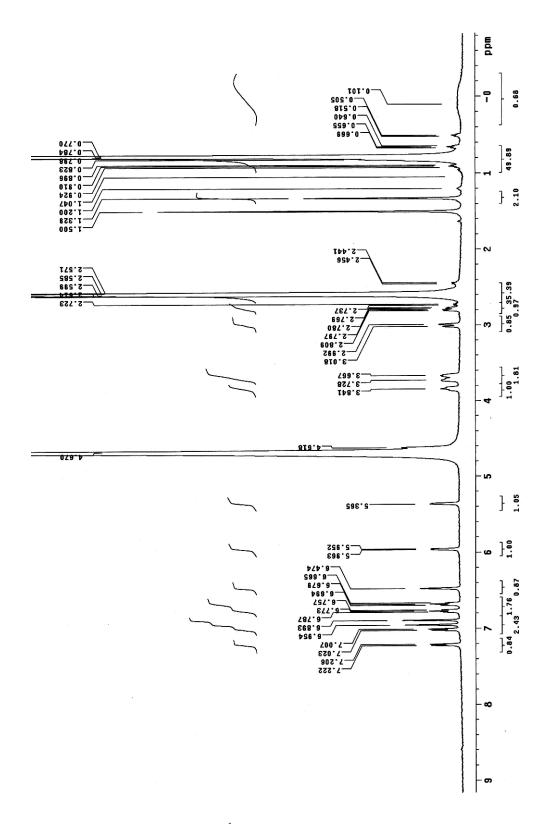


Figure S13. 6a ¹H-NMR in D₂O 500 MHz 2 °C.

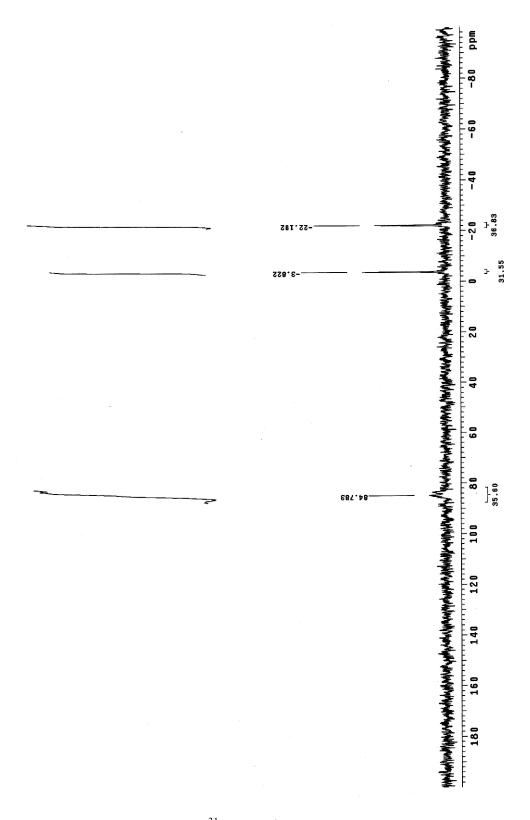


Figure S14. 6a ³¹P-NMR in D₂O 202 MHz 2 °C.

