

Article

On the Nature of the Transition State Characterizing Gated Molecular Encapsulations

Xiaoyong Lu, Bao-Yu Wang, Shigui Chen and Jovica D. Badjić *

Department of Chemistry and Biochemistry, The Ohio State University, 100 West 18th Avenue, Columbus, OH 43210, USA

* Author to whom correspondence should be addressed; E-Mail: badjic@chemistry.ohio-state.edu; Tel.: +1-614-247-8342; Fax: +1-614-292-1685.

Received: 17 July 2014; in revised form: 21 August 2014 / Accepted: 28 August 2014 /

Published: 11 September 2014

Abstract: Gated molecular encapsulations, with baskets of type **1**, are postulated to occur by the mechanism in which solvent molecule penetrates the inner space of **1**, through one of its apertures, while the residing guest simultaneously departs the cavity. In the transition state of the exchange, three pyridine-based gates are proposed to assume an open position with both incoming solvent and departing guest molecules interacting with the concave surface of the host. The More O'Ferrall-Jencks diagram and linear free energy relationships (LFERs) suggest a more advanced departure of the guest when bigger solvents partake in the displacement.

Keywords: molecular encapsulation; molecular gating; host-guest chemistry; linear free energy relationships; encapsulation mechanisms

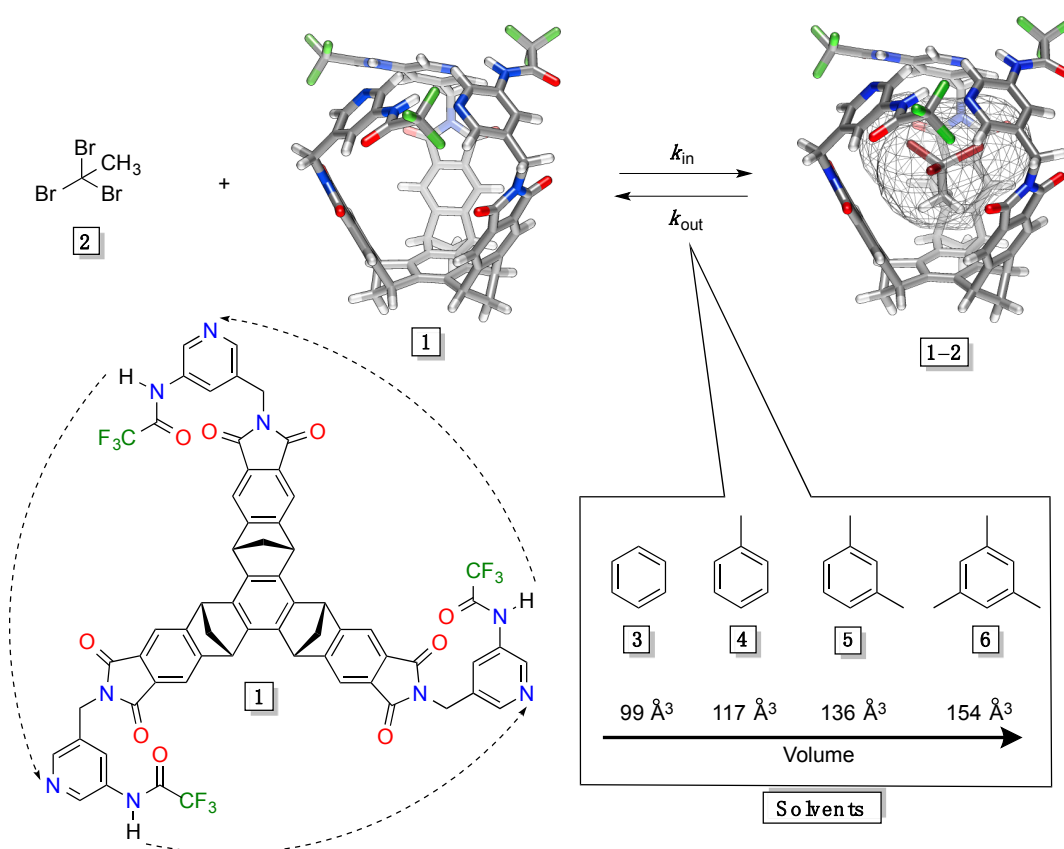
1. Introduction

The process of molecular encapsulation occurs, in concave hosts, via slippage, gating or full dissociation of self-assembled components [1]. Indeed, elucidating the mechanism of the operation of cavitands [2–5] has been an interest for controlling the outcome of chemical reactions [6], stabilizing reactive intermediates [7], or delivering useful molecules [8,9]. Accordingly, for the complexation of metal cations with crown ethers, Schneider and Cox reported that the rates by which cations access (k_{in}) the macrocycles are fast (approaching a diffusion-controlled limit) while their departure from the complex (k_{out}) is much slower corresponding to the thermodynamic stability (ΔG°) of the complex

itself [10]. Importantly, the corresponding mechanistic study [11] indicated an early transition state for the entrapment, resembling reactants to account for the absence of the selectivity in complexation events. For gated molecular encapsulations [12], however, a conformational change in a gated host creates an aperture permitting in/out exchange of guests. The complexation is, for this reason, occurring at a slower rate with the corresponding higher activation barrier referred to as constrictive binding [13]. As a result, one could tune the kinetics of molecular encapsulation by controlling conformational changes of the host [14]. Furthermore, the “opening” of some dynamic hosts, hemicarcerans [15,16] or self-folding capsules [17], were argued to constitute the rate-determining step of encapsulation. In other gating situations [18,19], however, the guest entrance generates enough van der Waals strain so that the encapsulation rate law exhibits a kinetic dependence in the concentration of both host and guest [20]. To our knowledge, there has been no particular study [21] about the nature of transition states characterizing gated encapsulations, and this is the objective of our paper.

Gated molecular baskets [22] have been designed and studied in our laboratory for almost a decade [23]. These hosts comprise a flat aromatic base that is fused to three bicycle[2.2.1]heptane rings forming a curved unit ($V = 220 \text{ \AA}^3$, Figure 1).

Figure 1. Energy-minimized (MMFFs, Spartan) and chemical structures of gated molecular basket **1** ($V = 220 \text{ \AA}^3$). The kinetics of in/out trafficking of guest **2** ($V = 107 \text{ \AA}^3$) to (k_{in}) and from (k_{out}) basket **1** was studied in solvents **3–6** ($99\text{--}154 \text{ \AA}^3$).



