



Phenolic Profiles of Six Unexplored Asteraceae Species from Asia: Comparison of Wild and Cultivated Plants

Daniil N. Olennikov ^{1,*}, Nadezhda K. Chirikova ²

¹ Laboratory of Medical and Biological Research, Institute of General and Experimental Biology, Siberian Division, Russian Academy of Science, 6 Sakhyanovoy Street, 670047 Ulan-Ude, Russia

² Department of Biochemistry and Biotechnology, North-Eastern Federal University, 58 Belinsky Street, 677027 Yakutsk, Russia

* Correspondence: olennikovdn@mail.ru; Tel.: +7-902-160-06-27

Supplementary content

Figure S1. Typical UV spectra of the basic phenolic compounds found in six Asteraceae species.

Figure S2. Flow chart for the experimental design.

Figure S3. HPLC-MS chromatograms of *Artemisia jacutica* herb in SIM mode.

Figure S4. HPLC-MS chromatograms of *Carduus nutans* subsp. *leiophyllus* herb in SIM mode.

Figure S5. HPLC-MS chromatograms of *Cirsium heterophyllum* herb in SIM mode.

Figure S6. HPLC-MS chromatograms of *Echinops davuricus* herb in SIM mode.

Figure S7. HPLC-MS chromatograms of *Ixeris chinensis* subsp. *versicolor* herb in SIM mode.

Figure S8. HPLC-MS chromatograms of *Lactuca sibirica* herb in SIM mode.

Table S1. Description of wild samples of Asteraceae species collected in Altacheiskii reserve for a period 2010–2020 and information on cultivation success.

Table S2. Plant cultivation conditions of six Asteraceae species.

Table S3. Plant sample description for HPLC assay.

Table S4. Reference standards used for HPLC profiling and quantification.

Table S5. HPLC and mass-spectral conditions used for metabolite separation

Table S6. Regression equations, correlation coefficients, standard deviation, limits of detection, limits of quantification and linear ranges for 55 reference standards.

Table S7. Retention times, molecular formulas, mass-spectral data for negative ionization, and identification level of compounds Aj-1–Aj-32 found in *Artemisia jacutica* herb.

Table S8. Retention times, molecular formulas, mass-spectral data for negative ionization, and identification level of compounds Cn-1–Cn-20 found in *Carduus nutans* subsp. *leiophyllus* herb.

Table S9. Extraction/chromatography conditions and spectral data of luteolin 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside and chrysoeriol 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside isolated from *Carduus nutans* subsp. *leiophyllus* leaves.

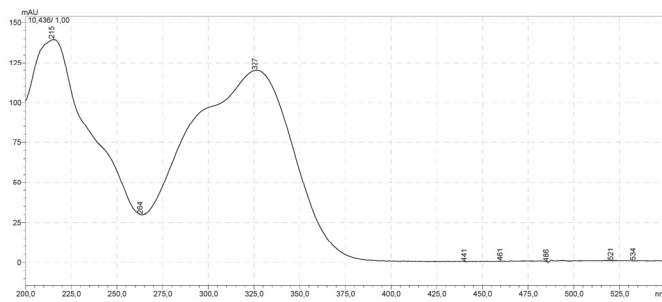
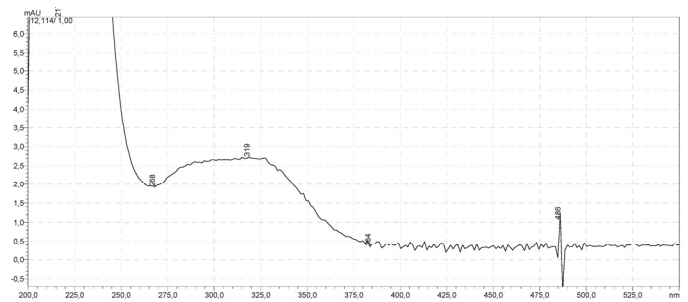
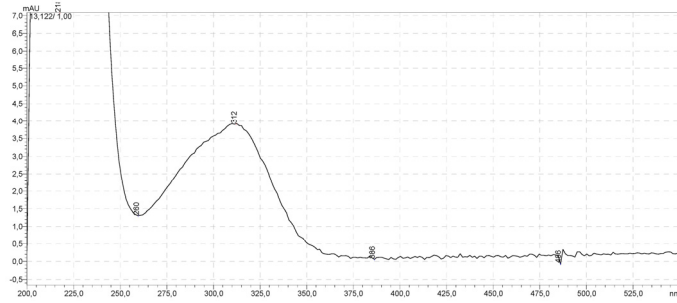
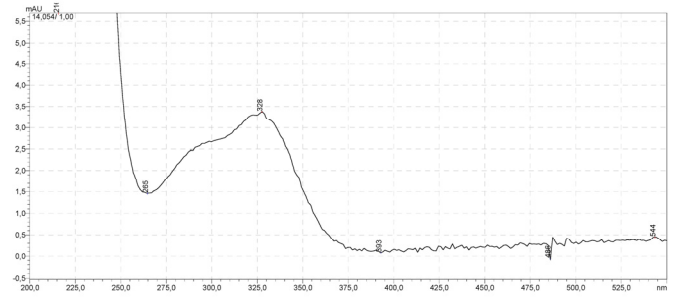
Table S10. Retention times, molecular formulas, mass-spectral data for negative ionization, and identification level of compounds Ch-1–Ch-24 found in *Cirsium heterophyllum* herb.

Table S11. Retention times, molecular formulas, mass-spectral data for negative ionization, and identification level of compounds El-1–El-19 found in *Echinops davuricus* herb.

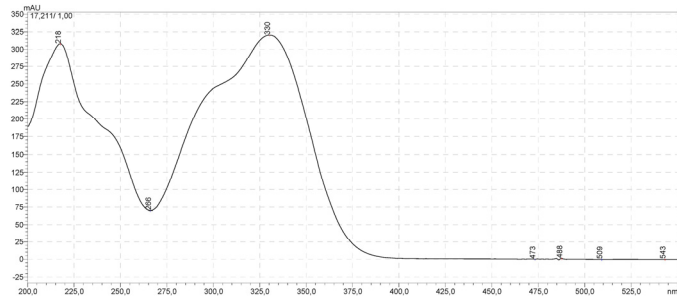
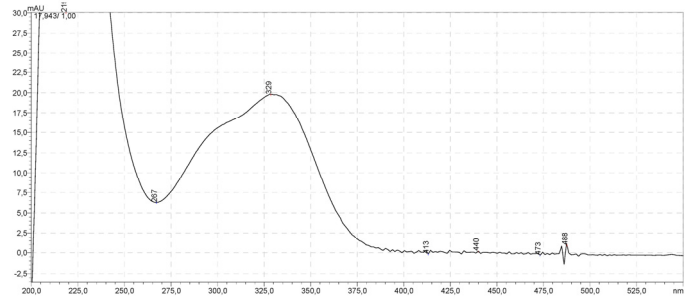
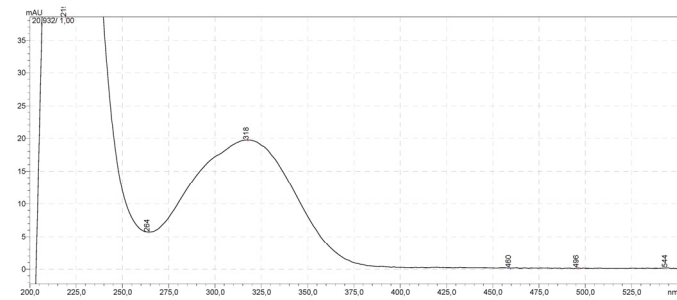
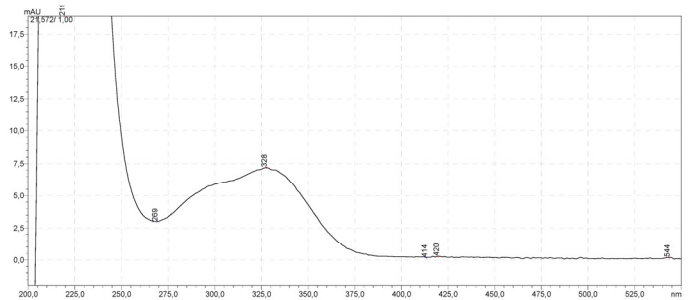
Table S12. Retention times, molecular formulas, mass-spectral data for negative ionization, and identification level of compounds Ic-1–Ic-25 found in *Ixeris chinensis* subsp. *versicolor* herb.

Table S13. Retention times, molecular formulas, mass-spectral data for negative ionization, and identification level of compounds Ls-1–Ls-41 found in *Lactuca sibirica* herb.

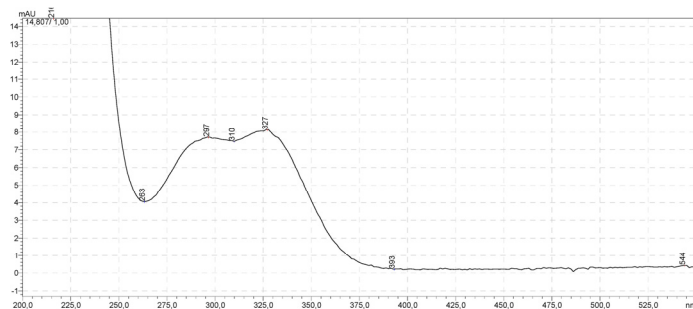
Table S14. Synopsis of compounds found in six Asteraceae species.

Caffeoyl-quinic acids, *trans*-; dicaffeoyl-quinic acidsCaffeoyl-quinic acids, *cis*-*p*-Coumaroyl-quinic acids

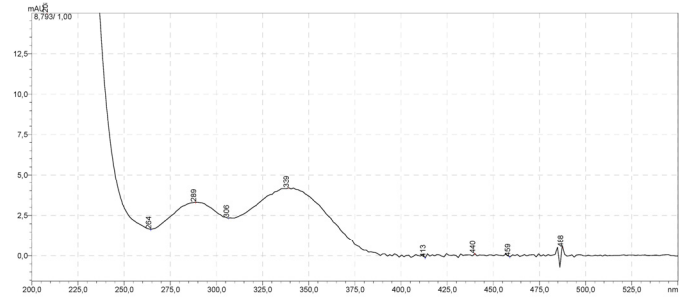
Feruloyl-quinic acids

Cichoric acid, *trans,trans*-; caftaric acidCichoric acid, *cis,trans*-*p*-Coumaroyl-caftaric acid

Feruloyl-caftaric acid

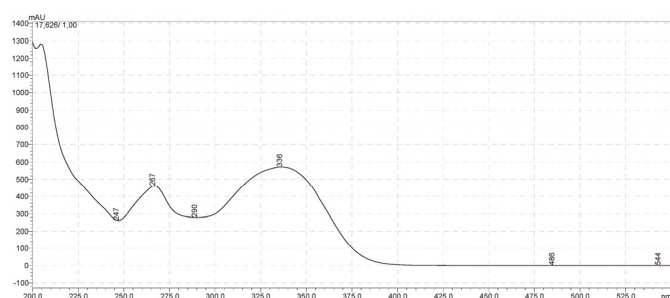


Vanilloyl quinic acids, divanilloyl quinic acids

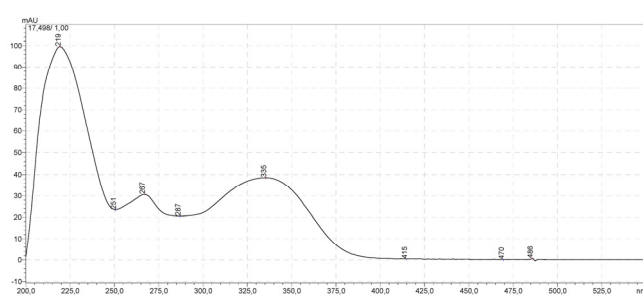


Cichoriin

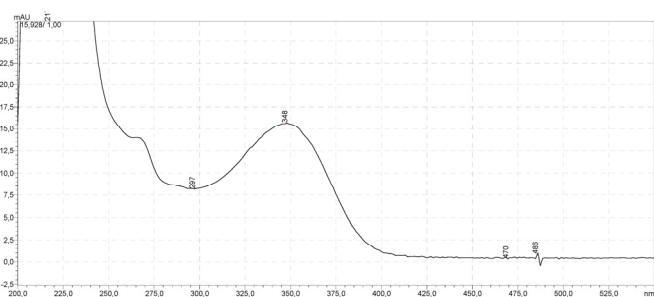
Figure S1. Typical UV spectra of the basic phenolic compounds found in six Asteraceae species.