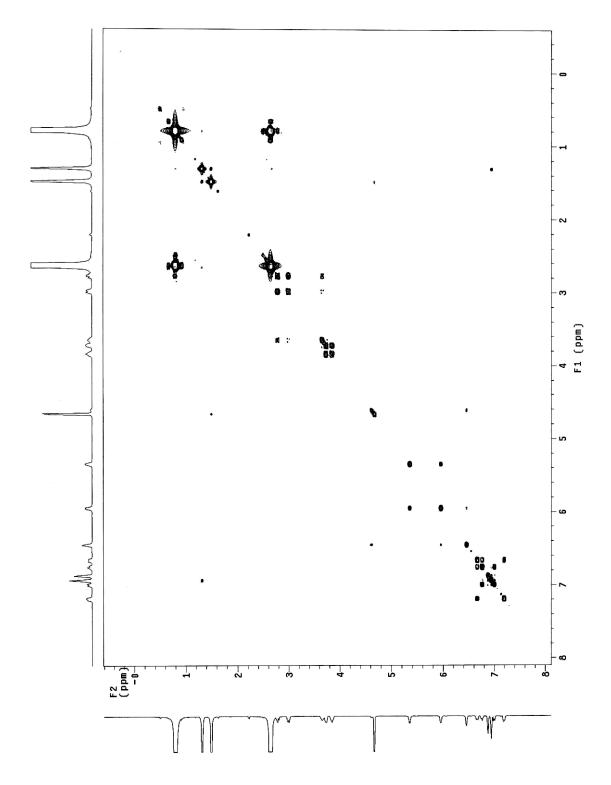


Figure S15. 6a COSY in D₂O 500 MHz 2 °C.

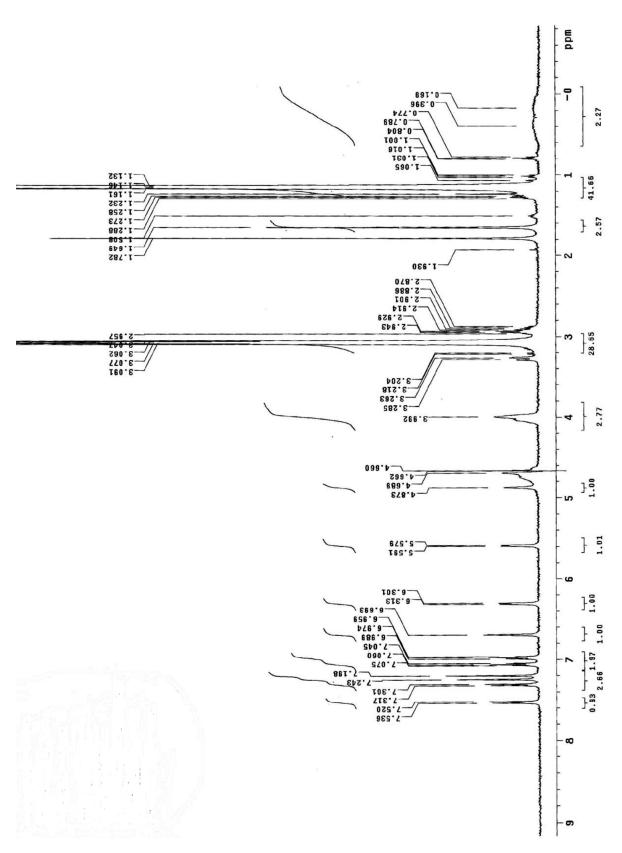


Figure S16. 6b ¹H-NMR in D₂O 500 MHz 25 °C Water Presat.

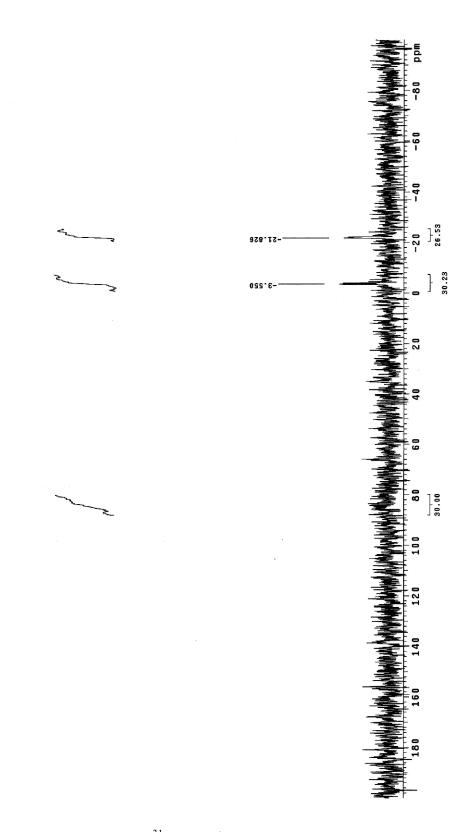
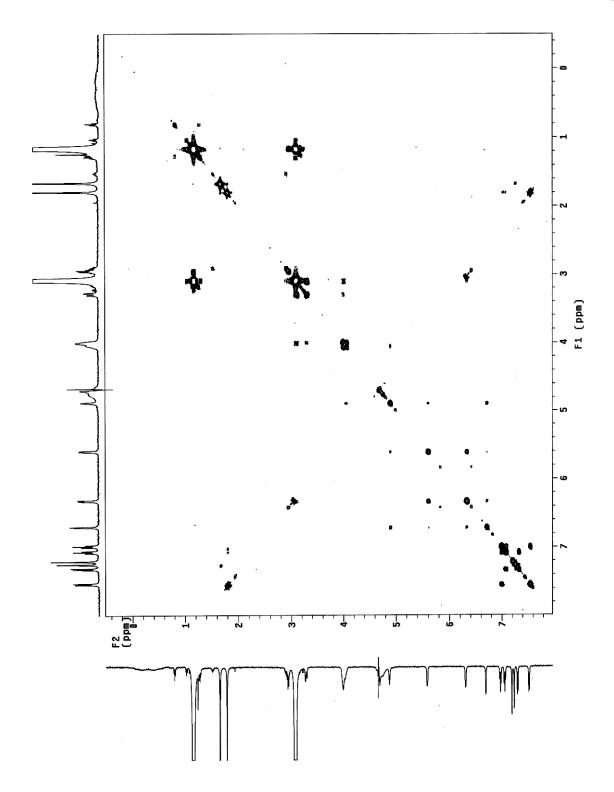
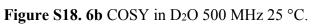