Three phthalimides extend this structure into a bowl-shaped platform. Three aminopyridine rings [24], containing hydrogen-bonding donor and acceptor sites, are attached to the rim of the platform to, via CH_2 “hinges”, act as gates that, by controlling the portal size, regulate the in/out encapsulation of

small haloalkanes ($V \sim 88\text{--}120 \text{ \AA}^3$) [25]. The formation of basket/guest complex was, in solvophobic CD_2Cl_2 solvent ($V = 61 \text{ \AA}^3$) [26], found to be first- while the dissociation is zeroth-order in guests (Figure 1) [20]. In essence, the basket undergoes a rapid thermal racemization ($\Delta G_{\text{rac}}^\ddagger < 11 \text{ kcal/mol}$) to, occasionally, permit the entrance/departure of guest molecules [27]. For a series of isosteric guests we found a linear dependence between the host-guest affinities (ΔG°) and the free energies of activation ($\Delta G_{\text{in}}^\ddagger$ and $\Delta G_{\text{out}}^\ddagger$), which was fit to the following equation: $\Delta G_{\text{in/out}}^\ddagger = \rho_{\text{in/out}} \Delta G^\circ + \delta$ [28] On the basis of rather small ρ_{in} values (0.08–0.25), we hypothesized that an early transition state characterizes gated encapsulations [27,28]; this was additionally supported with a relatively poor stereo-selectivity of entrapments ($k_{\text{in}}^{\text{R}}/k_{\text{in}}^{\text{S}} \sim 2\text{--}3$) [29]. To investigate the nature of the transition state of gated molecular encapsulations in greater details, we hereby used dynamic $^1\text{H-NMR}$ spectroscopy to measure rate coefficients (k_{out}) by which 1,1,1-tribromoethane **2** ($V = 107 \text{ \AA}^3$) departs from the interior of basket **1** in four aromatic solvents **3–6** (Figure 1). In earlier studies [20], we surmised that the egress of guest molecule should be followed with the entrapment of solvent (Figure 1). It follows that the rate of such guest swap should change with increasingly bigger solvent molecules ($99\text{--}154 \text{ \AA}^3$, Figure 1) via a steric imposition of the transition state of the transformation [21,30].

2. Results and Discussion

Basket **1** (Figure 1) was prepared by following an optimized synthetic procedure [14]. It was poorly soluble in *m*-xylene- d_{10} (*vide supra*) forming a suspension at 298.0–348.0 K (Figure S1). Upon addition of 1,1,1-tribromoethane **2** (0.8–50 molar equivalents, Figure S4), however, the solid basket **1** was extracted into *m*-xylene- d_{10} to, perhaps, give a [**1–2**] complex. Indeed, the formation of [**1–2**] ensued on the basis of $^1\text{H-NMR}$ spectroscopy and the intensity of signals corresponding to both host and guest (Figure 2A). DOSY NMR spectroscopic measurements (Figures S6–S9) were also in line with the formation of 1:1 complex in each solvent ($r_{\text{H}} \sim 7\text{--}8 \text{ \AA}$, Table 1) [31]; the experimental hydrodynamic radii correspond to the computed size ($d \sim 14 \text{ \AA}$, MMFFs) of [**1–2**] complex. Furthermore, two separate ^1H - singlets corresponding to the guest inside the host ($\delta = -1.34 \text{ ppm}$, Figure 2A) and in bulk solvent ($\delta = 2.81 \text{ ppm}$, Figure 2A) suggested a slow in/out exchange of CH_3CBr_3 occurring at 300.1 K on the “time scale. Accordingly, we completed $^1\text{H}, ^1\text{H-EXSY}$ NMR measurements (400 MHz, 300.1 K) to examine the kinetics characterizing the trafficking of complexed guest [**2**]_{in} into bulk solvent [**2**]_{out} (k_{out} , Figure 2A); note that due to the nature of EXSY experiment [32], the experimentally determined rate coefficient k_{out}^* is pseudo first-order in character (s^{-1} , Table 1) and in line with the rate $v_{\text{out}}^* = k_{\text{out}}^* [\text{1–2}]$. On the basis of the reaction’s stoichiometry, however, the reaction’s rate could be $v_{\text{out}} = k_{\text{out}} [\text{1–2}]$ (Figure 2A) and it follows that $k_{\text{out}}^* = k_{\text{out}}$ as long as the proposed model is valid. Since greater concentrations of guest [**2**] in solution (3.0–22.0 molar equivalents) had no measurable effects on the experimental rate constant k_{out}^* (Figure 2B) we conclude that the process of decomplexation is zeroth-order in 1,1,1-tribromoethane **2** [20]. This, in turn, corroborates the absence of interchange mechanism (Figure 2C) [19], whereby free guests substitute the one of the same kind trapped in the host. Alternatively, a molecule of solvent **5** could approach [**1–2**] to via its sizeable side aperture “push” the encapsulated guest out and thereby give complex [**1–5**] (Figure 2D); note that, in this situation [28], the gates ought to “open” so that the substitution takes place. In line with this mechanistic hypothesis, varying the size of solvent molecules

should have an effect on the rate by which guest **2** departs the basket: more sizeable compounds should create a greater van der Waals strain, while entering or exiting the host (vide infra), to affect the in/out exchange! Accordingly, we measured (^1H , ^1H -EXSY 300.1 K) rate coefficients k_{out} corresponding to 1,1,1-tribromoethane **2** departing from [1–2] complex in three additional solvents: Benzene- d_6 (**3**), toluene- d_8 (**4**) and mesitylene- d_{12} (**6**). The rate coefficients k_{out} were, importantly, found to increase (while k_{in} decreased) for smaller solvents (Table 1). In fact, there exists a linear free energy relationship (LFER) with $\log k_{\text{out}}$ being proportional to the volume of solvent molecules ($R^2 = 0.97$, Figure 3A).

Figure 2. (A) ^1H -NMR spectrum (400 MHz, 300.1 K) of C_3 symmetric basket **1** (0.5 mg) suspended in *m*-xylene- d_{10} and containing guest **2** (6.5 mM); (B) A plot showing experimental rate coefficients $k_{\text{out}} * (\text{s}^{-1})$, obtained from ^1H , ^1H -EXSY NMR (400 MHz) measurements, as a function of the concentration of guest **2** with basket **1** (1.1 mg) in *m*-xylene- d_{10} at 300.1 K; The interchange ((C), MMFFs/Spartan) or solvent-displacement ((D), MMFFs/Spartan) mechanisms of the exchange).

