

Review

Stem Cells for the Treatment of Neurodegenerative Diseases

Yong-Ping Wu, Wei-Shan Chen, Chong Teng and Ning Zhang *

Department of Orthopaedics, 2nd Affiliated Hospital, School of Medicine, Zhejiang University, #88 Jiefang Road, Hangzhou, 310009, China

* Author to whom correspondence should be addressed; E-Mail: zhangning98@gmail.com; Tel.: +86 57186021763; Fax: +86 57187022776.

Received: 13 August 2010; in revised form: 6 September 2010 / Accepted: 8 September 2010 / Published: 27 September 2010

Abstract: Neurodegenerative diseases are characterized by neurodegenerative changes or apoptosis of neurons involved in networks, leading to permanent paralysis and loss of sensation below the site of the injury. Cell replacement therapy has provided the basis for the development of potentially powerful new therapeutic strategies for a broad spectrum of human neurological diseases. In recent years, neurons and glial cells have successfully been generated from stem cells, and extensive efforts by investigators to develop stem cell-based brain transplantation therapies have been carried out. We review here notable previously published experimental and preclinical studies involving stem cell-based cell for neurodegenerative diseases and discuss the future prospects for stem cell therapy of neurological disorders in the clinical setting. Steady and solid progress in stem cell research in both basic and preclinical settings should support the hope for development of stem cell-based cell therapies for neurological diseases.

Keywords: stem cell; neurodegenerative diseases; treatment

1. Introduction

Neurodegenerative diseases, such as Parkinson's disease (PD), stroke, Huntington's disease (HD) and amyotrophic lateral sclerosis (ALS), are characterized by neurodegenerative changes or apoptosis of neurons involved in networks, leading to permanent paralysis and loss of sensation below the site of the injury [1]. Unfortunately, so far no successful treatment for neurodegenerative diseases has been

developed. Cell replacement therapy and gene transfer to the diseased or injured brain have provided the basis for the development of potentially powerful new therapeutic strategies for a broad spectrum of human neurological diseases. Stem cells are capable of repairing injured nervous tissue by replacing damaged cells, neuroprotection or the creation of an environment conducive to regeneration by endogenous cells [2]. The transplantation of stem cells may provide effective treatments due to the self-renewing and multipotential nature of these cells, including delivery of therapeutic factors to provide trophic support or missing gene products, mobilization of endogenous stem cells and replacement of lost or dysfunctional cells.

Meanwhile, human embryonic stem cells (ESCs) and adult stem cells have been coaxed into types of cells that repair neurodegenerative diseases insulation and replace nerve cells in neurodegenerative diseases [3]. The potency of these cells and the relative ease of isolating and expanding them are invaluable properties for clinical application. And some clinical trials have also been undertaken in neurodegenerative diseases [4,5]. Steady and solid progress in stem cell research in both basic and preclinical settings should support the hope for development of stem cell-based cell therapies for neurodegenerative diseases.

2. Stem Cell and Neurodegenerative Diseases

Here we review the scientific basis of stem cell therapies and discuss their prospects in Parkinson's disease, Huntington's disease, Alzheimer's disease, Amyotrophic Lateral Sclerosis, stroke and spinal cord injury. In each of these neurodegenerative diseases, we describe the ways in which stem cells might be used to treat these conditions, discussing the prospects and problems of translating laboratory findings into clinically useful therapies.

2.1. Stem cells and Parkinson's disease

Parkinson's disease is a neurodegenerative disorder characterized by the progressive loss of dopaminergic (DA) neurons in the substantia nigra pars compacta (SNpc) and a reduction in striatal dopamine. In PD, the loss of DA neurons in the SNpc leads to impaired information processing in the basal ganglia [6-8]. The main symptoms are rigidity, poor movement, tremors and postural instability [9,10]. Current therapies centre on the oral administration of L-dopa and dopamine receptor agonists, and on deep-brain stimulation in the sub-thalamic nucleus [10]. Although the pharmacological treatment is effective for some symptoms, it has some limitations because its effectiveness decreases over time and side effects develop [11,12]. Thus, an alternative approach for restoration of the damaged DA system is transplantation of DA-synthesizing cells. Human stem cells may provide sources of cells for use in the treatment of PD.

Studies show that stem cells overexpressing neurotrophic factors are able to induce neuroprotective and neuroregenerative effects after grafting in animal models [13,14]. Recently, continuously dividing immortalized cell lines of neural stem cells (NSCs) have been generated by introduction of oncogenes, and these immortalized NSC lines have advantages for basic studies of neural development and cell replacement therapy or gene therapy studies. To be clinically competitive, a stem-cell-based therapy must lead to long-lasting, significant improvement in mobility, ameliorate currently intractable symptoms, or counteract disease progression. Clinical trials of the transplantation of human fetal DA

neurons have shown that cell replacement can produce major, long-lasting improvement in some patients [15-17]. To make stem-cell therapy work for PD, dopaminergic neurons with the characteristics of substantia nigra neurons must be produced in large numbers [18].

For DA neurons generated from human ESCs and NSCs, survival after transplantation in animal models has been poor and needs to be markedly increased before clinical application will be possible [19]. One potential approach to prevent the death of existing neurons could be to transplant human stem cells engineered to express neuroprotective molecules such as glial-cell-line-derived neurotrophic factor (GDNF) [20]. Another study has demonstrated that retinoic acid treatment and transplanting ESCs to the lesioned brain can lead to the generation of putative DA neurons and functional recovery in Parkinsonian rat model [21]. Meanwhile, a study showed that L1-overexpressing stem cell-derived neural aggregates could enhance survival and migration of transplanted cells, differentiation into DA neurons, survival of endogenous DA neurons, and functional recovery after syngeneic transplantation in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of PD [22].

In addition, the effects of growth factors on embryonic DA neuron grafts were recently studied [23,24]. The data suggested that long-term and continuous neurotrophic factors support provided by Zuckerkandl's organ to the transplanted NSC derived DA neurons, helped in their better survival, axonal arborization and integration with host cells, leading to long-term functional restoration in the rat model of PD. For these reasons, it is scientifically and medically significant to test major refinements of cell therapies for Parkinson's disease.

2.2. Stem cells and Huntington's disease

Huntington's disease is a fatal, intractable disorder that is caused by polyglutamate expansions in the huntingtin protein [25-29]. Neuronal dysfunction and degeneration contribute to the progressive physiological, motor, cognitive and emotional disturbances characteristic of HD. One strategy for therapy of HD is to enhance neurogenesis, and the recent treatment of HD has centered on cell therapy strategies to protect vulnerable neuronal cell populations or to replace dysfunctional or dying cells [30-34]. Stem-cell therapy aims to restore or preserve brain function by replacing and protecting striatal neurons. At this time, using stem cells for the delivery of trophic factors and neuroprotection to prevent disease progression seems a more achievable clinical goal in HD than neuronal replacement.