Figure S17. 6b ³¹P-NMR in D₂O 202 MHz 2 °C.





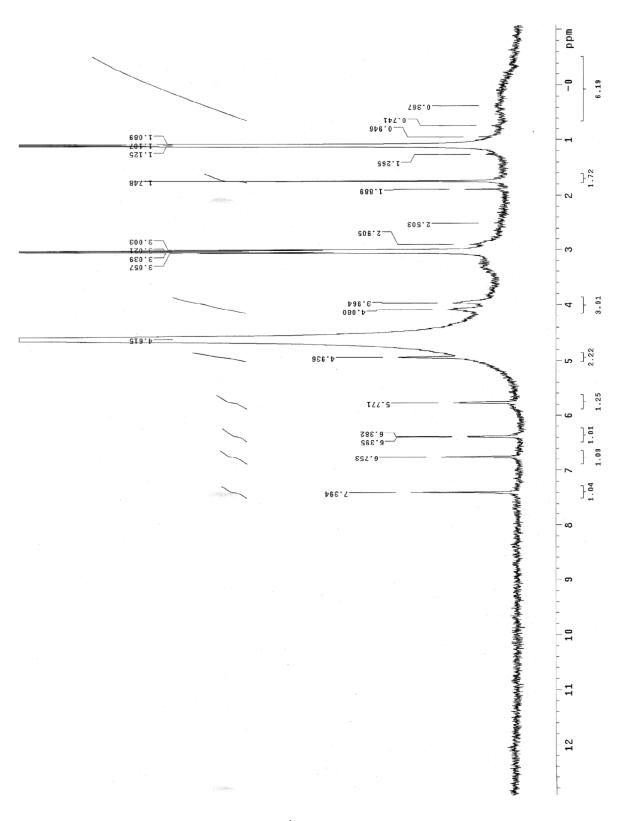


Figure S19. d4TTP α B *R*p ¹H-NMR in D₂O 400 MHz 25 °C.