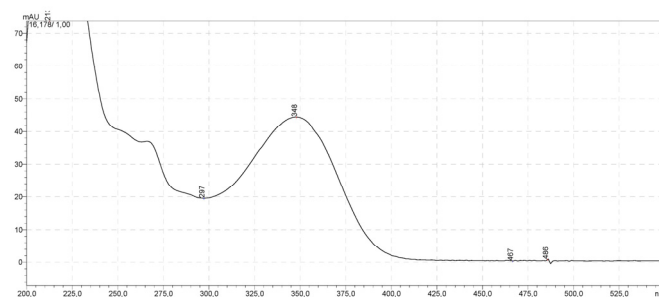
Apigenin 7-O-glycosides



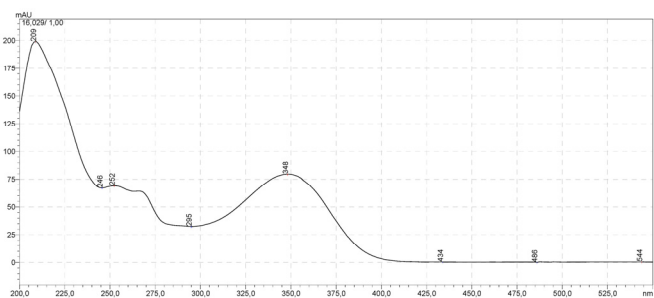
Acacetin/pectolinarigenin 7-O-glycosides



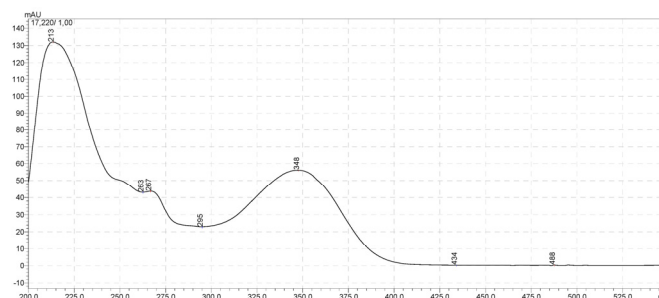
Luteolin 7-O-glycosides



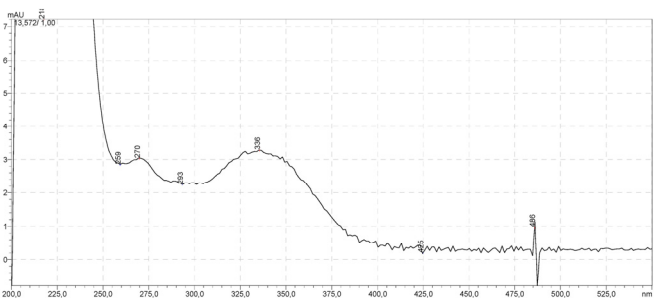
Chrysoeriol 7-O-glycosides



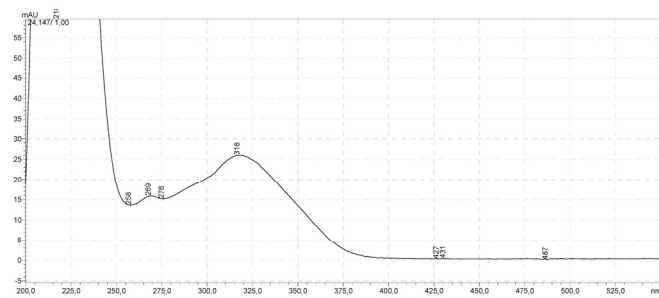
Luteolin 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside



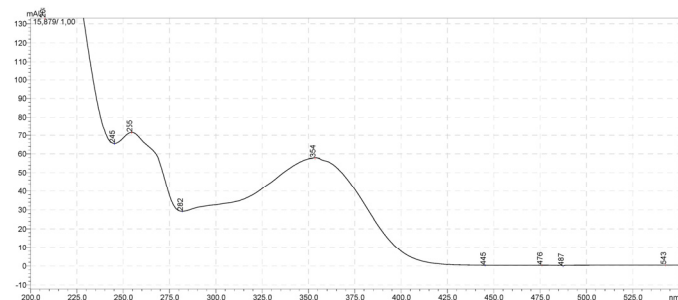
Chrysoeriol 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside



Schaftoside

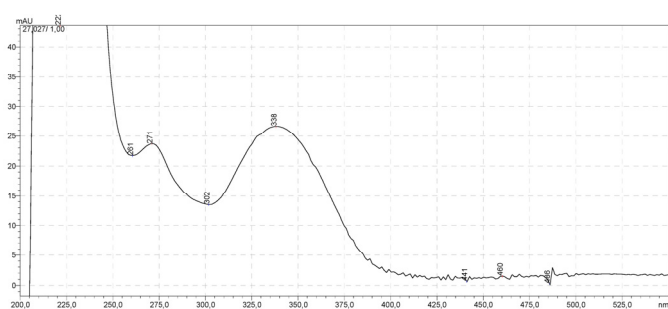


Echitin, echinacin

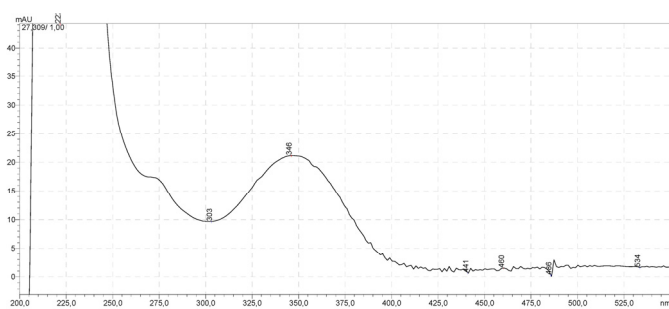


Quercetin/isorhamnetin 3-O-glycosides

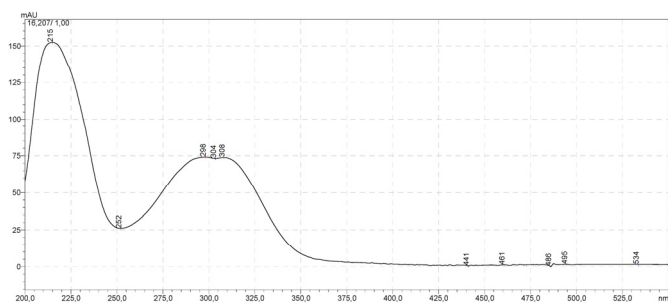
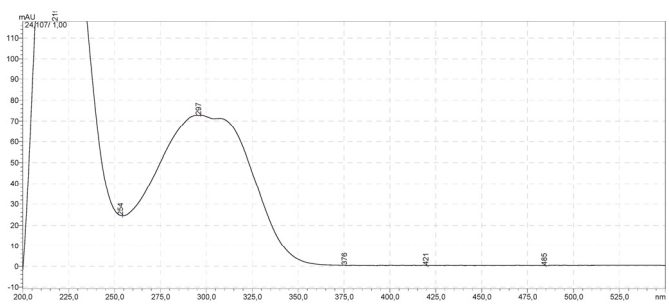
Figure S1. Continuation.



Cirsiliol



Chrysosplenetin

Tri-*p*-coumaroyl spermines / spermidinesTetra-*p*-coumaroyl spermines**Figure S1.** Continuation.

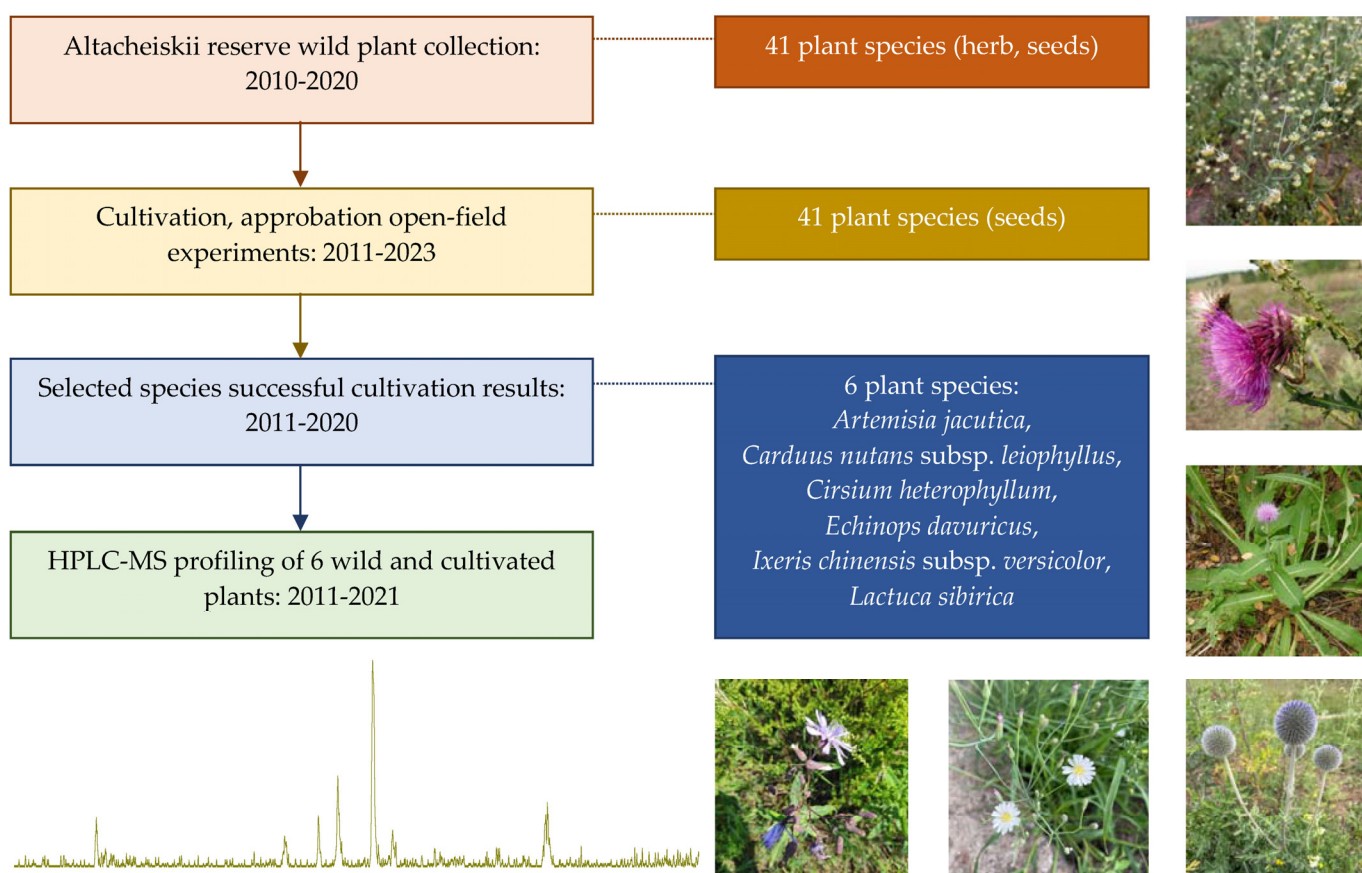


Figure S2. Flow chart for the experimental design.