Most of the recent work in stem cell therapy has been conducted in animal models of HD. The study showed that cell replacement using grafts of fetal striatal neurons promoted functional recovery, and some evidence from clinical trials indicates that this could also occur in patients [35]. The study demonstrated that hMSCs could affect endogenous cells in the striatum of HD mice by increasing cell proliferation, neuronal differentiation, and neuronal cell recruitment, and implantation of hMSCs could prevent striatal atrophy associated with HD [36]. Moreover, human NSCs implanted into the brains of rats were recently found to reduce motor impairments in experimental HD through trophic mechanisms [37]. Human NSCs appear to behave similarly to murine-derived NSCs in rodent models of HD. Intravenously transplanted human NSCs migrate to the striatum, reduce striatal atrophy, and contribute to functional improvement in a rodent lesion model of HD [38,39].

However, protocols and procedures developed from trials of fetal-derived cell transplantation in humans with HD lay the ground-work to move stem cell therapy into the clinic [40]. One of the first

challenges to stem cell therapy in HD is to determine which source of stem cells is most efficacious, and many sources have been examined. In addition to human ESCs, stem cells derived from mesenchyme in adults have been investigated as a readily available source of stem cells in HD. Following transplantation into a mouse model of HD, murine ESC-derived NPCs, genetically modified to promote neuronal differentiation, formed GABAergic neurons with appropriate out-growth [41]. Besides, already the method of graft preparation of NSCs for transplantation, as well as the timing of the transplantation procedure strongly could affect the survival of the donor cells when grafted into the quinolinic acid (QA)-lesioned striatum of adult mice [41]. Furthermore, the studies clearly pointed to the fact that immune responses in the CNS are complex, but cannot be ignored when it comes to repair strategies involving cellular transplants. New experimental studies are definitely needed in order to better comprehend and delineate the immunological privilege of the brain [42-44]. The results of those experimental studies will allow defining more precisely the extent to which immunosuppressive regimens can be adapted to the specific case of intracerebral transplants [45-47].

The grafts were also functionally beneficial given that the animals showed improvement in rotational behavior. In HD, mitochondrial defects are thought to play a significant part in the pathomechanism for cell death in HD pathology. Administration of 3NP, which inhibits mitochondrial succinate dehydrogenase (SDH), can induce the behavioral and anatomical features of HD in rodents and primates. Moreover, data demonstrated that stem cell factor (SCF), produced *in situ* in the lesioned striatum, was an important factor in promoting the engraftment of stem cells within the lesioned brain which could be able to activate the SCF receptor c-kit and its signaling pathway and to promote the migration and proliferation of mesenchymal and neural stem cells *in vitro* [48-50].

Taken together, neural implantation of stem cells may be of benefit in HD but a number of parameters of dose, treatment schedule, and route of administration need to be optimized.

2.3. Stem cells and Alzheimer's disease

Alzheimer's disease (AD) is characterized by neuronal and synaptic loss throughout the brain, involving the basal forebrain cholinergic system, amygdala, hippocampus and several cortical areas [33,51-54]. Although in AD massive neuronal loss only occurs in very few brain structures, such as the hippocampal CA1 and CA2 regions, the entorhinal cortex and the locus coeruleus, large parts of the brain are affected by pathological alterations and decreased neuronal metabolism [55-57]. Current therapies, such as treatment with acetylcholinesterase inhibitors to enhance cholinergic function, provide only partial and temporary alleviation of symptoms [34]. The pathological changes seen in AD offer an extremely problematic situation for cell replacement. The data show that neural stem cells release diffusible factors that may improve the survival of aged and degenerating neurons in human brains [58].

Alzheimer's disease amyloid precursor protein [13] has been implicated in many neurobiologic processes, but supporting evidence remains indirect [59-62]. Studies are confounded by the existence of two partially redundant APP homologues, APLP1 and APLP2 [63]. The stem cell culture provides an excellent tool to circumvent the problem of lack of viability of APP/APLP triple knockout mice and would help to explore the function of this intriguing protein further *in vitro* and *in vivo*. A morphological abnormality of neurally-differentiated NSCs has also been described, which we too

have seen in NSCs transfected with wild-type APP [64]. Although it remains unclear as to whether or not adult neurogenesis is essential for normal cognitive function in aging [65], it is tempting to speculate that the altered APP metabolism that impairs proper NSCs migration and differentiation could be a part of the pathological process of AD, particularly since aged transgenic APP mice exhibit neocortical neuronal loss [66,67]. Furthermore, although the rate of neuroregeneration in the adult brain may be minimal, it may be that, in the long run, such a deficit significantly reduces normal brain function. In addition to these drawbacks, the use of transplantation therapy for AD with NSCs may not be effective in an environment where APP metabolism is altered and might lead to excessive gliogenesis. They considered the regulation of APP processing to develop effective NSC transplantation therapy for AD patients [68].

Because stem cells can be genetically modified to carry new genes and have high migratory capacity after brain transplantation, they could be used in place of fibroblasts that are known for their immobility following transplantation for delivery of nerve growth factor (NGF) to prevent degeneration of basal forebrain cholinergic neurons [69,70]. However, because stem cells can be genetically modified and have migratory capacity after transplantation, they could be used for the delivery of factors that can modify the course of the disease [71,72]. In support of this approach, basal forebrain grafts of fibroblasts that produce NGF, which counteracts cholinergic neuronal death, stimulates cell function and improves memory in animal models, have been of some benefit in patients with AD [73].

2.4. Stem cells and Amyotrophic Lateral Sclerosis

Amyotrophic Lateral Sclerosis is an adult-onset neurodegenerative disorder characterized by degeneration and loss of motor neurons in the cerebral cortex, brainstem and spinal cord, leading to fatal paralysis [74,75]. A stem-cell therapy could restore or preserve the function of both upper and lower motor neurons, and new neurons could become integrated into existing neural circuitries [76,77].

Recent studies have indicated that it is possible to generate motor neurons in culture from stem cells that include ESCs and NSCs. Mouse ESC-derived motor neurons transplanted into motor neuron-injured rat spinal cord survived and extended axons into ventral root, and human ESCs transplanted into cerebrospinal fluid of rats with motor neuron injury migrated into spinal cord and led to improved motor function [78-80]. Reports have also shown that it is possible to generate lower motor neurons *in vitro* from stem cells of various sources, including ESCs and those from the fetal CNS [81]. Mouse ES-cell-derived motor neurons establish functional synapses with muscle fibres *in vitro*, and extend axons to ventral roots after transplantation into adult rats, but whether these neurons can integrate into existing neural circuitries and restore motor function has not been established. Whereas neuronal replacement in ALS patients seems a distant goal, using stem cells to prevent motor neurons from dying is a more realistic and shorter-term clinical approach. This prospect is supported by studies showing that human embryonic germ cells delivered into the cerebrospinal fluid of rats with motor neuron injury can migrate into the spinal cord and induce motor recovery, probably through neuroprotection [82-84]. The efficacy of this approach could be improved by genetically modifying the stem cells to secrete molecules that promote motor neuron survival.