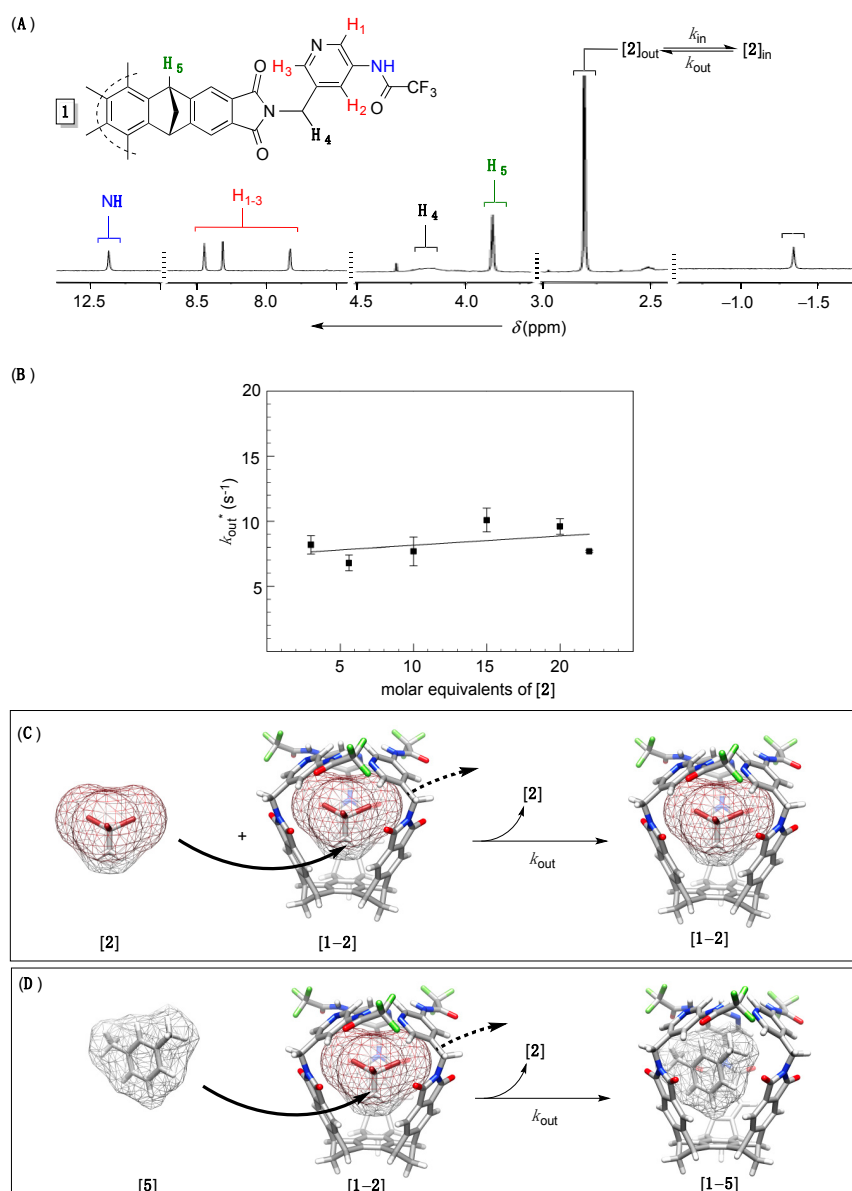
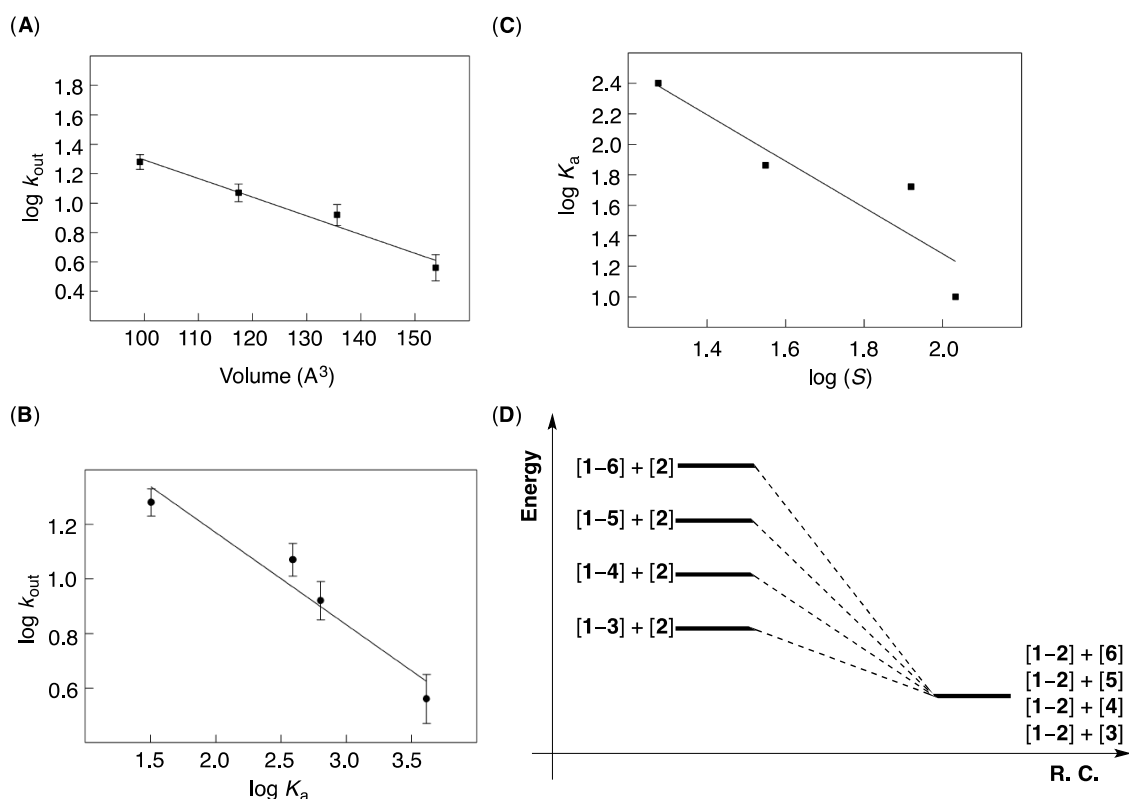


Figure 3. (A) A plot showing $\log k_{\text{out}}$ as function of van der Waals volume (Spartan) of solvents **3–6**. The data was fit to a linear function using the least-square method of analysis ($R^2 = 0.97$, Sigma Plot); (B) A plot showing $\log k_{\text{out}}$ as function of the stability $\log K_a$ of [1–2] complex in solvents **3–6**. The data was fit to a linear function using the least-square method of analysis ($R^2 = 0.94$, Sigma Plot); (C) A plot showing showing $\log K_a$ as function of the solubility $\log S$ of [1–2] complex in solvents **3–6**. The data was fit to a linear function using the least-square method of analysis ($R^2 = 0.94$, Sigma Plot); (D) Reaction coordinate diagram showing the relative stability of [1–solvent] complexes.