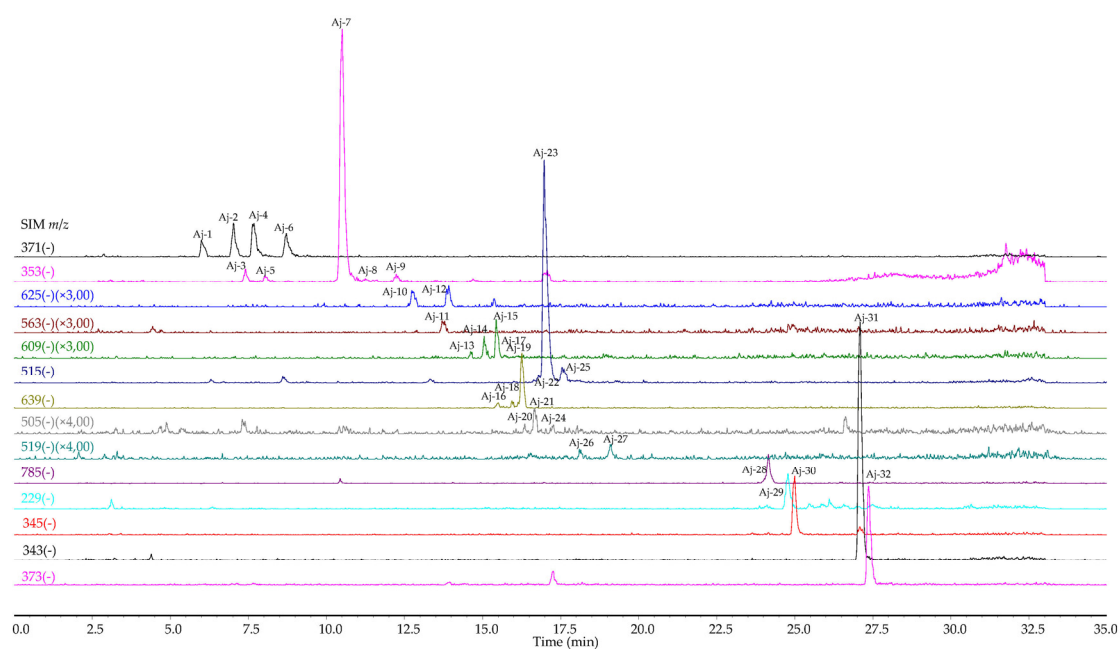


Figure S3. HPLC-MS chromatograms of *Artemisia jacutica* herb (wild sample AJW-1) in SIM mode (negative ionization; SIM m/z values in left panel). Compounds Aj-1–Aj-32 description see in Table S1.

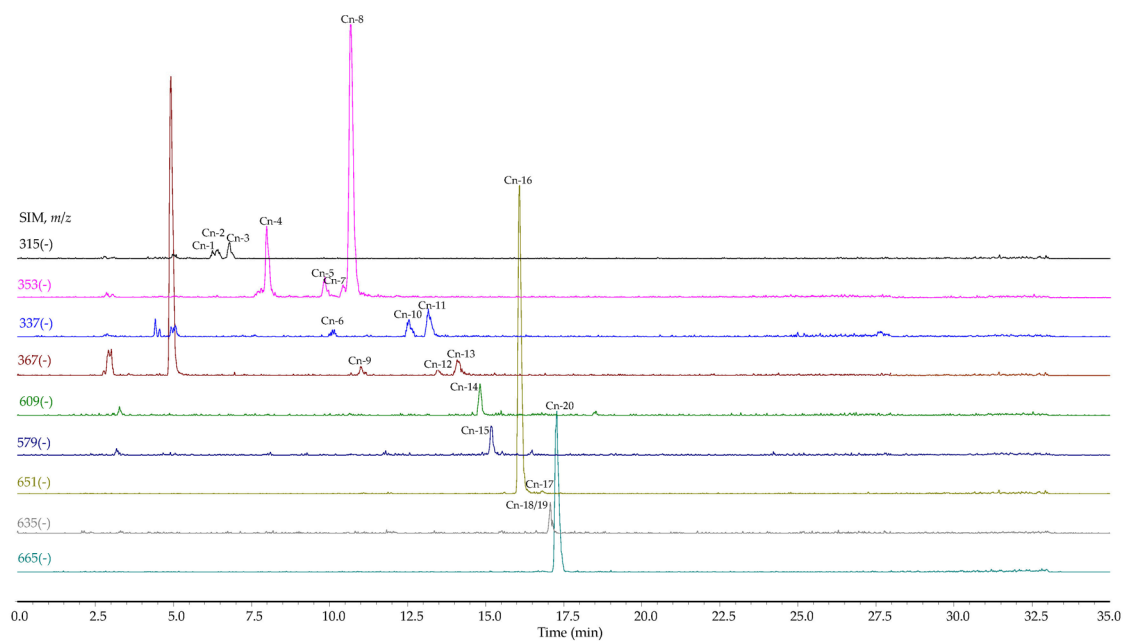


Figure S4. HPLC-MS chromatograms of *Carduus nutans* subsp. *leiophyllus* herb (wild sample CNW-1) in SIM mode (negative ionization; SIM m/z values in left panel). Compounds Cn-1–Cn-20 description see in Table S2.

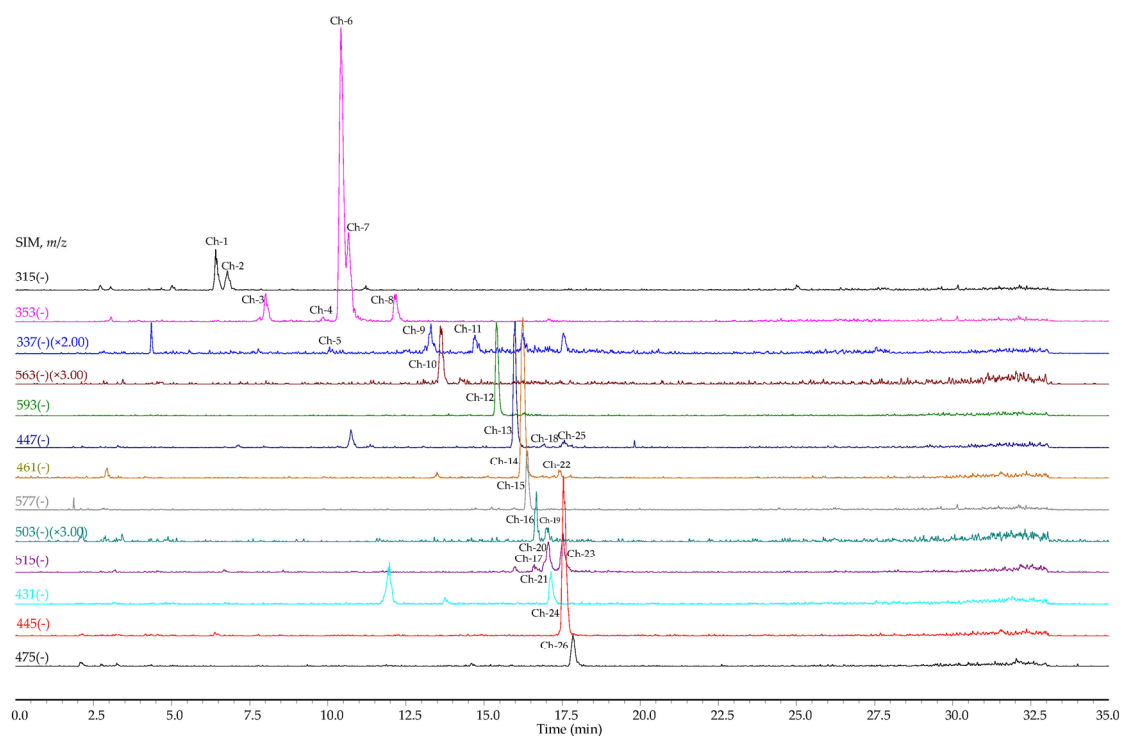


Figure S5. HPLC-MS chromatograms of *Cirsium heterophyllum* herb (wild sample CHW-1) in SIM mode (negative ionization; SIM m/z values in left panel). Compounds Ch-1–Ch-24 description see in Table S4.

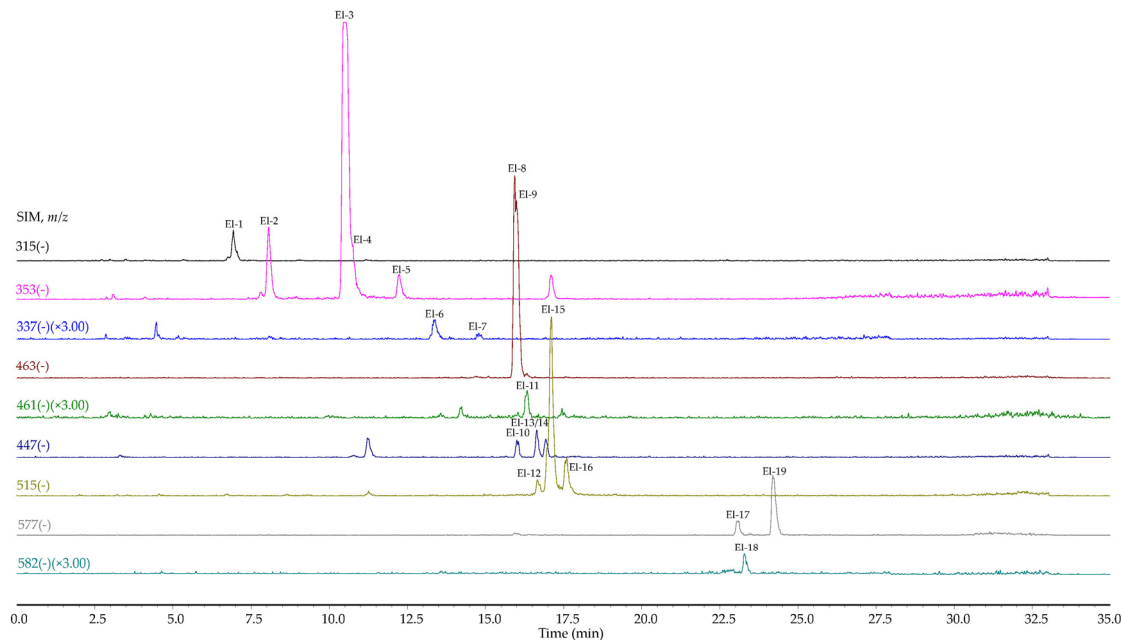


Figure S6. HPLC-MS chromatograms of *Echinops davuricus* herb (wild sample ELW-1) in SIM mode (negative ionization; SIM m/z values in left panel). Compounds EI-1–EI-19 description see in Table S5.