It is unrealistic to expect that the transplantation of stem cells or stem cell-derived motor neurons in ALS patients in a clinical setting replaces lost neurons, integrates into existing neural circuitry, and restores motor function. Rather, preventing cell death in host motor neurons via provision of neurotrophic factors by transplanted stem cells or stem cell-derived motor neurons is a more realistic and achievable approach [85-87]. For instance, a recent study showed that human cortical progenitors that were engineered to express GDNF survived implantation into the spinal cords of ALS rats and released the neurotrophic factor [88]. Furthermore the studies indicated that ESC-derived cell populations can be directed to express disease-relevant genes and to display characteristics of the disease-specific cell type. These genetically manipulated ESC-derived motor neurons can facilitate and advance the study of disease-specific cellular pathways, and serve as a model system to test new therapeutic approaches [23,89-91].

The recent breakthroughs in stem cell research might nevertheless provide possibilities for neural implantation and cell replacement therapy for patients with ALS. Kim *et al.* showed that intrathecal injection with an optimized cell number could be a potential route for stem cell therapy in ALS patients. They suggested that at this dose of 1×10^6 , the average number of motor neurons was significantly higher than others, and most injected hMSCs distributed in the ventricular system and subarachnoid space [92,93]. Additionally, the studies suggested that successful stem cell therapy for ALS likely would require that the cells be combined with other drugs or treatments, such as antioxidants and/or trophic molecules. Many exciting studies are taking this direction; both *in vitro* and *in vivo* studies have shown generation of motor neurons from human ESCs and functional engraftment of these motor neurons after transplantation into the developing chick and adult rodent spinal cord with axonal outgrowth toward muscle [88,94-97]. Recently, a Phase I clinical trial confirmed that MSCs transplantation into the spinal cord of ALS patients is safe and that MSCs might have a clinical use for future ALS cell based clinical trials [88].

2.5. Stems cell and stroke

Once stroke damage has maximized, little can be done to recover premorbid function. In addition to therapies aimed at improving cerebral blood flow, there has been increasing emphasis on neuroprotective strategies. Recent attention has focused on restoring brain function through cell transplantation [98-103]. A variety of cell types have been tried for restoration of brain function after stroke, mostly in rodent models. The technical and ethical difficulties associated with these cells promoted a search for alternatives. Autologous somatic stem cells are a very attractive source, and there are no ethical concerns and graft rejection is not an issue. However, it is not clear that somatic cells can be plastic enough and can be safely induced to a neural fate.

A recent study has reported that, in humans with ischemic infarct, intracerebral implantation of human teratocarcinoma NT2-derived neurons has resulted in functional improvement. As a first step towards this goal, human fetal NSCs were transplanted into the brains of stroke-damaged rats, resulting in the migration of new neurons towards the ischaemic lesion [104]. Other studies showed that monkey ES-cell-derived progenitors transplanted into the brains of mice after stroke differentiated into various types of neuron and glial cell, re-established connections with target areas, and led to improved motor function [105-109]. In the present report, the possible therapeutic strategy of the stem

cell transplantation for the stroke is discussed. Transplanted human NSCs migrated to the lesion site and differentiated into neurons and astrocytes, and three to twelve weeks post-transplantation, a functional improvement was observed in the transplanted animals compared with nongrafted controls on rotarod and turning-in-an-alley tests. For several months after a stroke, NS cells can generate new striatal neurons that migrate to the site of damage. It is now important to establish whether endogenous neurogenesis can contribute to functional recovery after stroke, and whether it occurs in humans [110].

The therapeutic efficacy of such strategies could be improved further by genetically modifying the stem cells, for example, by overexpressing an anti-apoptotic gene. Interestingly, the stroke-damaged adult rodent brain has some capacity for neuronal replacement from its own NSCs [111]. The distinct population of progenitor cells in the bone marrow is thought to retain the potential for both neural production and differentiation, and may contribute to a therapeutic strategy for stroke. And, because the regeneration of cortical neurons will be the basis for functional improvement in most stroke-damaged brains, it is also needed to know whether the adult brain's own NS cells can be triggered to produce cortical neurons [112]. For maximal functional recovery, however, regenerative therapy may need to follow combinatorial approaches, which may include cell replacement, trophic support, protection from oxidative stress, and the neutralization of the growth-inhibitory components for endogenous neuronal stem cells.

2.6. Stem cells and spinal cord injury

Spinal cord injury (SCI) invariably results in the loss of neurons and axonal degeneration at the lesion site, leading to permanent paralysis and loss of sensation below the site of the injury. It interrupts ascending and descending axonal pathways, and causes a loss of neurons and glia, inflammation and demyelination [113].

Preclinical studies have been performed on rats with a spinal cord injury and have shown that transplanted MSCs in the injured spinal cord survive, migrate into the host tissue and lead to axonal regeneration and motor function recovery [114]. Dasari *et al.* showed that expression of caspase-3 on both neurons and oligodendrocytes after SCI was significantly down-regulated by MSCs [114,115]. And treatment with MSCs had a positive effect on behavioral outcome and histopathological assessment after SCI. There is no cure, and the most common current treatment high-dose methylprednisolone is of questionable value. The transplantation of stem cells into injured spinal cord can lead to functional benefits, mainly through trophic factor secretion or the remyelination of spared axons. A recent study showed that human NS cells implanted into damaged mouse spinal cord generated new neurons and oligodendrocytes, leading to locomotor recovery [116].

In addition, MSCs are attractive targets for *ex vivo* cell and gene therapy. Ronsyn *et al.* investigated the feasibility of a plasmid-based strategy for genetic modification of human MSCs with enhanced green fluorescent protein (EGFP) and neurotrophin (NT) [117]. Bakshi *et al.* found that MSCs delivered by lumbar puncture (LP) reached the contused spinal cord tissues and exerted a significant beneficial effect by reducing cyst and injury size [118]. Astrocytic differentiation and aberrant axonal sprouting after NSCs implantation into injured rat spinal cord can cause hypersensitivity to stimuli [119]. With regard to the yet controversial immunological status of stem cells, it is important to predict strategies to overcome the potential immunoincompatibility. To reach this aim, two main possibilities

can be foreseen. Banking of hESCs including only 150 donors with unique blood groups could provide a beneficial HLA matching for most potential patients. If confirmed, such a bank could be generated under GMP conditions and would avoid the need of somatic cell nuclear transfer to customize hESCs, a yet not successful approach in human beings [120]. Although chimerism between ESCs and recipients has been reported, another strategy to confer some immune tolerance to HES would be to generate tolerogenic hematopoietic cells derived from them. Together, these strategies demonstrate the possibilities to overcome the immunologic barrier [121].

3. Conclusions

This review has discussed the major issues associated with stem cell therapy by transplantation for neurodegenerative diseases. Stem cells from a variety of sources have shown effectiveness in improving motor function after neurodegenerative diseases in animal experiments and clinical trials. Cell therapies in neurodegenerative disease are intended to protect neuronal populations susceptible to disease and replace dysfunctional or dying neurons. The use of both stem cell and growth factor-based therapies, although in its early stages, appears likely to contribute to future clinical strategies, including, but not limited to, neuroprotective and neuron replacement approaches. However, factors that control the differentiation, survival, and maturation of stem cells in the context of a host degenerative brain must be more thoroughly understood before stem cell therapy will prove to be a robust and safe strategy that can be transferred to the clinic. Furthermore, long-term and large scale multicenter clinical studies are required to determine further the precise therapeutic effect of stem cell transplantation.