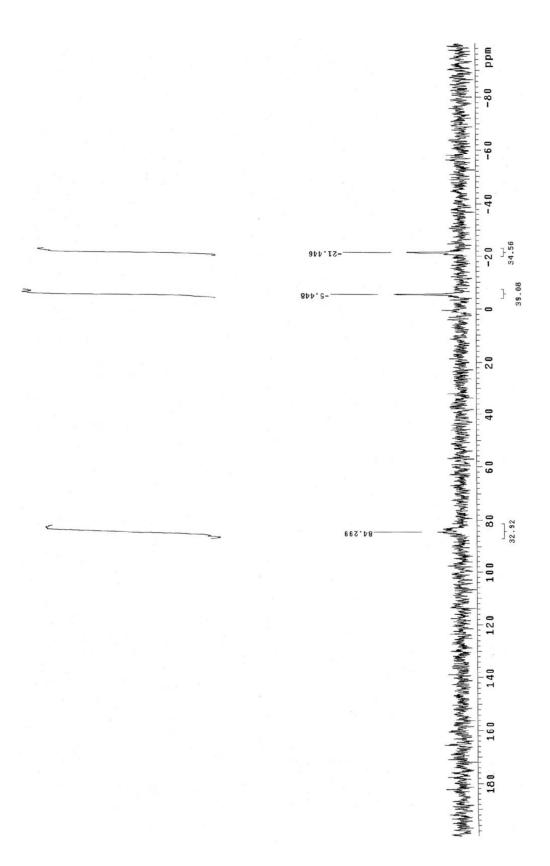


Figure S20. d4TTP α B *R*p ³¹P-NMR in D₂O 202 MHz 25 °C.

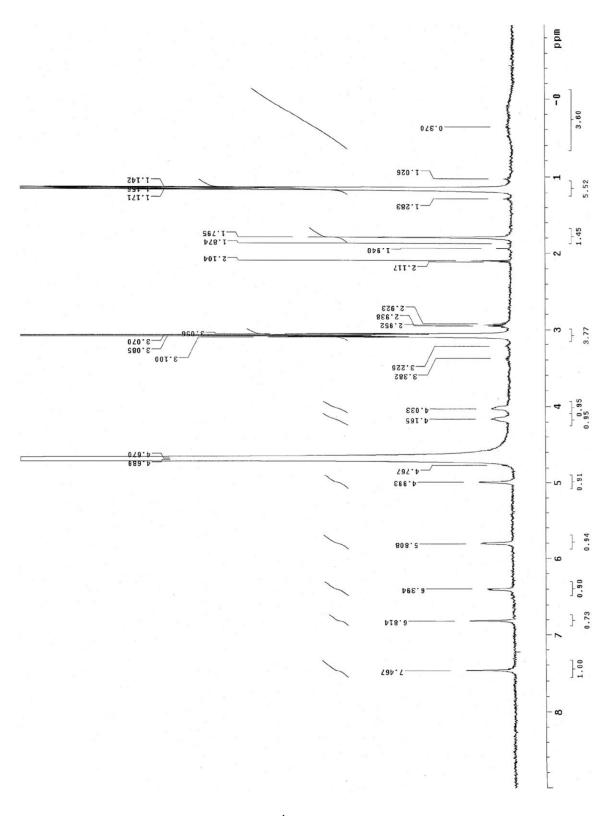


Figure S21. d4TTPαB *S*p ¹H-NMR in D₂O 500 MHz 25 °C.

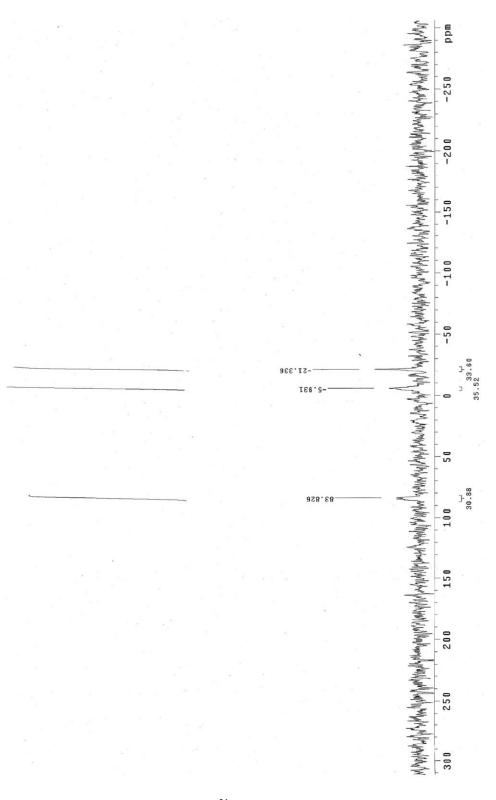


Figure S22. d4TTP α B Sp ³¹P-NMR in D₂O 162 MHz 25 °C.