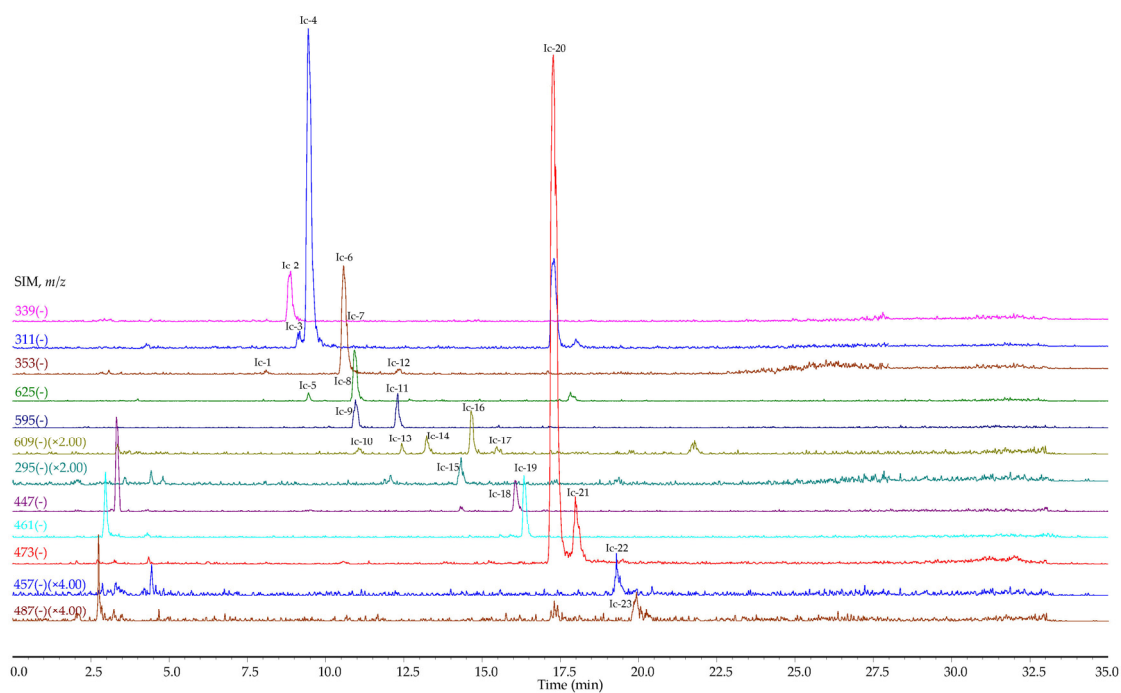


Figure S7. HPLC-MS chromatograms of *Ixeris chinensis* subsp. *versicolor* herb (wild sample ICW-1) in SIM mode (negative ionization; SIM m/z values in left panel). Compounds Ic-1–Ic-23 description see in Table S6.

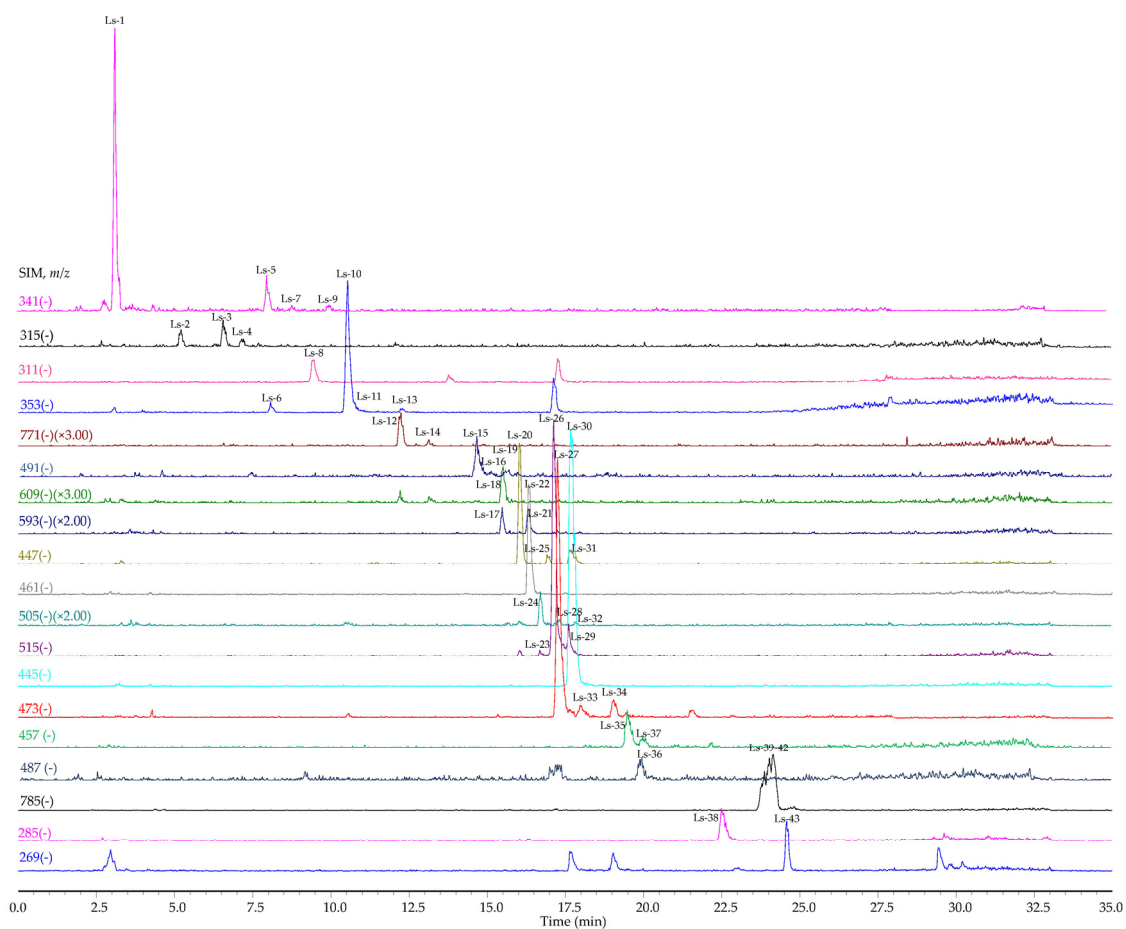


Figure S8. HPLC-MS chromatograms of *Lactuca sibirica* herb (wild sample LSW-1) in SIM mode (negative ionization; SIM m/z values in left panel). Compounds Ls-1–Ls-41 description see in Table S7.

Table S1. Description of wild samples of Asteraceae species collected in Altacheiskii reserve (<https://baikalzapovednik.ru/altacheisky#rec54745054>; accessed on 10 April 2024) for a period 2010–2020 and information on cultivation success.

No	Species	Seed and Herb Collection Year	Approbations Years	Cultivation Results*
AAW-1	<i>Achillea asiatica</i> Serg.	2010	2011, 2012, 2013	Ambiguously
ADW-1	<i>Antennaria dioica</i> (L.) Gaertn.	2014	2015, 2016	Failed
AGW-1	<i>Arctogeron gramineum</i> (L.) DC	2012	2013, 2014, 2015	Ambiguously
ANW-1	<i>Artemisia anethifolia</i> Web. ex Stechm.	2015	2016, 2017, 2018	Ambiguously
AEW-1	<i>Artemisia desertorum</i> Spreng.	2010	2011, 2012, 2013	Ambiguously
AFW-1	<i>Artemisia frigida</i> Willd.	2011	2012, 2014	Ambiguously
AIW-1	<i>Artemisia integrifolia</i> L.	2017	2018, 2019	Ambiguously
AJW-1	<i>Artemisia jacutica</i> Drobow	2016	2017, 2019, 2021	2-year cultivation success
APW-1	<i>Artemisia palustris</i> L.	2019	2020, 2021, 2022	Ambiguously
AUW-1	<i>Artemisia pubescens</i> var. <i>pubescens</i>	2020	2021, 2022	Ambiguously
ASW-1	<i>Artemisia sericea</i> Weber ex Stechm.	2020	2021, 2022	Ambiguously
ATW-1	<i>Artemisia tanacetifolia</i> L.	2010	2011, 2012, 2013	Ambiguously
ALW-1	<i>Aster alpinus</i> L.	2020	2021, 2022	Ambiguously
ABW-1	<i>Aster biennis</i> Ledeb.	2011	2012, 2013	Ambiguously
ARW-1	<i>Aster tataricus</i> L.	2010	2011, 2012, 2013	Ambiguously
CHW-1	<i>Cacalia hastata</i> L.	2018	2019, 2020	Ambiguously
CNW-1	<i>Carduus nutans</i> subsp. <i>leiophyllus</i> (Petrovič) Arènes	2015	2016, 2018, 2020	2-year cultivation success
CEW-1	<i>Cirsium esculentum</i> (Siev.) C.A. Mey.	2012	2013, 2014, 2015	Ambiguously
CHW-1	<i>Cirsium heterophyllum</i> (L.) Hill	2014	2015	6-year cultivation success
CSW-1	<i>Cirsium serratuloides</i> (L.) Hill	2019	2020, 2022	Ambiguously
CBW-1	<i>Crepis bungei</i> Ledeb.	2012	2013, 2014, 2015	Ambiguously
CTW-1	<i>Crepis tectorum</i> L.	2015	2016, 2017	Ambiguously
EDW-1	<i>Echinops davuricus</i> Fisch. ex Hornem.	2010	2011	9-year cultivation success
EAW-1	<i>Erigeron acris</i> L.	2017	2018, 2019	Ambiguously
FSW-1	<i>Filifolium sibiricum</i> (L.) Kitam.	2011	2012, 2013	Failed
GDW-1	<i>Galatella davurica</i> DC.	2014	2015, 2016, 2017	Failed
ICW-1	<i>Ixeris chinensis</i> subsp. <i>versicolor</i> (Fisch. ex Link) Kitam.	2014	2015	4-year cultivation success
LSW-1	<i>Lactuca sibirica</i> (L.) Benth. ex Maxim.	2014	2015	5-year cultivation success
LAW-1	<i>Leibnitzia anandria</i> (L.) Nakai	2018	2019, 2020	Failed
NPW-1	<i>Neopallasia pectinata</i> Poljak.	2020	2021, 2022, 2023	Ambiguously
PBW-1	<i>Pentanema britannica</i> (L.) D.Gut.Larr., Santos-Vicente, Anderb., E.Rico & M.M.Mart.Ort.	2014	2015, 2016, 2017	Ambiguously
PDW-1	<i>Picris davurica</i> Fisch.	2017	2018, 2019, 2020	Ambiguously
SAW-1	<i>Saussurea amara</i> (L.) DC.	2020	2021, 2022	Ambiguously
SSW-1	<i>Saussurea salicifolia</i> (L.) DC.	2010	2011, 2012	Ambiguously
SUW-1	<i>Scorzonera austriaca</i> Willd.	2018	2019, 2020	Ambiguously
SEW-1	<i>Senecio erucifolius</i> L.	2017	2018, 2019	Ambiguously
SDW-1	<i>Solidago dahurica</i> (Kitag.) Kitag. ex Juz.	2015	2016, 2017, 2019	Ambiguously
TLW-1	<i>Taraxacum leucanthum</i> (Ledeb.) Ledeb.	2014	2015, 2016, 2017	Failed
TIW-1	<i>Tephroseris integrifolia</i> (L.) Holub	2019	2020, 2021, 2022	Ambiguously
TSW-1	<i>Tephroseris subdentata</i> (Bunge) Holub	2011	2012, 2013, 2014	Ambiguously
YTW-1	<i>Youngia tenuifolia</i> (Willd.) Bab. et Stebb.	2015	2016, 2017, 2018	Ambiguously

* Ambiguously—mixed results, more research needed. Failed—failed to achieve successful cultivation.