Acknowledgements

Thanks must be given to Ying Yin, Sheng-Jie Xu for critical review of the original manuscript.

References and Notes

1. Kim, S.U.; de Vellis, J. Stem cell-based cell therapy in neurological diseases: a review. *J. Neurosci. Res.* **2009**, *87*, 2183-2200.
2. Lindvall, O.; Kokaia, Z.; Martinez-Serrano, A. Stem cell therapy for human neurodegenerative disorders-how to make it work. *Nat. Med.* **2004**, *10*, S42-S50.
3. Cui, Y.F.; Hargus, G.; Xu, J.C.; Schmid, J.S.; Shen, Y.Q.; Glatzel, M.; Schachner, M.; Bernreuther, C. Embryonic stem cell-derived L1 overexpressing neural aggregates enhance recovery in Parkinsonian mice. *Brain* **2010**, *133*, 189-204.
4. Grunt, R.F. Embryonic stem-cell research. *N. Engl. J. Med.* **2004**, *351*, 1797-1798.
5. Daley, G.Q. Missed opportunities in embryonic stem-cell research. *N. Engl. J. Med.* **2004**, *351*, 627-8.
6. Yang, D.; Zhang, Z.J.; Oldenburg, M.; Ayala, M.; Zhang, S.C. Human embryonic stem cell-derived dopaminergic neurons reverse functional deficit in parkinsonian rats. *Stem Cells* **2008**, *26*, 55-63.

7. Glass, C.K.; Saijo, K.; Winner, B.; Marchetto, M.C.; Gage, F.H. Mechanisms underlying inflammation in neurodegeneration. *Cell* **2010**, *140*, 918-934.
8. Dawson, T.M.; Dawson, V.L. Molecular pathways of neurodegeneration in Parkinson's disease. *Science* **2003**, *302*, 819-822.
9. Burnstein, R.M.; Foltynie, T.; He, X.; Menon, D.K.; Svendsen, C.N.; Caldwell, M.A. Differentiation and migration of long term expanded human neural progenitors in a partial lesion model of Parkinson's disease. *Int. J. Biochem. Cell Biol.* **2004**, *36*, 702-713.
10. Kim, J.H.; Auerbach, J.M.; Rodriguez-Gomez, J.A.; Velasco, I.; Gavin, D.; Lumelsky, N.; Lee, S.H.; Nguyen, J.; Sanchez-Pernaute, R.; Bankiewicz, K.; McKay, R. Dopamine neurons derived from embryonic stem cells function in an animal model of Parkinson's disease. *Nature* **2002**, *418*, 50-56.
11. Prokai-Tatrai, K.; Prokai, L. Prodrugs of thyrotropin-releasing hormone and related peptides as central nervous system agents. *Molecules* **2009**, *14*, 633-654.
12. Prokai-Tatrai, K.; Perjesi, P.; Rivera-Portalatin, N.M.; Simpkins, J.W.; Prokai, L. Mechanistic investigations on the antioxidant action of a neuroprotective estrogen derivative. *Steroids* **2008**, *73*, 280-288.
13. Cova, L.; Armentero, M.T.; Zennaro, E.; Calzarossa, C.; Bossolasco, P.; Busca, G.; Lambertenghi Delilieri, G.; Polli, E.; Nappi, G.; Silani, V.; Blandini, F. Multiple neurogenic and neurorescue effects of human mesenchymal stem cell after transplantation in an experimental model of Parkinson's disease. *Brain Res.* **2010**, *1311*, 12-27.
14. Venkataramana, N.K.; Kumar, S.K.; Balaraju, S.; Radhakrishnan, R.C.; Bansal, A.; Dixit, A.; Rao, D.K.; Das, M.; Jan, M.; Gupta, P.K.; Totey, S.M. Open-labeled study of unilateral autologous bone-marrow-derived mesenchymal stem cell transplantation in Parkinson's disease. *Transl. Res.* **2010**, *155*, 62-70.
15. Hedlund, E.; Pruszek, J.; Lardaro, T.; Ludwig, W.; Vinuela, A.; Kim, K.S.; Isacson, O. Embryonic stem cell-derived Pitx3-enhanced green fluorescent protein midbrain dopamine neurons survive enrichment by fluorescence-activated cell sorting and function in an animal model of Parkinson's disease. *Stem Cells* **2008**, *26*, 1526-1536.
16. Mine, Y.; Hayashi, T.; Yamada, M.; Okano, H.; Kawase, T. Environmental cue-dependent dopaminergic neuronal differentiation and functional effect of grafted neuroepithelial stem cells in parkinsonian brain. *Neurosurgery* **2009**, *65*, 741-753.
17. McLeod, M.; Hong, M.; Mukhida, K.; Sadi, D.; Ulalia, R.; Mendez, I. Erythropoietin and GDNF enhance ventral mesencephalic fiber outgrowth and capillary proliferation following neural transplantation in a rodent model of Parkinson's disease. *Eur. J. Neurosci.* **2006**, *24*, 361-370.
18. Ourednik, V.; Ourednik, J.; Xu, Y.F.; Zhang, Y.; Lynch, W.P.; Snyder, E.Y.; Schachner, M. Cross-Talk Between Stem Cells and the Dysfunctional Brain is Facilitated by Manipulating the Niche: Evidence from an Adhesion Molecule. *Stem Cells* **2009**, *27*, 2846-2856.
19. Oizumi, H.; Hayashita-Kinoh, H.; Hayakawa, H.; Arai, H.; Furuya, T.; Ren, Y.R.; Yasuda, T.; Seki, T.; Mizuno, Y.; Mochizuki, H. Alteration in the differentiation-related molecular expression in the subventricular zone in a mouse model of Parkinson's disease. *Neurosci. Res.* **2008**, *60*, 15-21.
20. Isacson, O. Cell therapy ahead for Parkinson's disease. *Science* **2009**, *326*, 1060.