To additionally probe the nature of the encapsulation events at hand, we used $^1\text{H-NMR}$ spectroscopy to quantify the affinity (K_a , M^{-1}) of basket **1** for trapping **2** in each solvent (Table 1). The binding was found to be strongest in “large” mesitylene- d_{12} ($V = 154 \text{ \AA}^3$) while weakest in “small” benzene- d_6 ($V = 99 \text{ \AA}^3$) [26]. The measured affinity constants (K_a , Table 1) suggest for the stability of [1–solvent] complexes to be in the order: [1–6] < [1–5] < [1–4] < [1–3] (Figure 3D). In other words, the stability of [1–2] could be comparable in solvents **3–6**, having $\epsilon \sim 2\text{--}4$ (Figure 3D) [33]. To further investigate this point of view, we used $^{19}\text{F-NMR}$ spectroscopy (with C_6F_6 as internal standard, Figure S14) to quantify the solubility (S , Table 1) of basket **1** in solvents **3–6**. The solubility delineates a dynamic equilibrium between basket **1** in its solid state (always the same in any solvent) and as solute in solution and is thus proportional to the stability of [1–solvent] complexes. As $\log(S)$ commensurates with $\log K_a$ (Figure 3C), we conclude that in these two experiments the stability of [1–solvent] complexes primarily determines the state of each equilibrium. Consequently, the stability of [1–2] complex is, in aromatic solvents **3–6**, comparable and therefore shown in Figure 3D as one state. At last, the experimental order of stability of [1–solvent] complexes tracks an increased population of the inner space of basket **1** (approximated PCs are given in Table 1) [26].

Table 1. Diffusion coefficients (D) of [1–2] complex were obtained from DOSY NMR (400 MHz) at 300.1 K and converted into hydrodynamic radii (r_H) using the Stokes-Einstein equation. The exchange rate constants k_{out} (s^{-1}) were obtained from $^1H, ^1H$ -EXSY NMR (300.1 K) and are shown as mean \pm standard deviation (>10 measurements). The equilibrium constants K_a (M^{-1}) were estimated from 1H NMR spectroscopic measurements (via signal integration) at 300.1 K, while k_{in} was calculated as $k_{in} = K_a k_{out}$. Solubility (S) of basket **1**, in **3–6**, was determined with the assistance of ^{19}F NMR spectroscopy at 300.1 K. Packing coefficients (PC) were calculated as PC (%) = $V_{guest}/V_{host's\ inner\ space}$ [26].

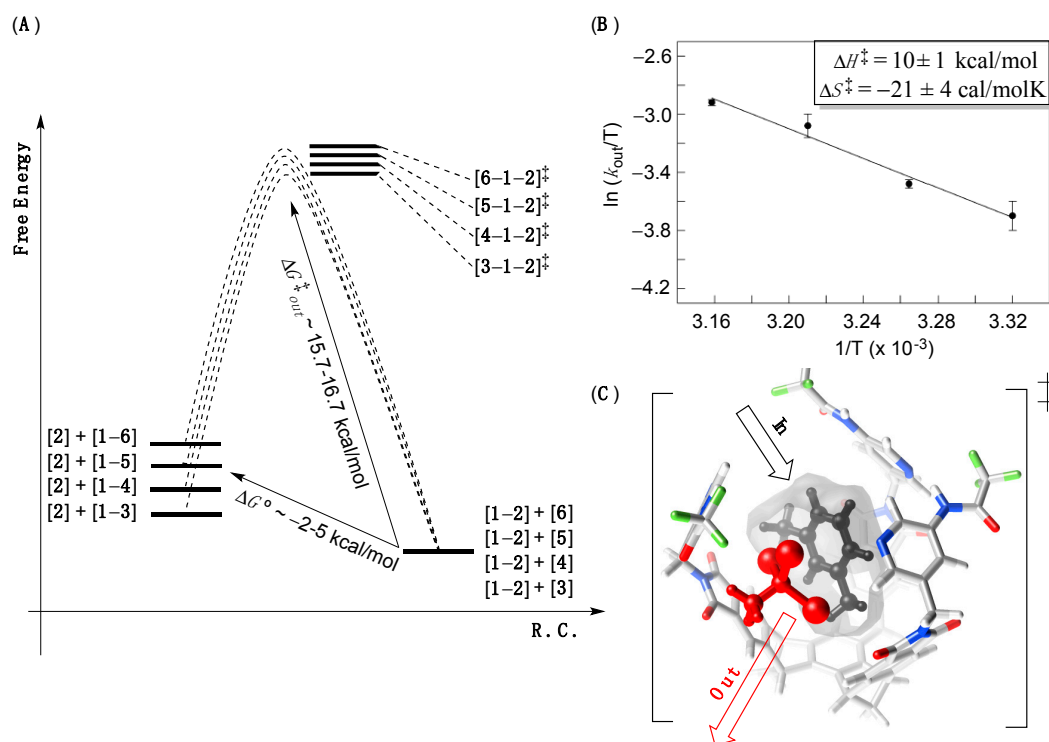
	C₆D₆ (3)	C₆D₅CD₃ (4)	C₆D₄ (CD₃)₂ (5)	C₆D₃ (CD₃)₃ (6)
D ($10^{-10} m^2 s^{-1}$)	4.7 ± 0.1	5.9 ± 0.2	4.47 ± 0.06	4.21 ± 0.03
r_H (Å)	7.7	6.7	8.5	8.0
k_{out} (s^{-1})	19.2 ± 2.2	11.8 ± 1.6	8.3 ± 1.3	3.6 ± 0.8
K_a (M^{-1})	32	390	637	4123
k_{in} ($M^{-1} s^{-1}$)	$[6.1 \pm 0.7] \times 10^2$	$[4.6 \pm 0.6] \times 10^3$	$[5.3 \pm 0.8] \times 10^3$	$[1.5 \pm 0.3] \times 10^4$
S ($\mu mol/dm^3$)	108 ± 11	83 ± 8	35 ± 4	19 ± 2
PC (%)	45	53	62	70