Table S2. Plant cultivation conditions of six Asteraceae species.

Species	Experi- mental Field Size, m×m	Experi- mental Fields, n (No.)	Distance Be- tween Plants, m	Soil Description	Climatic Data
<i>Artemisia jacutica</i>	50 × 60	3 (24-1b-1, 24- 1b-2, 24-1b-3)	0.6	Soil type: Dark humus. Fertile Layer: 70–90 cm. Soil P ₂ O ₅ content: 22–24 mg/100 g. Soil K ₂ O content: 60–64 mg/100 g. pH: 7.4. Humus content: 8–9%. To- tal N: 0.5–0.6%. C : N Ratio: 10–11. No organic/inor- ganic fertilizers used.	Average annual precipitation: 230–250 mm. Frost-free pe- riod: 110–116 days. Sum of temperatures above 10°C: 1900–2000.
<i>Carduus nutans</i> subsp. <i>leiophyllus</i>	70 × 70	4 (24-1b-7, 24- 1b-8, 24-1b-9, 24-1b-10)	1.2		
<i>Cirsium heterophyllum</i>	50 × 40	3 (24-1b-15, 24-1b-16, 24-1b-17)	0.6		
<i>Echinops davuricus</i>	70 × 90	3 (24-1b-27, 24-1b-28, 24-1b-29)	0.5		
<i>Ixeris chinensis</i> subsp. <i>versicolor</i>	40 × 40	4 (24-1b-35, 24-1b-36, 24-1b-37, 24-1b-38)	0.3		
<i>Lactuca sibirica</i>	40 × 50	5 (24-1b-52, 24-1b-53, 24-1b-54, 24-1b-55, 24-1b-56)	0.4		

Table S3. Plant sample description for HPLC assay.

Species	Experimental Field No	Year of Collection (plant age, years; sample numbers = repetition, one dry sample weight, kg)
<i>Artemisia jacutica</i>	24-1b-1	2021 (1; 5/0.1), 2022 (2; 5/0.5)
	24-1b-2	2021 (1; 5/0.1), 2022 (2; 5/0.7)
	24-1b-3	2021 (1; 5/0.1), 2022 (2; 5/0.6)
<i>Carduus nutans</i> subsp. <i>leiophyllus</i>	24-1b-7	2020 (1; 5/0.8), 2021 (2; 5/2.5)
	24-1b-8	2020 (1; 5/0.7), 2021 (2; 5/2.0)
	24-1b-9	2020 (1; 5/0.7), 2021 (2; 5/3.5)
	24-1b-10	2020 (1; 5/0.9), 2021 (2; 5/1.5)
<i>Cirsium heterophyllum</i>	24-1b-15	2015 (1; 5/0.2), 2016 (2; 5/0.7), 2018 (4; 5/1.4), 2020 (6; 5/2.9)
	24-1b-16	2015 (1; 5/0.1), 2016 (2; 5/0.6), 2018 (4; 5/1.2), 2020 (6; 5/2.4)
	24-1b-17	2015 (1; 5/0.2), 2016 (2; 5/0.5), 2018 (4; 5/1.2), 2020 (6; 5/2.5)
<i>Echinops davuricus</i>	24-1b-27	2011 (1; 4/0.3), 2013 (3; 4/0.9), 2015 (5; 4/1.6), 2017 (7; 4/1.7), 2019 (9; 4/1.5)
	24-1b-28	2011 (1; 4/0.2), 2013 (3; 4/0.8), 2015 (5; 4/1.2), 2017 (7; 4/1.4), 2019 (9; 4/1.5)
	24-1b-29	2011 (1; 4/0.4), 2013 (3; 4/0.8), 2015 (5; 4/1.4), 2017 (7; 4/1.4), 2019 (9; 4/1.5)
<i>Ixeris chinensis</i> subsp. <i>versicolor</i>	24-1b-35	2015 (1; 6/0.1), 2016 (2; 6/0.2), 2017 (3; 6/0.3), 2018 (4; 6/0.3)
	24-1b-36	2015 (1; 6/0.08), 2016 (2; 6/0.2), 2017 (3; 6/0.3), 2018 (4; 6/0.3)
	24-1b-37	2015 (1; 6/0.1), 2016 (2; 6/0.2), 2017 (3; 6/0.3), 2018 (4; 6/0.3)
	24-1b-38	2015 (1; 6/0.1), 2016 (2; 6/0.2), 2017 (3; 6/0.3), 2018 (4; 6/0.3)
<i>Lactuca sibirica</i>	24-1b-52	2015 (1; 5/0.1), 2016 (2; 5/0.2), 2017 (3; 5/0.4), 2019 (5; 5/0.3)
	24-1b-53	2015 (1; 5/0.1), 2016 (2; 5/0.15), 2017 (3; 5/0.3), 2019 (5; 5/0.3)
	24-1b-54	2015 (1; 5/0.1), 2016 (2; 5/0.2), 2017 (3; 5/0.3), 2019 (5; 5/0.3)
	24-1b-55	2015 (1; 5/0.1), 2016 (2; 5/0.17), 2017 (3; 5/0.4), 2019 (5; 5/0.3)
	24-1b-56	2015 (1; 5/0.1), 2016 (2; 5/0.2), 2017 (3; 5/0.4), 2019 (5; 5/0.3)

Table S4. Reference standards used for HPLC profiling and quantification.

Analyzed Compound	Reference Compound. HPLC-Profiling	Reference Compound. Quantification
Benzoates		
Protocatechuic acid <i>O</i> -hexosides	Protocatechuic acid 4- <i>O</i> -β-glucoside. ChemFaces. Cat. No.: CFN95557. Pu- rity ≥98%	Protocatechuic acid 4- <i>O</i> -β-glucoside. ChemFaces. Cat. No.: CFN95557. Pu- rity ≥98%
Protocatechuic acid 4- <i>O</i> -glucoside		
Vanilloyl quinic acid (probably 1- <i>O</i> -isomer)	Putative identification. Ref. [40–42]	Vanillic acid 4-β-D-glucoside. Med- ChemExpress. Cat. No.: HY-114760. Purity ≥98%. Conversion factor (differ- ence in MW): 0.97.
Vanilloyl quinic acid (probably 4- <i>O</i> -isomer)		
Vanilloyl quinic acid (probably 5- <i>O</i> -isomer)		
Divanilloyl quinic acid (probably 3,4- <i>O</i> -isomer)		Vanillic acid 4-β-D-glucoside. Med- ChemExpress. Cat. No.: HY-114760. Purity ≥98%. Conversion factor (differ- ence in MW): 0.67.
Divanilloyl quinic acid (probably 3,5- <i>O</i> -isomer)		
Divanilloyl quinic acid (probably 4,5- <i>O</i> -isomer)		
Coumarins		
Esculetin 7- <i>O</i> -glucoside	Cichoriin. ChemFaces. Cat. No.: CFN95196. Purity ≥98%	Cichoriin. ChemFaces. Cat. No.: CFN95196. Purity ≥98%
Hydroxycinnamates		
3- <i>O</i> -Caffeoyl glucaric acid 4- <i>O</i> -Caffeoyl glucaric acid 2- <i>O</i> -Caffeoyl glucaric acid 5- <i>O</i> -Caffeoyl glucaric acid	Laboratory collection. Ref. [43]	<i>trans</i> -Caffeoyl-6- <i>O</i> -D-gluconic acid. MedChemExpress. Cat. No.: HY- N11466. Purity ≥98%. Conversion fac- tor (difference in MW): 0.96.
Caftaric acid isomer Caftaric acid	Caftaric acid. ChemFaces. Cat. No.: CFN00384. Purity ≥98%	Caftaric acid. ChemFaces. Cat. No.: CFN00384. Purity ≥98%
Cichoric acid (<i>di-trans</i> isomer) Cichoric acid (<i>cis-trans</i> isomer) Cichoric acid (<i>cis-cis</i> isomer)	Cichoric acid. ChemFaces. Cat. No.: CFN99725. Purity ≥98%	Cichoric acid. ChemFaces. Cat. No.: CFN99725. Purity ≥98%
Coutaric acid	Coutaric acid. SLS Scientific Labora- tory Supplies. Cat. No.: SIGPHL83843. Purity ≥96%	Coutaric acid. SLS Scientific Labora- tory Supplies. Cat. No.: SIGPHL83843. Purity ≥96%
<i>p</i> -Coumaroyl-caffeoyl-tartaric acids	Putative identification. Ref. [44,45]	Cichoric acid. ChemFaces. Cat. No.: CFN99725. Purity ≥98%. Conversion factor (difference in MW): 1.04.
Feruloyl-caffeoyl-tartaric acid		Cichoric acid. ChemFaces. Cat. No.: CFN99725. Purity ≥98%. Conversion factor (difference in MW): 0.81.
3- <i>O-p</i> -Coumaroyl quinic acid	3- <i>O-p</i> -Coumaroylquinic acid. Sigma- Aldrich. Cat. No.: 935557. Purity ≥95%	3- <i>O-p</i> -Coumaroylquinic acid. Sigma- Aldrich. Cat. No.: 935557. Purity ≥95%
4- <i>O-p</i> -Coumaroyl quinic acid	4- <i>O</i> -Coumaroylquinic acid. Chem- Faces. Cat. No.: CFN95508. Purity ≥98%	4- <i>O</i> -Coumaroylquinic acid. Chem- Faces. Cat. No.: CFN95508. Purity ≥98%
5- <i>O-p</i> -Coumaroyl quinic acid	5- <i>O</i> -Coumaroylquinic acid. Chem- Faces. Cat. No.: CFN95507. Purity ≥98%	5- <i>O</i> -Coumaroylquinic acid. Chem- Faces. Cat. No.: CFN95507. Purity ≥98%
3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	Chlorogenic acid. MedChemExpress. Cat. No.: HY-N0055. Purity ≥99.35%.	Chlorogenic acid. MedChemExpress. Cat. No.: HY-N0055. Purity ≥99.35%.
4- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	Cryptochlorogenic acid. Med- ChemExpress. Cat. No.: HY-N0787. Purity ≥99.88%.	Cryptochlorogenic acid. Med- ChemExpress. Cat. No.: HY-N0787. Purity ≥99.88%.
4- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)		
5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	Neochlorogenic acid. Med- ChemExpress. Cat. No.: HY-N0722. Purity ≥99.07%.	Neochlorogenic acid. Med- ChemExpress. Cat. No.: HY- N0722. Purity ≥99.07%.
5- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	<i>Cis</i> -5-Caffeoylquinic acid. BenchChem. Cat. No.: B1222021. Purity ≥98%.	<i>Cis</i> -5-Caffeoylquinic acid. BenchChem. Cat. No.: B1222021. Purity ≥98%.
3,4-Di- <i>O</i> -caffeoyl quinic acid	3,4-Dicaffeoylquinic acid. Med- ChemExpress. Cat. No.: HY-N0057. Purity ≥98.64%.	3,4-Dicaffeoylquinic acid. Med- ChemExpress. Cat. No.: HY-N0057. Purity ≥98.64%.
3,5-Di- <i>O</i> -caffeoyl quinic acid	3,5-Di- <i>O</i> -caffeoylquinic acid. Chem- Faces. Cat. No.: CFN93712. Purity ≥98%	3,5-Di- <i>O</i> -caffeoylquinic acid. Chem- Faces. Cat. No.: CFN93712. Purity ≥98%