21. Akerud, P.; Holm, P.C.; Castelo-Branco, G.; Sousa, K.; Rodriguez, F.J.; Arenas, E. Persephin-overexpressing neural stem cells regulate the function of nigral dopaminergic neurons and prevent their degeneration in a model of Parkinson's disease. *Mol. Cell. Neurosci.* **2002**, *21*, 205-222.
22. Shin, J.H.; Dawson, V.L.; Dawson, T.M. SnapShot: Pathogenesis of Parkinson's Disease. *Cell* **2009**, *139*, e1-e2.
23. Marchetto, M.C.; Winner, B.; Gage, F.H. Pluripotent stem cells in neurodegenerative and neurodevelopmental diseases. *Hum. Mol. Genet.* **2010**, *19*, R71-R76.
24. Nakamura, T.; Lipton, S.A. Redox regulation of mitochondrial fission, protein misfolding, synaptic damage, and neuronal cell death: potential implications for Alzheimer's and Parkinson's diseases. *Apoptosis* **2010**. doi: 10.1007/s10495-010-0476-x.
25. Steiner, B.; Wolf, S.; Kempermann, G. Adult neurogenesis and neurodegenerative disease. *Regen. Med.* **2006**, *1*, 15-28.
26. Zietlow, R.; Lane, E.L.; Dunnett, S.B.; Rosser, A.E. Human stem cells for CNS repair. *Cell Tissue Res.* **2008**, *331*, 301-322.
27. Dunnett, S.B.; Rosser, A.E. Stem cell transplantation for Huntington's disease. *Exp. Neurol.* **2007**, *203*, 279-292.
28. Kim, M.; Lee, S.T.; Chu, K.; Kim, S.U. Stem cell-based cell therapy for Huntington disease: a review. *Neuropathology* **2008**, *28*, 1-9.
29. McKay, R. Stem cells in the central nervous system. *Science* **1997**, *276*, 66-71.
30. Keene, C.D.; Rodrigues, C.M.P.; Eich, T.; Chhabra, M.S.; Steer, C.J.; Low, W.C. Tauroursodeoxycholic acid, a bile acid, is neuroprotective in a transgenic model of Huntington's disease. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 10671-10676.
31. Ramaswamy, S.; Shannon, K.M.; Kordower, J.H. Huntington's disease: Pathological mechanisms and therapeutic strategies. *Cell Transplant.* **2007**, *16*, 301-312.
32. Johann, V.; Schiefer, J.; Sass, C.; Mey, J.; Brook, G.; Kruttgen, A.; Schlangen, C.; Bernreuther, C.; Schachner, M.; Dihne, M.; Kosinski, C.M. Time of transplantation and cell preparation determine neural stem cell survival in a mouse model of Huntington's disease. *Exp. Brain Res.* **2007**, *177*, 458-470.
33. Cherubini, A.; Spoletini, I.; Peran, P.; Luccichenti, G.; Di Paola, M.; Sancesario, G.; Gianni, W.; Giubilei, F.; Bossu, P.; Sabatini, U.; Caltagirone, C.; Spalletta, G. A multimodal MRI investigation of the subventricular zone in mild cognitive impairment and Alzheimer's disease patients. *Neurosci. Lett.* **2010**, *469*, 214-218.
34. Wang, J.M.; Singh, C.; Liu, L.; Irwin, R.W.; Chen, S.; Chung, E.J.; Thompson, R.F.; Brinton, R.D. Allopregnanolone reverses neurogenic and cognitive deficits in mouse model of Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 6498-6503.
35. McBride, J.L.; Behrstock, S.P.; Chen, E.Y.; Jakel, R.J.; Siegel, I.; Svendsen, C.N.; Kordower, J.H. Human neural stem cell transplants improve motor function in a rat model of Huntington's disease. *J. Comp. Neurol.* **2004**, *475*, 211-219.
36. Snyder, B.R.; Chiu, A.M.; Prockop, D.J.; Chan, A.W. Human multipotent stromal cells (MSCs) increase neurogenesis and decrease atrophy of the striatum in a transgenic mouse model for Huntington's disease. *Plos One* **2010**, *5*, e9347.

37. Krystkowiak, P.; Gaura, V.; Labalette, M.; Rialland, A.; Remy, P.; Peschanski, M.; Bachoud-Levi, A.C. Alloimmunisation to donor antigens and immune rejection following foetal neural grafts to the brain in patients with Huntington's disease. *Plos One* **2007**, *2*, e166.
38. Lorincz, M.T.; Zawistowski, V.A. Expanded CAG repeats in the murine Huntington's disease gene increases neuronal differentiation of embryonic and neural stem cells. *Mol. Cell Neurosci.* **2009**, *40*, 1-13.
39. Kim, S.U. Genetically engineered human neural stem cells for brain repair in neurological diseases. *Brain Develop* **2007**, *29*, 193-201.
40. Clelland, C.D.; Barker, R.A.; Watts, C. Cell therapy in Huntington disease. *Neurosurg. Focus* **2008**, *24*, E9.
41. Ebert, A.D.; Barber, A.E.; Heins, B.M.; Svendsen, C.N. *Ex vivo* delivery of GDNF maintains motor function and prevents neuronal loss in a transgenic mouse model of Huntington's disease. *Exp. Neurol.* **2010**, *224*, 155-162.
42. Yang, C.R.; Yu, R.K. Intracerebral transplantation of neural stem cells combined with trehalose ingestion alleviates pathology in a mouse model of Huntington's disease. *J. Neurosci. Res.* **2009**, *87*, 26-33.
43. Roberts, T.J.; Price, J.; Williams, S.C.; Modo, M. Preservation of striatal tissue and behavioral function after neural stem cell transplantation in a rat model of Huntington's disease. *Neuroscience* **2006**, *139*, 1187-1199.
44. McBride, J.L.; Ramaswamy, S.; Gasmi, M.; Bartus, R.T.; Herzog, C.D.; Brandon, E.P.; Zhou, L.; Pitzer, M.R.; Berry-Kravis, E.M.; Kordower, J.H. Viral delivery of glial cell line-derived neurotrophic factor improves behavior and protects striatal neurons in a mouse model of Huntington's disease. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 9345-9350.
45. Laowtammathron, C.; Cheng, E.; Cheng, P.H.; Snyder, B.R.; Yang, S.H.; Johnson, Z.; Lorthongpanich, C.; Kuo, H.C.; Parnpai, R.; Chan, A.W. Monkey hybrid stem cells develop cellular features of Huntington's disease. *BMC Cell Biol.* **2010**, *11*, 12.
46. Mareschi, K.; Novara, M.; Rustichelli, D.; Ferrero, I.; Guido, D.; Carbone, E.; Medico, E.; Madon, E.; Vercelli, A.; Fagioli, F. Neural differentiation of human mesenchymal stem cells: Evidence for expression of neural markers and eag K⁺ channel types. *Exp. Hematol.* **2006**, *34*, 1563-1572.
47. Pineda, J.R.; Rubio, N.; Akerud, P.; Urban, N.; Badimon, L.; Arenas, E.; Alberch, J.; Blanco, J.; Canals, J.M. Neuroprotection by GDNF-secreting stem cells in a Huntington's disease model: optical neuroimage tracking of brain-grafted cells. *Gene Ther.* **2007**, *14*, 118-128.
48. Molero, A.E.; Gokhan, S.; Gonzalez, S.; Feig, J.L.; Alexandre, L.C.; Mehler, M.F. Impairment of developmental stem cell-mediated striatal neurogenesis and pluripotency genes in a knock-in model of Huntington's disease. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 21900-21905.
49. Huang, A.H.; Snyder, B.R.; Cheng, P.H.; Chan, A.W. Putative dental pulp-derived stem/stromal cells promote proliferation and differentiation of endogenous neural cells in the hippocampus of mice. *Stem Cells* **2008**, *26*, 2654-2663.
50. Bantubungi, K.; Blum, D.; Cuvelier, L.; Wislet-Gendebien, S.; Rogister, B.; Brouillet, E.; Schiffmann, S.N. Stem cell factor and mesenchymal and neural stem cell transplantation in a rat model of Huntington's disease. *Mol. Cell Neurosci.* **2008**, *37*, 454-470.