Evidently, more favorable decomplexations of [1–2] (smaller $\log K_a$, Figure 3B) are, in solvents **3–6**, occurring at a faster rate (bigger $\log k_{out}$). The slope of the linear $\log K_a/\log k_{out}$ function (Figure 3B) is equal to 0.3 (Figure 3B), with a greater span of ΔG° ($\Delta\Delta G^\circ \sim 3$ kcal/mol) than ΔG_{out}^\ddagger ($\Delta G_{out}^\ddagger \sim 1$ kcal/mol) values, along the series. On the basis of the transition state theory, ΔG_{out}^\ddagger corresponds to the equilibrium between ([1–2] + [solvent]) and [solvent–1–2] ‡ whereas ΔG° describes the balance between ([1–2] + [solvent]) and ([1–solvent] + [2]). Since the initial states are for both processes the same and comparable in energy (Figure 3D), the observed LFER in Figure 3B must indicate a greater span in the stability of [1–solvent] complexes than the corresponding transition states [solvent–1–2] ‡ (Figure 4A)! To partly account for the observation, we note that the transition state for gated exchange must be an “open” form of the basket with (a) no intramolecular hydrogen bond(s) and (b) somewhat extended surface. Such transient species should “more easily” accommodate differently sized and shaped solvents, than [1–solvent] complex having a “closed” surface, to give rise to a more narrow distribution of ΔG_{out}^\ddagger values.

The energy separation between [solvent–1–2] ‡ and the corresponding ground states is rather large (>10 kcal/mol, Figure 4A) and in line with a unique structure of this fleeting complex lacking N-H-N intramolecular hydrogen bonds [34]. With the assistance of $^1H, ^1H$ -EXSY spectroscopy, we measured rate coefficients k_{out} characterizing the departure of **2** from [1–2] at various temperatures in *m*-xylene- d_{10} (Table S5). From the Eyring plot (Figure 4B), we determined the activation parameters $\Delta H^\ddagger = 10 \pm 1$ kcal/mol and $\Delta S^\ddagger = -21 \pm 4$ kcal/mol K characterizing the process [21]. The negative entropy of activation ($\Delta S^\ddagger < 0$) is indeed in line with an associative mechanism of exchange, resembling classical S_N2 reactions, with simultaneous (but not necessarily synchronous) egress of **2** and ingress of *m*-xylene- d_{10} (Figure 4C)! In spite of the postulated one-step mechanism (Figure 2D), the exchange process may also be a two-step process including rapid opening of the basket’s one gate (first step) followed by a slower swapping of the entrapped with the incoming compounds and the simultaneous opening of two additional gates (second step). It is indeed difficult to repudiate this particular mechanistic scenario, although we have not observed any intermediate either spectroscopically or

kinetically (the saturation kinetics) [18] to suggest the two-step process [23]. In addition, an earlier study [26] indicated that for a guest populating the basket's inner space to a greater extent ($PC > 0.3$, Table 1) the basket's racemization necessitates simultaneous opening of all three gates.

Figure 4. (A) A reaction coordinate diagram depicting kinetic and thermodynamic characteristics of the encapsulation; (B) The Eyring plot showing $\log k_{\text{out}}/T$ as function of $1/T$ corresponding to the decomplexation of [1–2] complex in *m*-xylene- d_{10} . The data was fit to a linear function using the least-square method of analysis ($R^2 = 0.98$, SigmaPlot); (C) The energy minimized [5–1–2] ‡ complex (MMFFs, Spartan) showing *m*-xylene- d_{10} entering while 1,1,1-tribromoethane **2** exiting the basket's inner space.

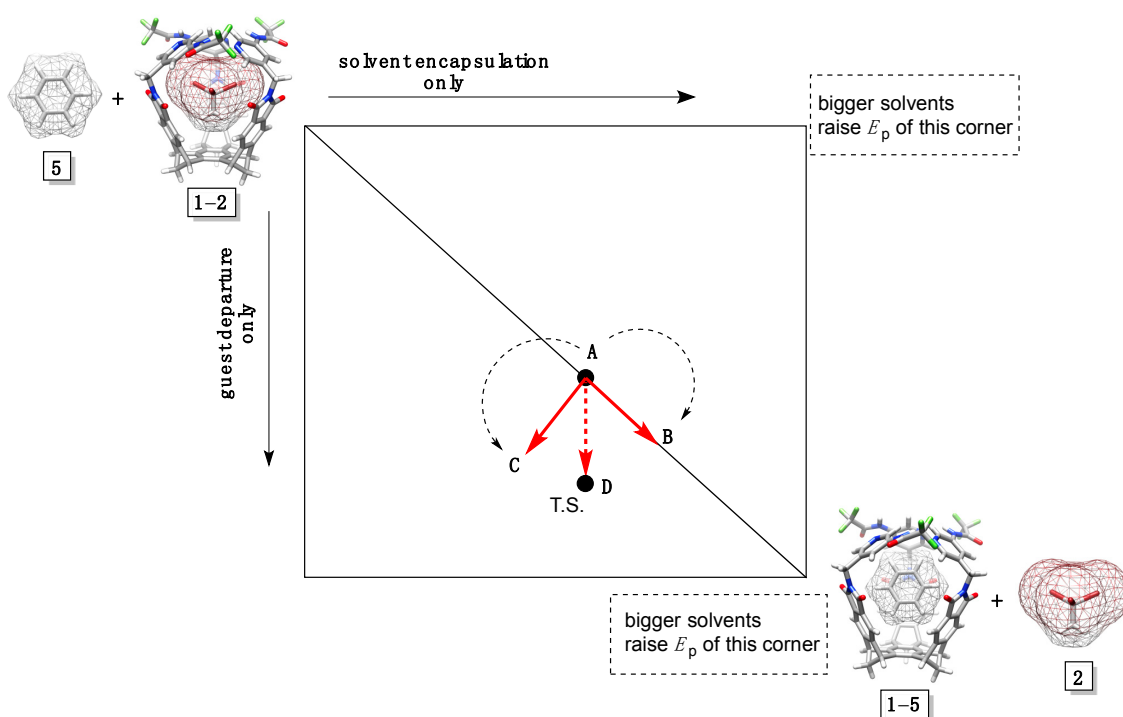


To assess possible changes in the structure of the transition states for the postulated displacement mechanism (Figure 2D), we used the More O'Ferrall-Jencks diagram (Figure 5) [35]. Three reaction coordinates, as in classical nucleophilic substitutions [36], are included: (a) x-axis presents the solvent encapsulation without guest departure; (b) y-axis depicts the sole departure of guest (akin to S_N1); and (c) the diagonal coordinate is a combination of the two pathways (akin to S_N2). The movement of the transition state, with a change in the potential energy of the system (z-axis), proceeds along the diagonal reaction coordinate (Hammond effect) but, in accord with this approach, should also be balanced with a shift perpendicular to it (anti-Hammond effect) [36].