4,5-Di-O-caffeoyl quinic acid	4,5-Di-O-caffeoyl quinic acid. Med-ChemExpress. Cat. No.: HY-N11077. Purity ≥98%.	4,5-Di-O-caffeoyl quinic acid. Med-ChemExpress. Cat. No.: HY-N11077. Purity ≥98%.
3-O-Feruloyl quinic acid	3-O-Feruloyl quinic acid. ChemFaces. Cat. No.: CFN92393. Purity ≥98%.	3-O-Feruloyl quinic acid. ChemFaces. Cat. No.: CFN92393. Purity ≥98%.
4-O-Feruloyl quinic acid	4-O-Feruloyl quinic acid. ChemFaces. Cat. No.: CFN92392. Purity ≥98%.	4-O-Feruloyl quinic acid. ChemFaces. Cat. No.: CFN92392. Purity ≥98%.
5-O-Feruloyl quinic acid	5-O-Feruloyl quinic acid. ChemFaces. Cat. No.: CFN92889. Purity ≥98%.	5-O-Feruloyl quinic acid. ChemFaces. Cat. No.: CFN92889. Purity ≥98%.
<i>Flavonoids</i>		
Apigenin 7-O-glucoside	Apigenin 7-O-glucoside. Extrasynthese. Cat. No.: 1004 S. Purity ≥99%.	Apigenin 7-O-glucoside. Extrasynthese. Cat. No.: 1004 S. Purity ≥99%.
Apigenin 7-O-glucuronide	Apigenin 7-O-glucuronide. Med-ChemExpress. Cat. No.: HY-N1454. Purity ≥98.83%.	Apigenin 7-O-glucuronide. Med-ChemExpress. Cat. No.: HY-N1454. Purity ≥98.83%.
Apigenin 7-O-rutinoside	Isorhoifolin. Cayman Chemical. Cat. No.: 34036. Purity ≥95%	Isorhoifolin. Cayman Chemical. Cat. No.: 34036. Purity ≥95%
Apigenin 7-O-(2''-O- <i>p</i> -coumaroyl)-glucoside	Laboratory collection. Ref. [46]	Apigenin 4'-O-(2'',6''-di-O- <i>E-p</i> -coumaroyl)glucoside. ChemFaces. Cat. No.: CFN95276. Purity ≥98%. Conversion factor (difference in MW): 1.25.
Apigenin 7-O-(6''-O- <i>p</i> -coumaroyl)-glucoside		
Apigenin di-O-hexoside-O-acetates	Putative identification. Ref. [47]	Apigenin 7-O-glucoside. Extrasynthese. Cat. No.: 1004 S. Purity ≥99%. Conversion factor (difference in MW): 0.68.
Apigenin 6-C-glucoside-8-C-arabinoside	Schaftoside. ChemFaces. Cat. No.: CFN90197. Purity ≥98%.	Schaftoside. ChemFaces. Cat. No.: CFN90197. Purity ≥98%.
Acacetin 7-O-glucoside	Tilianin. ChemFaces. Cat. No.: CFN92764. Purity ≥98%.	Tilianin. ChemFaces. Cat. No.: CFN92764. Purity ≥98%.
Luteolin 5-O-glucoside	Luteolin 5-O-glucoside. Med-ChemExpress. Cat. No.: HY-N2008. Purity ≥97.18%.	Luteolin 5-O-glucoside. Med-ChemExpress. Cat. No.: HY-N2008. Purity ≥97.18%.
Luteolin 7-O-glucoside	Luteolin 7-glucoside. Sigma-Aldrich. Cat. No.: 49968. Purity ≥98%.	Luteolin 7-glucoside. Sigma-Aldrich. Cat. No.: 49968. Purity ≥98%
Luteolin 3'-O-glucoside	Luteolin 3'-glucoside. Phytolab GmbH & Co. KG. Cat. No.: 85840. Purity ≥98%.	Luteolin 3'-glucoside. Phytolab GmbH & Co. KG. Cat. No.: 85840. Purity ≥98%.
Luteolin 4'-O-glucoside	Luteolin 4'-O-glucoside. Extrasynthese. Cat. No.: 1412 S. Purity ≥98%.	Luteolin 4'-O-glucoside. Extrasynthese. Cat. No.: 1412 S. Purity ≥98%.
Luteolin 7-O-rutinoside	Luteolin 7-rutinoside. Med-ChemExpress. Cat. No.: HY-N6647. Purity ≥98.47%.	Luteolin 7-rutinoside. Med-ChemExpress. Cat. No.: HY-N6647. Purity ≥98.47%.
Luteolin 7-O-(2''-O-glucosyl)-glucoside	Laboratory collection. Ref. [48]	Luteolin 7-glucoside. Sigma-Aldrich. Cat. No.: 49968. Purity ≥98%. Conversion factor (difference in MW): 0.73.
Luteolin 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside	Isolated in present work.	Luteolin 7-glucoside. Sigma-Aldrich. Cat. No.: 49968. Purity ≥98%. Conversion factor (difference in MW): 0.69.
Luteolin O-hexoside-O-pentoside	Putative identification. Ref. [49]	Luteolin 7-glucoside. Sigma-Aldrich. Cat. No.: 49968. Purity ≥98%. Conversion factor (difference in MW): 0.77.
Luteolin di-O-hexoside-O-acetate	Putative identification. Ref. [49]	Luteolin 7-glucoside. Sigma-Aldrich. Cat. No.: 49968. Purity ≥98%. Conversion factor (difference in MW): 0.67.
Luteolin tri-O-hexosides	Putative identification. Ref. [49]	Luteolin 7-glucoside. Sigma-Aldrich. Cat. No.: 49968. Purity ≥98%. Conversion factor (difference in MW): 0.58.
Chrysoeriol 7-O-glucoside	Chrysoeriol 7-O-glucoside. ChemFaces. Cat. No.: CFN93021. Purity ≥98%.	Chrysoeriol 7-O-glucoside. ChemFaces. Cat. No.: CFN93021. Purity ≥98%.
Chrysoeriol 4'-O-glucoside	Laboratory collection. Ref. [50]	Chrysoeriol 7-O-glucoside. ChemFaces. Cat. No.: CFN93021. Purity ≥98%.

Chrysoeriol 7- <i>O</i> -(2''- <i>O</i> -acetyl)-glucoside	Laboratory collection. Ref. [50]	Chrysoeriol 7- <i>O</i> -glucoside. ChemFaces. Cat. No.: CFN93021. Purity ≥98%. Conversion factor (difference in MW): 0.92.
Chrysoeriol 7- <i>O</i> -(6''- <i>O</i> -acetyl)-glucoside		
Chrysoeriol 7- <i>O</i> -(2''- <i>O</i> -(6'''- <i>O</i> -acetyl)-glucosyl)-glucoside	Isolated in present work.	Chrysoeriol 7- <i>O</i> -glucoside. ChemFaces. Cat. No.: CFN93021. Purity ≥98%. Conversion factor (difference in MW): 0.69.
6-Hydroxyluteolin di- <i>O</i> -hexoside	Putative identification. Ref. [51]	6-Hydroxyluteolin 7- <i>O</i> -glucoside. ChemFaces. Cat. No.: CFN91094. Purity ≥98%. Conversion factor (difference in MW): 0.74.
Pectolarigenin 7- <i>O</i> -glucoside	Pectolarigenin 7- <i>O</i> -glucoside. Geopharma Co. Cat. No.: 35-5373-11-5. Purity ≥98%.	Pectolarigenin 7- <i>O</i> -glucoside. Geopharma Co. Cat. No.: 35-5373-11-5. Purity ≥98%.
Kaempferol 3- <i>O</i> -neohesperidoside	Kaempferol 3- <i>O</i> -neohesperidoside. MedChemExpress. Cat. No.: HY-107207. Purity ≥99.86%.	Kaempferol 3- <i>O</i> -neohesperidoside. MedChemExpress. Cat. No.: HY-107207. Purity ≥99.86%.
Kaempferol 3- <i>O</i> -rutinoside	Nicotiflorin. Selleck Chemicals. Cat. No.: S3267. Purity ≥99.91%.	Nicotiflorin. Selleck Chemicals. Cat. No.: S3267. Purity ≥99.91%.
Kempferol 3- <i>O</i> -sophoroside	Kempferol 3- <i>O</i> -sophoroside. MedChemExpress. Cat. No.: HY-N2055. Purity ≥99.84%.	Kempferol 3- <i>O</i> -sophoroside. MedChemExpress. Cat. No.: HY-N2055. Purity ≥99.84%.
Kaempferol 3- <i>O</i> -gentiobioside	Kaempferol 3- <i>O</i> -gentiobioside. ChemFaces. Cat. No.: CFN92384. Purity ≥98%.	Kaempferol 3- <i>O</i> -gentiobioside. ChemFaces. Cat. No.: CFN92384. Purity ≥98%.
Kaempferol 7- <i>O</i> -glucoside	Kaempferol 7- <i>O</i> -glucoside. Cayman Chemical. Cat. No.: CAY34347. Purity ≥98%.	Kaempferol 7- <i>O</i> -glucoside. Cayman Chemical. Cat. No.: CAY34347. Purity ≥98%.
Kaempferol di- <i>O</i> -hexoside	Putative identification. Ref. [52]	Kempferol 3- <i>O</i> -sophoroside. MedChemExpress. Cat. No.: HY-N2055. Purity ≥99.84%.
Quercetin 3- <i>O</i> -glucoside	Isoquercitrin. Sigma-Aldrich. Cat. No.: 00140585. Purity ≥95%.	Isoquercitrin. Sigma-Aldrich. Cat. No.: 00140585. Purity ≥95%.
Quercetin 3- <i>O</i> -galactoside	Quercetin 3- <i>O</i> -galactoside. Cayman Chemical. Cat. No.: 18648. Purity ≥98%.	Quercetin 3- <i>O</i> -galactoside. Cayman Chemical. Cat. No.: 18648. Purity ≥98%.
Quercetin 3- <i>O</i> -(2''- <i>O</i> -arabinosyl)-glucoside	Putative identification. Ref. [53]	Isoquercitrin. Sigma-Aldrich. Cat. No.: 00140585. Purity ≥95%. Conversion factor (difference in MW): 0.78.
Quercetin 3- <i>O</i> -(6''- <i>O</i> -arabinosyl)-glucoside		
Quercetin 3- <i>O</i> -neohesperidoside	Quercetin 3- <i>O</i> -neohesperidoside. MedChemExpress. Cat. No.: HY-N7976. Purity ≥99.54%.	Quercetin 3- <i>O</i> -neohesperidoside. MedChemExpress. Cat. No.: HY-N7976. Purity ≥99.54%.
Quercetin 3- <i>O</i> -rungsoside	Laboratory collection. Ref. [54]	Rutin. Sigma-Aldrich. Cat. No.: R5143. Purity ≥94%.
Quercetin 3- <i>O</i> -(4''- <i>O</i> -rhamnosyl)-glucoside		
Quercetin 3- <i>O</i> -rutinoside	Rutin. Sigma-Aldrich. Cat. No.: R5143. Purity ≥94%.	Rutin. Sigma-Aldrich. Cat. No.: R5143. Purity ≥94%.
Quercetin 3- <i>O</i> -sophoroside	Quercetin 3- <i>O</i> -sophoroside. ChemFaces. Cat. No.: CFN90630. Purity ≥98%.	Quercetin 3- <i>O</i> -sophoroside. ChemFaces. Cat. No.: CFN90630. Purity ≥98%.
Quercetin 3- <i>O</i> -gentiobioside	Quercetin 3- <i>O</i> -gentiobioside. MedChemExpress. Cat. No.: HY-N4089. Purity ≥99.92%.	Quercetin 3- <i>O</i> -gentiobioside. MedChemExpress. Cat. No.: HY-N4089. Purity ≥99.92%.
Quercetin 3- <i>O</i> -(2''- <i>O</i> -acetyl)-glucoside	Laboratory collection. Ref. [55]	Quercetin 3- <i>O</i> -(6-acetylglucoside). Extrasynthese. Cat. No.: 1099. Purity ≥85%.
Quercetin 3- <i>O</i> -(3''- <i>O</i> -acetyl)-glucoside		
Quercetin 3- <i>O</i> -(4''- <i>O</i> -acetyl)-glucoside		
Quercetin 3- <i>O</i> -(6''- <i>O</i> -acetyl)-glucoside	Quercetin 3- <i>O</i> -(6-acetylglucoside). Extrasynthese. Cat. No.: 1099. Purity ≥85%.	Quercetin 3- <i>O</i> -(6-acetylglucoside). Extrasynthese. Cat. No.: 1099. Purity ≥85%.
Isorhamnetin 3- <i>O</i> -(2''- <i>O</i> -acetyl)-glucoside	Laboratory collection. Ref. [54]	Isorhamnetin 3- <i>O</i> -glucoside. MedChemExpress. Cat. No.: HY-N0777. Purity ≥99.95%. Conversion factor (difference in MW): 0.92.
Isorhamnetin 3- <i>O</i> -(6''- <i>O</i> -acetyl)-glucoside		