51. Oliveira, A.A., Jr.; Hodges, H.M. Alzheimer's disease and neural transplantation as prospective cell therapy. *Curr. Alzheimer Res.* **2005**, *2*, 79-95.
52. Lee, J.K.; Jin, H.K.; Endo, S.; Schuchman, E.H.; Carter, J.E.; Bae, J.S. Intracerebral Transplantation of Bone Marrow-Derived Mesenchymal Stem Cells Reduces Amyloid-Beta Deposition and Rescues Memory Deficits in Alzheimer's Disease Mice by Modulation of Immune Responses. *Stem Cells* **2010**, *28*, 329-343.
53. Blurton-Jones, M.; Kitazawa, M.; Martinez-Coria, H.; Castello, N.A.; Muller, F.J.; Loring, J.F.; Yamasaki, T.R.; Poon, W.W.; Green, K.N.; LaFerla, F.M. Neural stem cells improve cognition via BDNF in a transgenic model of Alzheimer disease. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 13594-13599.
54. Wu, S.; Sasaki, A.; Yoshimoto, R.; Kawahara, Y.; Manabe, T.; Kataoka, K.; Asashima, M.; Yuge, L. Neural stem cells improve learning and memory in rats with Alzheimer's disease. *Pathobiology* **2008**, *75*, 186-194.
55. Ryu, J.K.; Cho, T.; Wang, Y.T.; McLarnon, J.G. Neural progenitor cells attenuate inflammatory reactivity and neuronal loss in an animal model of inflamed AD brain. *J. Neuroinflammation* **2009**, *6*, 39.
56. Wu, L.; Sluiter, A.A.; Guo, H.F.; Balesar, R.A.; Swaab, D.F.; Zhou, J.N.; Verwer, R.W. Neural stem cells improve neuronal survival in cultured postmortem brain tissue from aged and Alzheimer patients. *J. Cell Mol. Med.* **2008**, *12*, 1611-1621.
57. Bergmans, B.A.; Shariati, S.A.M.; Habets, R.L.P.; Verstreken, P.; Schoonjans, L.; Muller, U.; Dotti, C.G.; De Strooper, B. Neurons Generated from APP/APLP1/APLP2 Triple Knockout Embryonic Stem Cells Behave Normally *in Vitro* and *in Vivo*: Lack of Evidence for a Cell Autonomous Role of the Amyloid Precursor Protein in Neuronal Differentiation. *Stem Cells* **2010**, *28*, 399-406.
58. Zhongling, F.; Gang, Z.; Lei, Y. Neural stem cells and Alzheimer's disease: challenges and hope. *Am. J. Alzheimers Dis. Other Demen.* **2009**, *24*, 52-57.
59. Holtzman, D.M. Alzheimer's disease: Moving towards a vaccine. *Nature* **2008**, *454*, 418-420.
60. Waldau, B.; Shetty, A.K. Behavior of neural stem cells in the Alzheimer brain. *Cell Mol. Life Sci.* **2008**, *65*, 2372-2384.
61. Lee, J.K.; Jin, H.K.; Bae, J.S. Bone marrow-derived mesenchymal stem cells reduce brain amyloid-beta deposition and accelerate the activation of microglia in an acutely induced Alzheimer's disease mouse model. *Neurosci. Lett.* **2009**, *450*, 136-141.
62. Wang, Q.; Matsumoto, Y.; Shindo, T.; Miyake, K.; Shindo, A.; Kawanishi, M.; Kawai, N.; Tamiya, T.; Nagao, S. Neural stem cells transplantation in cortex in a mouse model of Alzheimer's disease. *J. Med. Invest.* **2006**, *53*, 61-69.
63. Lee, H.J.; Lee, J.K.; Lee, H.; Carter, J.E.; Chang, J.W.; Oh, W.; Yang, Y.S.; Suh, J.G.; Lee, B.H.; Jin, H.K.; Bae, J.S. Human umbilical cord blood-derived mesenchymal stem cells improve neuropathology and cognitive impairment in an Alzheimer's disease mouse model through modulation of neuroinflammation. *Neurobiol. Aging* **2010**. doi: 10.1016/j.neurobiolaging.2010.03.024.
64. Salloway, S.; Mintzer, J.; Weiner, M.F.; Cummings, J.L. Disease-modifying therapies in Alzheimer's disease. *Alzheimers Dement.* **2008**, *4*, 65-79.

65. Sugaya, K. Possible use of autologous stem cell therapies for Alzheimer disease. *Neurobiol. Aging* **2004**, *25*, S24-S25.
66. Xuan, A.G.; Luo, M.; Ji, W.D.; Long, D.H. Effects of engrafted neural stem cells in Alzheimer's disease rats. *Neurosci. Lett.* **2009**, *450*, 167-171.
67. Imitola, J. Prospects for neural stem cell-based therapies for neurological diseases. *Neurotherapeutics* **2007**, *4*, 701-714.
68. Sugaya, K.; Merchant, S. How to Approach Alzheimer's Disease Therapy Using Stem Cell Technologies. *J. Alzheimers Dis.* **2008**, *15*, 241-254.
69. Santilli, G.; Lamorte, G.; Carlessi, L.; Ferrari, D.; Rota Nodari, L.; Binda, E.; Delia, D.; Vescovi, A.L.; De Filippis, L. Mild hypoxia enhances proliferation and multipotency of human neural stem cells. *Plos One* **2010**, *5*, e8575.
70. Heese, K.; Low, J.W.; Inoue, N. Nerve growth factor, neural stem cells and Alzheimer's disease. *Neurosignals* **2006**, *15*, 1-12.
71. Mucke, L. Neuroscience: Alzheimer's disease. *Nature* **2009**, *461*, 895-897.
72. Sugaya, K.; Kwak, Y.D.; Ohmitsu, O.; Marutle, A.; Greig, N.H.; Choumrina, E. Practical issues in stem cell therapy for Alzheimer's disease. *Curr. Alzheimer Res.* **2007**, *4*, 370-377.
73. Giacobini, E.; Becker, R.E. One hundred years after the discovery of Alzheimer's disease. A turning point for therapy? *J. Alzheimers Dis.* **2007**, *12*, 37-52.
74. Papadeas, S.T.; Maragakis, N.J. Advances in stem cell research for Amyotrophic Lateral Sclerosis. *Curr. Opin. Biotechnol.* **2009**, *20*, 545-551.
75. Yan, J.; Xu, L.; Welsh, A.M.; Chen, D.; Hazel, T.; Johe, K.; Koliatsos, V.E. Combined immunosuppressive agents or CD4 antibodies prolong survival of human neural stem cell grafts and improve disease outcomes in amyotrophic lateral sclerosis transgenic mice. *Stem Cells* **2006**, *24*, 1976-1985.
76. Silani, V.; Calzarossa, C.; Cova, L.; Ticozzi, N. Stem cells in amyotrophic lateral sclerosis: motor neuron protection or replacement? *CNS Neurol. Disord. Drug Target.* **2010**, *9*, 314-324.
77. Silani, V.; Cova, L.; Corbo, M.; Ciammola, A.; Polli, E. Stem-cell therapy for amyotrophic lateral sclerosis. *Lancet* **2004**, *364*, 200-202.
78. Yang, E.J.; Jiang, J.H.; Lee, S.M.; Hwang, H.S.; Lee, M.S.; Choi, S.M. Electroacupuncture reduces neuroinflammatory responses in symptomatic amyotrophic lateral sclerosis model. *J. Neuroimmunol.* **2010**, *223*, 84-91.
79. Thonhoff, J.R.; Ojeda, L.; Wu, P. Stem cell-derived motor neurons: applications and challenges in amyotrophic lateral sclerosis. *Curr. Stem Cell Res. Ther.* **2009**, *4*, 178-199.
80. Lepore, A.C.; Maragakis, N.J. Targeted stem cell transplantation strategies in ALS. *Neurochem. Int.* **2007**, *50*, 966-975.
81. de Aguilar, J.L.G.; Echaniz-Laguna, A.; Fergani, A.; Rene, F.; Meininger, V.; Loeffler, J.P.; Dupuis, L. Amyotrophic lateral sclerosis: all roads lead to Rome. *J. Neurochem.* **2007**, *101*, 1153-1160.
82. Neusch, C.; Bahr, M.; Schneider-Gold, C. Glia cells in amyotrophic lateral sclerosis: new clues to understanding an old disease? *Muscle Nerve* **2007**, *35*, 712-724.
83. Karumbayaram, S.; Kelly, T.K.; Paucar, A.A.; Roe, A.J.; Umbach, J.A.; Charles, A.; Goldman, S.A.; Kornblum, H.I.; Wiedau-Pazos, M. Human embryonic stem cell-derived motor neurons