Bulkier solvent molecules form less stable complexes with basket **1** so that the top and bottom right corners are, in Figure 5, raised in the series **3–6**. Accordingly, the transition state shifts along AB and AC vectors to give rise to the AD vector. It follows that with an increase in the solvent size, the degree of solvent encapsulation stays constant while the magnitude of the guest departure increases. To put it in the language of classical organic chemistry, bulkier solvents enforce “ S_N1 -like” mechanism of the supramolecular substitution: More sizeable mesitylene- d_{12} pushes the departing 1,1,1-tribromoethane **2**

to a greater degree than benzene-d₆ in the transition state [36,37]. The analysis is reasonable since bigger mesitylene-d₁₂ provides less space than smaller benzene-d₆ for departing compound **2** within the limited inner space of basket **1**. At last, the LFER in Figure 3A shows a linear dependence for the rate of the guest departure ($\log k_{\text{out}}$) as a function of the size (“nucleophilicity”) of solvents **3–6** thereby supporting the More O’Ferrall-Jencks analysis in Figure 5. In fact, this plot is analogous to Brønsted relationships [37] ($\log k = \beta_{\text{Nuc}} \cdot \text{p}K_{\text{a}} + n$) that are used for characterizing the degree of covalent bond formation (β_{Nuc} ; which is the degree of solvent’s encapsulation in our case) in the transition state of substitution reactions as a function of nucleophilicity [35].

Figure 5. A More O’Ferrall-Jencks plot for solvents **3–6** (only **5** is shown) displacing 1,1,1-tribromoethane **2** in basket **1**.



3. Experimental Section

All chemicals were purchased from commercial sources, and used as received unless stated otherwise. Solvents **3**, **4** and **6** were purchased from Cambridge Isotope Laboratories: benzene-d₆ (D-99.5%), toluene-d₈ (D-99.5%) and mesitylene-d₁₂ (D-98%). *m*-Xylene-d₁₀ (D-98%) was obtained from Sigma-Aldrich. All solvents (used in synthesis, see Scheme in supporting information) were dried prior to use according to standard literature protocols. Chromatography purifications were performed using silica gel 60 (Sorbent Technologies 40–75 μm , 200 \times 400 mesh). Thin-layer chromatography (TLC) was performed on silica-gel plate w/UV254 (200 μm). ¹H and ¹³C-NMR spectra were recorded (400 and 100 MHz, respectively), on a Bruker DPX-400 spectrometer unless otherwise noted. ¹⁹F NMR spectra were performed (376.54 MHz) on Bruker Biospin spectrometer with hexafluorobenzene (C₆F₆) as internal standard. All spectra were referenced using the solvent residual signal. The chemical shift values are expressed as δ (ppm) values. Coupling patterns are abbreviated as s (singlet), d (doublet), t (triplet), m (multiplet). Temperatures for all NMR measurements were

calibrated with neat methanol as a standard. High-resolution mass spectrometric measurements were completed on Bruker Micromass Q-TOF II spectrometer.

Compound **S2** (see Scheme S1 in SI): A solution of bromine (0.52 g, 3.3 mmol) in CCl_4 (7 mL) was added to a solution of compound **S1** (1.0 g, 3.0 mmol) in CCl_4 (20 mL) over 5 min at 77 °C. Following, the solvent was removed under reduced pressure to give brown oil. The oil was loaded on a thin pad of silica (SiO_2), and then washed with dichloromethane (100 mL). The solvent was removed under reduced pressure to give compound **S2** as a light yellow solid (1.5 g, 99% yield; 2:1 mixture of two diastereomers). ^1H NMR (400 MHz, CDCl_3): δ (ppm) = 7.76–7.61 (m, 2 H, ArH), 5.33 (d, J = 4.0 Hz, 0.3 H, CH), 4.45 (d, J = 2.8 Hz, 0.7 H, CH), 3.93 (s, 6 H, $2 \times \text{CH}_3$), 2.83 (m, 0.7 H, CH_2), 2.72 (d, J = 8.1 Hz, 0.3 H, CH_2), 2.38 (m, 0.3 H, CH_2), 2.28 (m, 0.7 H, CH_2). ^{13}C NMR (100.62 MHz, CDCl_3): δ (ppm) = 168.05, 168.01, 167.98, 167.68, 147.63, 146.94, 146.67, 145.67, 132.60, 131.50, 131.30, 131.27, 125.47, 125.14, 124.87, 122.05, 67.27, 66.90, 65.07, 63.69, 63.25, 63.11, 55.42, 52.94, 52.93, 52.88, 52.44, 47.23, 46.78. HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{13}\text{Br}_3\text{O}_4\text{Na}$ $[\text{M}+\text{Na}]^+$ 516.8256, found 516.8272.

Compound **S3** (see Scheme S1 in SI): A solution of potassium *tert*-butoxide (0.65 g, 1.3 mmol) in dry THF (20 mL) was added, by a syringe pump (3 h), to a stirred solution of compound **S2** (0.22 g, 2.0 mmol) in dry THF (100 mL) at 0 °C. After the addition was complete, the reaction mixture was stirred for additional 2 h at 0 °C. Following, the reaction mixture was quenched with cold acidic water (HCl aq), and the solvent evaporated under reduced pressure. The solid residue was partitioned between water (100 mL) and ethyl acetate (3×80 mL). Combined organic layers were dried over Na_2SO_4 and then evaporated under reduced pressure. The solid residue was purified by column chromatography (SiO_2 , hexane/ethyl acetate = 3:1) to yield compound **S3** as a white solid (0.43 g, 80% yield); m.p. 110–111 °C. ^1H -NMR (400 MHz, CDCl_3): δ (ppm) = 7.65 (s, 2 H, $2 \times \text{CH}$), 4.00 (m, 2 H, $2 \times \text{CH}$), 3.90 (s, 6 H, $2 \times \text{CH}_3$), 2.79 (m, 1 H, CH_2), 2.42 (m, 1 H, CH_2). ^{13}C NMR (100.62 MHz, CDCl_3): δ (ppm) = 166.78, 150.38, 132.01, 128.90, 120.85, 65.69, 57.30, 51.40. HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{12}\text{Br}_2\text{O}_4\text{Na}$ $[\text{M}+\text{Na}]^+$ 438.8975, found 438.8972.