Apigenin	Apigenin. ChemFaces. Cat. No.: CFN98843. Purity ≥98%.	Apigenin. ChemFaces. Cat. No.: CFN98843. Purity ≥98%.
Luteolin	Luteolin. ChemFaces. Cat. No.: CFN98784. Purity ≥98%.	Luteolin. ChemFaces. Cat. No.: CFN98784. Purity ≥98%.
Cirsiliol	Cirsiliol. MedChemExpress. Cat. No.: HY-110399. Purity ≥99.25%.	Cirsiliol. MedChemExpress. Cat. No.: HY-110399. Purity ≥99.25%.
Cirsilineol	Cirsilineol. MedChemExpress. Cat. No.: HY-119347. Purity ≥98%.	Cirsilineol. MedChemExpress. Cat. No.: HY-119347. Purity ≥98%.
Axillarin	Axillarin. ChemFaces. Cat. No.: CFN89530. Purity ≥98%.	Axillarin. ChemFaces. Cat. No.: CFN89530. Purity ≥98%.
Chrysosplenetin	Chrysosplenetin. ChemFaces. Cat. No.: CFN97026. Purity ≥98%.	Chrysosplenetin. ChemFaces. Cat. No.: CFN97026. Purity ≥98%.
<i>Phenylamines</i>		
Tri- <i>O-p</i> -coumaroyl spermines	Putative identification. Ref. [S16]	<i>N</i> 1, <i>N</i> 5, <i>N</i> 10-(<i>E</i>)-Tri- <i>p</i> -coumaroylspermidine. ChemFaces. Cat. No.: CFN95256. Purity ≥98%. Conversion factor (difference in MW): 0.91.
Tetra- <i>O-p</i> -coumaroyl spermines	Putative identification. Ref. [S16]	<i>N</i> 1, <i>N</i> 5, <i>N</i> 10-(<i>E</i>)-Tri- <i>p</i> -coumaroylspermidine. ChemFaces. Cat. No.: CFN95256. Purity ≥98%. Conversion factor (difference in MW): 0.74.
<i>N</i> ¹ , <i>N</i> ⁵ , <i>N</i> ¹⁰ -Tri- <i>O</i> -(<i>EEE</i>)- <i>p</i> -coumaroyl-spermidine	<i>N</i> 1, <i>N</i> 5, <i>N</i> 10-(<i>E</i>)-Tri- <i>p</i> -coumaroylspermidine. ChemFaces. Cat. No.: CFN95256. Purity ≥98%.	<i>N</i> 1, <i>N</i> 5, <i>N</i> 10-(<i>E</i>)-Tri- <i>p</i> -coumaroylspermidine. ChemFaces. Cat. No.: CFN95256. Purity ≥98%.

Table S5. HPLC and mass-spectral conditions used for metabolite separation and identification.

Mode No, SPE Eluate	Column	Column temp., °C	Eluents A/B Com- position	Gradient Program, %B	Flow Rate, µL/min
Mode 1 (for <i>Artemisia jacutica</i> , <i>Carduus nutans</i> subsp. <i>leiophyllum</i> , <i>Cirsium heterophyllum</i> , <i>Echinops davuricus</i> , <i>Ixeris chinensis</i> subsp. <i>versicolor</i>)	ReproSil-Pur 120 C18-AQ (4.6 × 250 mm, 5 µm; Dr. Maisch GmbH, Ammerbuch, Germany)	30	A: 0.1% HCOOH in water; B: 0.1% HCOOH in MeCN	0–20 min 2–80% B, 20–30 min 80–100% B, 30–35 min 100% B, 35–40 min 100–2% B	150
Mode 2 (for <i>Lactuca sibirica</i>)	ReproSil-Pur 120 C18-AQ (4.6 × 250 mm, 5 µm; Dr. Maisch GmbH, Ammerbuch, Germany)	25	A: 0.1% HCOOH in water; B: 0.1% HCOOH in MeCN	0–10 min 2–54% B, 10–20 min 54–75% B, 20–30 min 75– 100% B, 30–35 min 100% B, 35–40 min 100–2% B	150
	Injection Volume, µL	UV-Vis spectral range, nm	ESI Mode	Temperature ESI Interface / Desolvation Line / Heat Block, °C	
Mode 1 (for <i>Artemisia jacutica</i> , <i>Carduus nutans</i> subsp. <i>leiophyllum</i> , <i>Cirsium heterophyllum</i> , <i>Echinops davuricus</i> , <i>Ixeris chinensis</i> subsp. <i>versicolor</i>)	10	200–600	Negative	300 / 200 / 400	
Mode 2 (for <i>Lactuca sibirica</i>)	10	200–600	Negative	300 / 200 / 400	
	Flow Rate Nebulizing Gas (N ₂) / Heating Gas (air) / Collision-Induced Dissociation Gas (Ar), µL/min	Source Voltage, kV	Collision Energy, eV	MS Scanning Range, m/z	
Mode 1 (for <i>Artemisia jacutica</i> , <i>Carduus nutans</i> subsp. <i>leiophyllum</i> , <i>Cirsium heterophyllum</i> , <i>Echinops davuricus</i> , <i>Ixeris chinensis</i> subsp. <i>versicolor</i>)	3 / 10 / 0.3	3	–10–35	80–2000	
Mode 2 (for <i>Lactuca sibirica</i>)	3 / 10 / 0.3	3	–10–35	80–2000	

Table S6. Regression equations, correlation coefficients (r^2), standard deviation (S_{YX}), limits of detection (LOD), limits of quantification (LOQ) and linear ranges for 55 reference standards.