- expressing SOD1 mutants exhibit typical signs of motor neuron degeneration linked to ALS. *Dis. Model Mech.* **2009**, *2*, 189-195.
84. Di Giorgio, F.P.; Boulting, G.L.; Bobrowicz, S.; Eggan, K.C. Human embryonic stem cell-derived motor neurons are sensitive to the toxic effect of glial cells carrying an ALS-causing mutation. *Cell Stem Cell* **2008**, *3*, 637-648.
 85. Vercelli, A.; Mereuta, O.M.; Garbossa, D.; Muraca, G.; Mareschi, K.; Rustichelli, D.; Ferrero, I.; Mazzini, L.; Madon, E.; Fagioli, F. Human mesenchymal stem cell transplantation extends survival, improves motor performance and decreases neuroinflammation in mouse model of amyotrophic lateral sclerosis. *Neurobiol. Dis.* **2008**, *31*, 395-405.
 86. Calvo, A.; Moglia, C.; Balma, M.; Chio, A. Involvement of immune response in the pathogenesis of amyotrophic lateral sclerosis: a therapeutic opportunity? *CNS Neurol. Disord. Drug Target.* **2010**, *9*, 325-330.
 87. Suzuki, M.; McHugh, J.; Tork, C.; Shelley, B.; Klein, S.M.; Aebischer, P.; Svendsen, C.N. GDNF Secreting Human Neural Progenitor Cells Protect Dying Motor Neurons, but Not Their Projection to Muscle, in a Rat Model of Familial ALS. *Plos One* **2007**, *2*, e689.
 88. Inoue, H. Neurodegenerative disease-specific induced pluripotent stem cell research. *Exp. Cell Res.* **2010**, *316*, 2560-2564.
 89. Mazzini, L.; Ferrero, I.; Luparello, V.; Rustichelli, D.; Gunetti, M.; Mareschi, K.; Testa, L.; Stecco, A.; Tarletti, R.; Miglioretti, M.; Fava, E.; Nasuelli, N.; Cisari, C.; Massara, M.; Vercelli, R.; Oggioni, G.D.; Carriero, A.; Cantello, R.; Monaco, F.; Fagioli, F. Mesenchymal stem cell transplantation in amyotrophic lateral sclerosis: A Phase I clinical trial. *Exp. Neurol.* **2010**, *223*, 229-237.
 90. Mazzini, L.; Vercelli, A.; Mareschi, K.; Ferrero, I.; Testa, L.; Fagioli, F. Mesenchymal stem cells for ALS patients. *Amyotroph. Lateral Scler.* **2009**, *10*, 123-124.
 91. Corti, S.; Locatelli, F.; Papadimitriou, D.; Del Bo, R.; Nizzardo, M.; Nardini, M.; Donadoni, C.; Salani, S.; Fortunato, F.; Strazzer, S.; Bresolin, N.; Comi, G.P. Neural stem cells LewisX+ CXCR4+ modify disease progression in an amyotrophic lateral sclerosis model. *Brain* **2007**, *130*, 1289-1305.
 92. Boillee, S.; Vande Velde, C.; Cleveland, D.W. ALS: A disease of motor neurons and their nonneuronal neighbors. *Neuron* **2006**, *52*, 39-59.
 93. Kim, H.; Kim, H.Y.; Choi, M.R.; Hwang, S.; Nam, K.H.; Kim, H.C.; Han, J.S.; Kim, K.S.; Yoon, H.S.; Kim, S.H. Dose-dependent efficacy of ALS-human mesenchymal stem cells transplantation into cisterna magna in SOD1-G93A ALS mice. *Neurosci. Lett.* **2010**, *468*, 190-194.
 94. Garbuzova-Davis, S.; Sanberg, P.R. Feasibility of cell therapy for amyotrophic lateral sclerosis. *Exp. Neurol.* **2009**, *216*, 3-6.
 95. Choi, M.R.; Kim, H.Y.; Park, J.Y.; Lee, T.Y.; Baik, C.S.; Chai, Y.G.; Jung, K.H.; Park, K.S.; Roh, W.; Kim, K.S.; Kim, S.H. Selection of optimal passage of bone marrow-derived mesenchymal stem cells for stem cell therapy in patients with amyotrophic lateral sclerosis. *Neurosci. Lett.* **2010**, *472*, 94-98.
 96. Gornall, J. Stem cell renegades or pioneers? *BMJ* **2010**, *340*, c2041.