Compound **S4** (see Scheme S1 in SI): A solid mixture of $\text{Pd}(\text{OAc})_2$ (57 mg, 0.25 mmol), Ph_3P (0.13 g, 0.51 mmol), Bu_4NBr (1.7 g, 5.1 mmol), K_2CO_3 (3.5 g, 25.4 mmol) and 4Å molecular sieves (4.7 g) was added to a solution of dry dioxane (100 mL) under an atmosphere of argon. Compound **S3** (0.53 g, 1.3 mmol) was added and the reaction mixture stirred at 100 °C for additional 48 h. After, the reaction suspension was cooled to a room temperature and filtered with a filter paper. The solid residue was washed with ethyl acetate (~150 mL), and the organic solvent evaporated under reduced pressure to yield a crude solid residue. The residue was partitioned between water (100 mL) and ethyl acetate (3×80 mL). Combined organic layers were evaporated under vacuum, and the solid residue purified by column chromatography (SiO_2 , hexane/ethyl acetate = 2:1) to yield compound **S4** (59 mg, 20% yield); note that *anti* stereoisomer of **S4** was also isolated (60 mg, 20% yield). ^1H -NMR (400 MHz, CDCl_3): δ (ppm) = 7.43 (s, 6 H, $6 \times \text{CH}$), 4.41 (s, 6 H, $6 \times \text{CH}$), 3.78 (s, 18 H, $6 \times \text{CH}_3$), 2.52 (m, 6 H, $3 \times \text{CH}_2$). ^1H -NMR of *anti* diastereomer of **S4** (400 MHz, CDCl_3): δ (ppm) = 7.64 (s, 2 H, $2 \times \text{CH}$), 7.54 (s, 2 H, $2 \times \text{CH}$), 7.51 (s, 2 H, $2 \times \text{CH}$), 4.40 (s, 2 H, $2 \times \text{CH}$), 4.37, (s, 2 H, $2 \times \text{CH}$), 4.36 (s, 2 H, $2 \times \text{CH}$), 3.81–3.90 (s, 18 H, $2 \times \text{CH}_3$), 2.48 (m, 2 H, CH_2), 2.44 (m, 1 H, CH_2), 2.37 (m, 2 H, CH_2), 2.24 (m, 1 H, CH_2).

The supplementary information includes: ^1H -NMR Binding Studies, DOSY NMR/ ^1H , ^1H -EXSY spectroscopic measurements, variable temperature NMR measurements and ^{19}F -NMR quantification of the solubility of basket **1**.

4. Conclusions

A fundamental understanding of encapsulation processes mediated by dynamic hosts is important for controlling the outcome of chemical reactions [38], resolution of enantiomers by gating [29], and completing a precise delivery of molecules [39,40]. The results of our study contribute to such efforts. In particular, the experimental data suggest that the transition state corresponding to the rate-determining step of gated molecular encapsulations encompasses a conglomerate of the host and two molecules exchanging. In particular, the solvent molecule “pushes” a guest, trapped in the gated basket, to give rise to a transient structure containing both species within the concave surface of the unfolded basket. Importantly, bigger solvents are deduce to enforce “S_N1-like” mechanism [41] of the exchange by advancing the degree of the guest departure.

Supplementary Mterials

Supplementary materials can be accessed at: <http://www.mdpi.com/1420-3049/19/9/14292/s1>.

Acknowledgments

This work was financially supported with funds obtained from the National Science Foundation under CHE-1305179.

Author Contributions

Xiaoyong Lu, Bao-Yu Wang and Shigui Chen completed all experimental measurement. Jovica D. Badjic wrote the paper.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Rieth, S.; Hermann, K.; Wang, B.-Y.; Badjic, J.D. Controlling the dynamics of molecular encapsulation and gating. *Chem. Soc. Rev.* **2011**, *40*, 1609–1622.
2. Palmer, L.C.; Rebek, J., Jr. The ins and outs of molecular encapsulation. *Org. Biomol. Chem.* **2004**, *2*, 3051–3059.
3. Davis, A.V.; Fiedler, D.; Seeber, G.; Zahl, A.; van Eldik, R.; Raymond, K.N. Guest exchange dynamics in an M₄L₆ tetrahedral host. *J. Am. Chem. Soc.* **2006**, *128*, 1324–1333.
4. Marquez, C.; Hudgins, R.R.; Nau, W.M. Mechanism of host-guest complexation by cucurbituril. *J. Am. Chem. Soc.* **2004**, *126*, 5806–5816.
5. Ferrand, Y.; Chandramouli, N.; Kendhale, A.M.; Aube, C.; Kauffmann, B.; Grelard, A.; Laguerre, M.; Dubreuil, D.; Huc, I. Long-range effects on the capture and release of a chiral guest by a helical molecular capsule. *J. Am. Chem. Soc.* **2012**, *134*, 11282–11288.
6. Liu, S.; Gan, H.; Hermann, A.T.; Rick, S.W.; Gibb, B.C. Kinetic resolution of constitutional isomers controlled by selective protection inside a supramolecular nanocapsule. *Nat. Chem.* **2010**, *2*, 847–852.