97. Mazzini, L.; Mareschi, K.; Ferrero, I.; Vassallo, E.; Oliveri, G.; Nasuelli, N.; Oggioni, G.D.; Testa, L.; Fagioli, F. Stem cell treatment in Amyotrophic Lateral Sclerosis. *J. Neurol. Sci.* **2008**, *265*, 78-83.
98. Savitz, S.I.; Rosenbaum, D.M.; Dinsmore, J.H.; Wechsler, L.R.; Caplan, L.R. Cell transplantation for stroke. *Ann. Neurol.* **2002**, *52*, 266-275.
99. Cui, X.; Chopp, M.; Zacharek, A.; Roberts, C.; Lu, M.; Savant-Bhonsale, S.; Chen, J. Chemokine, vascular and therapeutic effects of combination Simvastatin and BMSC treatment of stroke. *Neurobiol. Dis.* **2009**, *36*, 35-41.
100. Hayase, M.; Kitada, M.; Wakao, S.; Itokazu, Y.; Nozaki, K.; Hashimoto, N.; Takagi, Y.; Dezawa, M. Committed neural progenitor cells derived from genetically modified bone marrow stromal cells ameliorate deficits in a rat model of stroke. *J. Cerebr. Blood Flow Metabol.* **2009**, *29*, 1409-1420.
101. Liu, Z.; Li, Y.; Zhang, X.; Savant-Bhonsale, S.; Chopp, M. Contralesional axonal remodeling of the corticospinal system in adult rats after stroke and bone marrow stromal cell treatment. *Stroke* **2008**, *39*, 2571-2577.
102. Chen, J.; Li, Y.; Katakowski, M.; Chen, X.; Wang, L.; Lu, D.; Lu, M.; Gautam, S.C.; Chopp, M. Intravenous bone marrow stromal cell therapy reduces apoptosis and promotes endogenous cell proliferation after stroke in female rat. *J. Neurosci. Res.* **2003**, *73*, 778-786.
103. Zhao, M.Z.; Nonoguchi, N.; Ikeda, N.; Watanabe, T.; Furutama, D.; Miyazawa, D.; Funakoshi, H.; Kajimoto, Y.; Nakamura, T.; Dezawa, M.; Shibata, M.A.; Otsuki, Y.; Coffin, R.S.; Liu, W.D.; Kuroiwa, T.; Miyatake, S. Novel therapeutic strategy for stroke in rats by bone marrow stromal cells and ex vivo HGF gene transfer with HSV-1 vector. *J. Cerebr. Blood Flow Metabol.* **2006**, *26*, 1176-1188.
104. Haas, S.; Weidner, N.; Winkler, J. Adult stem cell therapy in stroke. *Curr. Opin. Neurol.* **2005**, *18*, 59-64.
105. Roh, J.K.; Jung, K.H.; Chu, K. Adult stem cell transplantation in stroke: its limitations and prospects. *Curr. Stem Cell Res. Ther.* **2008**, *3*, 185-196.
106. Li, Y.; McIntosh, K.; Chen, J.; Zhang, C.; Gao, Q.; Borneman, J.; Raginski, K.; Mitchell, J.; Shen, L.; Zhang, J.; Lu, D.; Chopp, M. Allogeneic bone marrow stromal cells promote glial-axonal remodeling without immunologic sensitization after stroke in rats. *Exp. Neurol.* **2006**, *198*, 313-325.
107. Zhang, C.; Li, Y.; Chen, J.; Gao, Q.; Zacharek, A.; Kapke, A.; Chopp, M. Bone marrow stromal cells upregulate expression of bone morphogenetic proteins 2 and 4, gap junction protein connexin-43 and synaptophysin after stroke in rats. *Neuroscience* **2006**, *141*, 687-695.
108. Li, Y.; Chen, J.; Zhang, C. L.; Wang, L.; Lu, D.; Katakowski, M.; Gao, Q.; Shen, L. H.; Zhang, J.; Lu, M.; Chopp, M. Gliosis and brain remodeling after treatment of stroke in rats with marrow stromal cells. *Glia* **2005**, *49*, 407-417.
109. Kubinova, S.; Sykova, E. Nanotechnology for treatment of stroke and spinal cord injury. *Nanomedicine (Lond)* **2010**, *5*, 99-108.
110. Lu, M.; Chen, J.; Lu, D.; Yi, L.; Mahmood, A.; Chopp, M. Global test statistics for treatment effect of stroke and traumatic brain injury in rats with administration of bone marrow stromal cells. *J. Neurosci. Meth.* **2003**, *128*, 183-190.

111. Hicks, A.; Schallert, T.; Jolkkonen, J. Cell-based therapies and functional outcome in experimental stroke. *Cell Stem Cell* **2009**, *5*, 139-140.
112. Li, Y.; Chopp, M. Marrow stromal cell transplantation in stroke and traumatic brain injury. *Neurosci. Lett.* **2009**, *456*, 120-123.
113. Takahashi, K.; Yamanaka, S., Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* **2006**, *126*, 663-676.
114. Ogawa, Y.; Sawamoto, K.; Miyata, T.; Miyao, S.; Watanabe, M.; Nakamura, M.; Bregman, B.S.; Koike, M.; Uchiyama, Y.; Toyama, Y.; Okano, H. Transplantation of in vitro-expanded fetal neural progenitor cells results in neurogenesis and functional recovery after spinal cord contusion injury in adult rats. *J. Neurosci. Res.* **2002**, *69*, 925-933.
115. Perrier, A.L.; Tabar, V.; Barberi, T.; Rubio, M.E.; Bruses, J.; Topf, N.; Harrison, N.L.; Studer, L. Derivation of midbrain dopamine neurons from human embryonic stem cells. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 12543-12548.
116. Nisbet, D.R.; Moses, D.; Gengenbach, T.R.; Forsythe, J.S.; Finkelstein, D.I.; Horne, M.K. Enhancing neurite outgrowth from primary neurones and neural stem cells using thermoresponsive hydrogel scaffolds for the repair of spinal cord injury. *J. Biomed. Mater. Res. A* **2009**, *89A*, 24-35.
117. Song, S.J.; Song, S.J.; Zhang, H.L.; Cuevas, J.; Sanchez-Ramos, J. Comparison of neuron-like cells derived from bone marrow stem cells to those differentiated from adult brain neural stem cells. *Stem Cells Dev.* **2007**, *16*, 747-756.
118. Lee, H.; Shamy, G.A.; Elkabetz, Y.; Schofield, C.M.; Harrison, N.L.; Panagiotakos, G.; Socci, N.D.; Tabar, V.; Studer, L. Directed differentiation and transplantation of human embryonic stem cell-derived motoneurons. *Stem Cells* **2007**, *25*, 1931-1939.
119. Keirstead, H.S.; Nistor, G.; Bernal, G.; Totoiu, M.; Cloutier, F.; Sharp, K.; Steward, O. Human embryonic stem cell-derived oligodendrocyte progenitor cell transplants remyelinate and restore locomotion after spinal cord injury. *J. Neurosci.* **2005**, *25*, 4694-4705.
120. Ronsyn, M.W.; Daans, J.; Spaepen, G.; Chatterjee, S.; Vermeulen, K.; D'Haese, P.; Van Tendeloo, V.F.; Van Marck, E.; Ysebaert, D.; Berneman, Z.N.; Jorens, P.G.; Ponsaerts, P. Plasmid-based genetic modification of human bone marrow-derived stromal cells: analysis of cell survival and transgene expression after transplantation in rat spinal cord. *BMC Biotechnol.* **2007**, *7*, 90.
121. Roy, N.S.; Cleren, C.; Singh, S.K.; Yang, L.; Beal, M.F.; Goldman, S.A. Functional engraftment of human ES cell-derived dopaminergic neurons enriched by coculture with telomerase-immortalized midbrain astrocytes. *Nat. Med.* **2006**, *12*, 1259-68.