7. Lin, Z.; Sun, J.; Efremovska, B.; Warmuth, R. Assembly of water-soluble, dynamic, covalent, container molecules and their application in the room-temperature stabilization of protoadamantene. *Chem.-A Eur. J.* **2012**, *18*, 12864–12872.
8. Cao, L.; Hettiarachchi, G.; Briken, V.; Isaacs, L. Cucurbit [7] uril containers for targeted Delivery of oxaliplatin to cancer cells. *Angew. Chem. Int. Ed.* **2013**, *52*, 12033–12037.
9. Helgeson, R.C.; Hayden, A.E.; Houk, K.N. Controlled gating of a hemicarcerand by disulfide-dithiol interchange. *J. Org. Chem.* **2010**, *75*, 570–575.
10. Cox, B.G.; Schneider, H. Kinetics and mechanism of macrocyclic complex formation. *Pure Appl. Chem.* **1990**, *62*, 2259–2268.
11. Cox, B.G.; Garcia-Rosas, J.; Schneider, H. Solvent dependence of the kinetics of formation and dissociation of cryptate complexes. *J. Am. Chem. Soc.* **1981**, *103*, 1054–1059.
12. Houk, K.N.; Nakamura, K.; Sheu, C.; Keating, A.E. Gating as a control element in constrictive binding and guest release by hemicarcerands. *Science* **1996**, *273*, 627–629.
13. Cram, D.J.; Tanner, M.E.; Knobler, C.B. Host-guest complexation. 58. Guest release and capture by hemicarcerands introduces the phenomenon of constrictive binding. *J. Am. Chem. Soc.* **1991**, *113*, 7717–7727.
14. Wang, B.-Y.; Rieth, S.; Badjic, J.D. Tuning the rate of molecular translocation. *J. Am. Chem. Soc.* **2009**, *131*, 7250–7252.
15. Sheu, C.; Houk, K.N. The Role of gating in constrictive binding: molecular mechanics and statistical thermodynamics studies of complexes of a flexible hemicarcerand with neutral guests. *J. Am. Chem. Soc.* **1996**, *118*, 8056–8070.
16. Liu, Y.; Warmuth, R. Isotope effects as probe of constrictive and intrinsic binding in hemicarceplexes. *Org. Lett.* **2007**, *9*, 2883–2886.
17. Rudkevich, D.M.; Hilmersson, G.; Rebek, J., Jr. Self-folding cavitands. *J. Am. Chem. Soc.* **1998**, *120*, 12216–12225.
18. Santamaria, J.; Martin, T.; Hilmersson, G.; Craig, S.L.; Rebek, J., Jr. Guest exchange in an encapsulation complex: A supramolecular substitution reaction. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 8344–8347.
19. Craig, S.L.; Lin, S.; Chen, J.; Rebek, J., Jr. An NMR study of the rates of single-molecule exchange in a cylindrical host capsule. *J. Am. Chem. Soc.* **2002**, *124*, 8780–8781.
20. Hermann, K.; Rieth, S.; Taha, H.A.; Wang, B.-Y.; Hadad, C.M.; Badjic, J.D. On the mechanism of action of gated molecular baskets: The synchronicity of the revolving motion of gates and in/out trafficking of guests. *Beilstein J. Org. Chem.* **2012**, *8*, 90–99.
21. Yoon, J.; Cram, D.J. Decomplexation rate comparisons of hemicarceplexes whose single unique host bridge is changed in length and blocking power. *Chem. Commun.* **1997**, 1505–1506, doi:10.1039/a702380d.
22. Maslak, V.; Yan, Z.; Xia, S.; Gallucci, J.; Hadad, C.M.; Badjic, J.D. Design, synthesis, and conformational dynamics of a gated molecular basket. *J. Am. Chem. Soc.* **2006**, *128*, 5887–5894.
23. Hermann, K.; Ruan, Y.; Hardin, A.; Hadad, C.M.; Badjic, J.D. Gated molecular baskets. *Chem. Soc. Rev.* **2014**, doi:10.1039/c4cs00140k.
24. Wang, B.-Y.; Bao, X.; Yan, Z.; Maslak, V.; Hadad, C.M.; Badjic, J.D. A 3-fold “Butterfly Valve” in command of the encapsulation’s kinetic stability. Molecular baskets at work. *J. Am. Chem. Soc.* **2008**, *130*, 15127–15133.

25. Wang, B.-Y.; Bao, X.; Stojanovic, S.; Hadad, C.M.; Badjic, J.D. Encapsulation of guests within a gated molecular basket: Thermodynamics and selectivity. *Org. Lett.* **2008**, *10*, 5361–5364.
26. Ruan, Y.; Wang, B.-Y.; Erb, J.M.; Chen, S.; Hadad, C.M.; Badjic, J.D. On the role of guests in enforcing the mechanism of action of gated baskets. *Orga. Biomol. Chem.* **2013**, *11*, 7667–7675.
27. Rieth, S.; Badjic, J.D. The effect of the dynamics of revolving gates on the kinetics of molecular encapsulation—the activity/selectivity relationship. *Chem.-A Eur. J.* **2011**, *17*, 2562–2565.
28. Rieth, S.; Bao, X.; Wang, B.-Y.; Hadad, C.M.; Badjic, J.D. Gated molecular recognition and dynamic discrimination of guests. *J. Am. Chem. Soc.* **2010**, *132*, 773–776.
29. Wang, B.-Y.; Stojanovic, S.; Turner, D.A.; Young, T.L.; Hadad, C.M.; Badjic, J.D. The entrapment of chiral guests with gated baskets: Can a kinetic discrimination of enantiomers be governed through gating? *Chem.-A Eur. J.* **2013**, *19*, 4767–4775.
30. Chapman, K.T.; Still, W.C. A remarkable effect of solvent size on the stability of a molecular complex. *J. Am. Chem. Soc.* **1989**, *111*, 3075–3077.
31. Ruan, Y.; Dalkilic, E.; Peterson, P.W.; Pandit, A.; Dastan, A.; Brown, J.D.; Polen, S.M.; Hadad, C.M.; Badjic, J.D. Trapping of organophosphorus chemical nerve agents in water with amino acid functionalized baskets. *Chem.-A Eur. J.* **2014**, *20*, 4251–4256.
32. Perrin, C.L.; Dwyer, T.J. Application of two-dimensional NMR to kinetics of chemical exchange. *Chem. Rev.* **1990**, *90*, 935–967.
33. Lide, D.R. *CRC Handbook of Chemistry and Physics*, 83rd ed.; CRC Press: Boca Raton, FL, USA, 2002.
34. Hammond, G.S. A correlation of reaction rates. *J. Am. Chem. Soc.* **1955**, *77*, 334–338.
35. Anslyn, E.V.; Dougherty, D.A. *Modern Physical Organic Chemistry*; University Science Books: Sausalito, CA, USA, 2006.
36. Harris, J.M.; Shafer, S.G.; Moffatt, J.R.; Becker, A.R. Prediction of S_N2 transition state variation by the use of More O’Ferrall plots. *J. Am. Chem. Soc.* **1979**, *101*, 3295–3300.
37. Bordwell, F.; Hughes, D.L. S_N2 reactions of nitranions with benzyl chlorides. *J. Am. Chem. Soc.* **1984**, *106*, 3234–3240.
38. Wang, B.-Y.; Zujovic, T.; Turner, D.A.; Hadad, C.M.; Badjic, J.D. Design, preparation, and study of catalytic gated baskets. *J. Org. Chem.* **2012**, *77*, 2675–2688.
39. Yoshizawa, M.; Klosterman, J.K.; Fujita, M. Functional molecular flasks: New properties and reactions within discrete, self-assembled hosts. *Angew. Chem. Int. Ed.* **2009**, *48*, 3418–3438.
40. Wang, H.; Liu, F.; Helgeson, R.C.; Houk, K.N. Reversible photochemically gated transformation of a hemicarcerand to a carcerand. *Angew. Chem. Int. Ed.* **2013**, *52*, 655–659.
41. Jencks, W.P. When is an intermediate not an intermediate? Enforced mechanisms of general acid-base, catalyzed, carbocation, carbanion, and ligand exchange reaction. *Acc. Chem. Res.* **1980**, *13*, 161–169.

Sample Availability: Samples of the compounds are not available.