Compound	Ionization ^a	CE ^b (eV)	Regression equation ^c		r^2	S_{YX}	LOD/ LOQ ($\mu\text{g/mL}$)	Linear range ($\mu\text{g/mL}$)
			a	$b \cdot 10^6$				
Protocatechuic acid 4- <i>O</i> - β -glucoside	N	-30	0.9114	-0.6312	0.9887	6.37·10 ⁻²	0.23/0.70	0.7–100.0
Vanillic acid 4- β -D-glucoside	N	-15	1.0864	-0.5271	0.9983	1.04·10 ⁻²	0.03/0.10	0.10–500.0
Cichoriin	N	-25	1.3620	-0.0820	0.9961	9.91·10 ⁻²	0.21/0.72	0.8–100.0
<i>trans</i> -Caffeoyl-6- <i>O</i> -D-gluconic acid	N	-35	0.9562	-0.0521	0.9971	7.79·10 ⁻²	0.27/0.82	0.9–100.0
Caftaric acid	N	-10	2.6538	-0.1376	0.9990	1.17·10 ⁻²	0.01/0.04	0.10–100.0
Cichoric acid	N	-15	1.9478	-0.2369	0.9984	1.03·10 ⁻²	0.02/0.05	0.05–250.0
Coutaric acid	N	-10	1.5632	-0.0376	0.9983	5.14·10 ⁻²	0.11/0.33	0.40–100.0
3- <i>O</i> - <i>p</i> -Coumaroylquinic acid	N	-25	1.0627	-1.2531	0.9893	0.62·10 ⁻²	0.02/0.06	0.10–400.0
4- <i>O</i> -Coumaroylquinic acid	N	-20	1.4689	-0.3641	0.9990	5.69·10 ⁻²	0.12/0.38	0.40–400.0
5- <i>O</i> -Coumaroylquinic acid	N	-20	2.6340	-0.2411	0.9973	2.74·10 ⁻²	0.03/0.10	0.10–350.0
Chlorogenic acid	N	-15	2.4176	-1.5647	0.9994	0.40·10 ⁻²	0.005/0.02	0.02–300.0
Cryptochlorogenic acid	N	-15	2.7365	-1.0690	0.9996	0.51·10 ⁻²	0.006/0.02	0.02–300.0
Neochlorogenic acid	N	-15	2.9021	-1.4184	0.9998	0.39·10 ⁻²	0.004/0.01	0.02–300.0
<i>Cis</i> -5-Caffeoylquinic acid	N	-15	5.8022	-0.8040	0.9990	1.14·10 ⁻²	0.007/0.02	0.02–500.0
3,4-Dicaffeoylquinic acid	N	-10	0.1756	-0.0144	0.9967	3.01·10 ⁻²	0.56/1.71	2.00–850.0
3,5-Di- <i>O</i> -caffeoylquinic acid	N	-20	1.4412	-0.6211	0.9930	11.25·10 ⁻²	0.26/0.78	0.80–100.0
4,5-Di- <i>O</i> -caffeoyl quinic acid	N	-15	1.1123	-1.0289	0.9862	0.41·10 ⁻²	0.01/0.04	0.05–400.0
3- <i>O</i> -Feruloyl quinic acid	N	-20	0.9562	-0.0521	0.9971	7.79·10 ⁻²	0.27/0.82	0.90–100.0
4- <i>O</i> -Feruloyl quinic acid	N	-20	1.3387	-0.0284	0.9981	9.50·10 ⁻²	0.23/0.71	0.80–100.0
5- <i>O</i> -Feruloyl quinic acid	N	-10	0.9518	-0.0267	0.9990	1.03·10 ⁻²	0.03/0.10	0.10–100.0
Apigenin 7- <i>O</i> -glucoside	N	-30	1.1269	-0.9560	0.9864	0.93·10 ⁻²	0.02/0.08	0.10–400.0
Apigenin 7- <i>O</i> -glucuronide	N	-15	3.6748	-0.7069	0.9987	0.90·10 ⁻²	0.008/0.02	0.02–400.0
Isorhoifolin	N	-15	4.9634	-0.5047	0.9972	0.63·10 ⁻²	0.004/0.01	0.01–400.0
Apigenin 4'- <i>O</i> -(2'',6''-di- <i>O</i> - <i>E</i> - <i>p</i> -coumaroyl)glucoside	N	-30	2.3312	-0.4563	0.9803	14.92·10 ⁻²	0.21/0.64	0.7–100.0
Apigenin 7- <i>O</i> -glucoside	N	-25	1.1492	-0.6010	0.9980	4.68·10 ⁻²	0.14/0.41	0.50–400.0
Schaftoside	N	-20	1.4412	-0.6211	0.9930	11.25·10 ⁻²	0.26/0.78	0.8–100.0
Tilianin	N	-25	1.3620	-0.0820	0.9961	9.91·10 ⁻²	0.21/0.72	0.8–100.0
Luteolin 5- <i>O</i> -glucoside	N	-25	1.9634	-0.7458	0.9963	2.59·10 ⁻²	0.04/0.14	0.20–350.0
Luteolin 7-glucoside	N	-20	1.8233	-0.7962	0.9975	2.02·10 ⁻²	0.04/0.11	0.20–350.0
Luteolin 3'-glucoside	N	-15	2.0330	-0.2516	0.9992	1.09·10 ⁻²	0.02/0.05	0.05–300.0
Luteolin 4'- <i>O</i> -glucoside	N	-10	0.1537	-0.0189	0.9960	3.83·10 ⁻²	0.82/2.49	2.50–850.0
Luteolin 7-rutinoside	N	-30	1.3722	-0.0829	0.9973	9.93·10 ⁻²	0.24/0.72	0.80–100.0
Chrysoeriol 7- <i>O</i> -glucoside	N	-30	1.3620	-0.0820	0.9961	9.91·10 ⁻²	0.21/0.72	0.80–100.0
6-Hydroxyluteolin 7- <i>O</i> -glucoside	N	-20	1.0634	-0.0933	0.9902	10.01·10 ⁻²	0.31/0.94	1.00–100.0
Pectolinarigenin 7- <i>O</i> -glucoside	N	-10	1.9610	-0.5271	0.9993	0.94·10 ⁻²	0.02/0.05	0.05–250.0
Kaempferol 3- <i>O</i> -neohesperidoside	N	-15	1.1105	-0.3211	0.9937	4.18·10 ⁻²	0.12/0.38	0.40–400.0
Nicotiflorin	N	-30	1.9634	-0.4511	0.9952	9.18·10 ⁻²	0.15/0.46	0.5–100.0
Kempferol 3- <i>O</i> -sophoroside	N	-10	1.4611	-0.0731	0.9982	4.08·10 ⁻²	0.09/0.27	0.30–100.0
Kaempferol 3-gentiobioside	N	-10	1.5330	-0.0863	0.9985	4.15·10 ⁻²	0.09/0.27	0.30–100.0
Kaempferol 7- <i>O</i> -glucoside	N	-20	2.0859	-0.9171	0.9980	6.18·10 ⁻²	0.03/0.09	0.1–100.0
Isoquercitrin	N	-10	1.6278	-0.0428	0.9990	7.11·10 ⁻²	0.14/0.44	0.50–100.0
Quercetin 3-D-galactoside	N	-10	0.2569	-0.0316	0.9973	2.52·10 ⁻²	0.33/1.00	1.00–750.0
Quercetin 3- <i>O</i> -neohesperidoside	N	-20	1.9871	-0.6871	0.9984	5.63·10 ⁻²	0.09/0.28	0.40–400.0
Rutin	N	-20	1.2716	-0.7389	0.9897	9.14·10 ⁻²	0.23/0.72	0.80–400.0
Quercetin 3- <i>O</i> -sophoroside	N	-20	1.6705	-0.4374	0.9988	12.79·10 ⁻²	0.25/0.77	0.8–100.0
Quercetin 3- <i>O</i> -gentiobioside	N	-20	1.2703	-0.7911	0.9814	15.26·10 ⁻²	0.40/1.20	1.5–100.0
Quercetin-3- <i>O</i> -(6-acetylglucoside)	N	-20	1.1103	-0.9217	0.9901	14.33·10 ⁻²	0.42/1.29	1.5–100.0
Isorhamnetin 3- <i>O</i> -glucoside	N	-15	0.9375	-0.7631	0.9982	1.42·10 ⁻²	0.05/0.15	0.20–350.0

Apigenin	N	-20	2.4751	-0.2081	0.9976	2.04·10 ⁻²	0.03/0.10	0.10–350.0
Luteolin	N	-20	1.8267	-0.4160	0.9990	11.73·10 ⁻²	0.21/0.67	0.70–400.0
Cirsiliol	N	-15	1.1541	-0.4691	0.9987	1.06·10 ⁻²	0.03/0.10	0.10–350.0
Cirsilineol	N	-20	2.0384	-0.3640	0.9975	2.02·10 ⁻²	0.03/0.10	0.10–350.0
Axillarin	N	-10	1.2416	-0.3615	0.9901	3.02·10 ⁻²	0.08/0.24	0.3–100.0
Chrysosplenetin	N	-15	7.064	-1.533	0.9992	1.92·10 ⁻²	0.009/0.03	0.03–500.0
N1,N5,N10-(E)-Tri- <i>p</i> -coumaroylspermidine	N	-10	0.9911	-0.0379	0.9988	2.05·10 ⁻²	0.07/0.21	0.3–100.0

^a Ionization mode : N—negative, P—positive. ^b CE—collision energy. ^c Regression equation: $y = a \cdot x + b$.

Table S7. Retention times (t_R), molecular formulas (MF), mass-spectral data for negative ionization (MSD), and identification level (IL) of compounds Aj-1–Aj-32 found in *Artemisia jacobitica* herb.

No	t_R , min	Compound	MF	MSD, m/z	IL
Aj-1	6.00	3- <i>O</i> -Caffeoyl glucaric acid	C ₁₅ H ₁₆ O ₁₁	371, 179	1a
Aj-2	7.01	4- <i>O</i> -Caffeoyl glucaric acid	C ₁₅ H ₁₆ O ₁₁	371, 179	1a
Aj-3	7.41	4- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Aj-4	7.66	2- <i>O</i> -Caffeoyl glucaric acid	C ₁₅ H ₁₆ O ₁₁	371, 179	1a
Aj-5	8.06	4- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Aj-6	8.71	5- <i>O</i> -Caffeoyl glucaric acid	C ₁₅ H ₁₆ O ₁₁	371, 179	1a
Aj-7	10.51	5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Aj-8	11.28	3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Aj-9	12.25	5- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Aj-10	12.77	6-Hydroxyluteolin di- <i>O</i> -hexoside	C ₂₇ H ₃₀ O ₁₇	625, 463, 301	2
Aj-11	13.71	Schaftoside (apigenin 6- <i>C</i> -glucoside-8- <i>C</i> -arabinoside)	C ₂₆ H ₂₈ O ₁₄	563	1b
Aj-12	13.90	Quercetin 3- <i>O</i> -gentiobioside (quercetin 3- <i>O</i> -(6''- <i>O</i> -glucosyl)-glucoside)	C ₂₇ H ₃₀ O ₁₇	625, 463, 301	1b
Aj-13	14.61	Calendoflavobioside (quercetin 3- <i>O</i> -neohesperidoside)	C ₂₇ H ₃₀ O ₁₆	609, 463, 301	1a
Aj-14	15.05	Rutin (quercetin 3- <i>O</i> -rutinoside)	C ₂₇ H ₃₀ O ₁₆	609, 463, 301	1b
Aj-15	15.41	Calendoside II (quercetin 3- <i>O</i> -rungsioside)	C ₂₇ H ₃₀ O ₁₆	609, 463, 301	1a
Aj-16	15.52	Tri- <i>O</i> - <i>p</i> -coumaroyl spermine	C ₃₇ H ₄₄ N ₄ O ₆	639, 493, 347, 163	2
Aj-17	15.92	Calendoside I (quercetin 3- <i>O</i> -(4''- <i>O</i> -rhamnosyl)-glucoside)	C ₂₇ H ₃₀ O ₁₆	609, 463, 301	1a
Aj-18	15.94	Tri- <i>O</i> - <i>p</i> -coumaroyl spermine	C ₃₇ H ₄₄ N ₄ O ₆	639, 493, 347, 163	2
Aj-19	16.26	Tri- <i>O</i> - <i>p</i> -coumaroyl spermine	C ₃₇ H ₄₄ N ₄ O ₆	639, 493, 347, 163	2
Aj-20	16.37	Quercetin 3- <i>O</i> -(2''- <i>O</i> -acetyl)-glucoside	C ₂₃ H ₂₂ O ₁₃	505, 463, 301	1a
Aj-21	16.67	Quercetin 3- <i>O</i> -(6''- <i>O</i> -acetyl)-glucoside	C ₂₃ H ₂₂ O ₁₃	505, 463, 301	1a
Aj-22	16.81	3,4-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Aj-23	16.97	3,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Aj-24	17.27	Quercetin 3- <i>O</i> -(4''- <i>O</i> -acetyl)-glucoside	C ₂₃ H ₂₂ O ₁₃	505, 463, 301	1a
Aj-25	17.57	4,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Aj-26	18.13	Isorhamnetin 3- <i>O</i> -(2''- <i>O</i> -acetyl)-glucoside	C ₂₄ H ₂₄ O ₁₃	519, 477, 315	1a
Aj-27	19.12	Isorhamnetin 3- <i>O</i> -(6''- <i>O</i> -acetyl)-glucoside	C ₂₄ H ₂₄ O ₁₃	519, 477, 315	1a
Aj-28	24.17	Tetra- <i>O</i> - <i>p</i> -coumaroyl spermine	C ₄₆ H ₅₀ N ₄ O ₈	785, 639, 493, 347, 163	2
Aj-29	24.80	Cirsiliol	C ₁₇ H ₁₄ O ₇	329	1b
Aj-30	25.01	Axyllarin	C ₁₇ H ₁₄ O ₈	345	1b
Aj-31	27.10	Cirsilineol	C ₁₈ H ₁₆ O ₇	343	1b
Aj-32	27.37	Chrysosplenetin	C ₁₉ H ₁₈ O ₈	373	1b

* Identification levels: (1) identified compounds after comparison of retention times, UV and mass-spectral data with previously isolated compounds (1a) or commercial reference standard (1b); (2) putatively annotated compounds after comparison of retention times, UV and mass-spectral data with literature data.

Table S8. Retention times (t_R), molecular formulas (MF), mass-spectral data for negative ionization (MSD), and identification level (IL) of compounds Cn-1–Cn-20 found in *Carduus nutans* subsp. *leiophyllus* herb.

No	t_R , min	Compound	MF	MSD, m/z	IL
Cn-1	6.27	Protocatechuic acid <i>O</i> -hexoside (Cn-3 isomer)	C ₁₃ H ₁₆ O ₉	315, 153	2
Cn-2	6.41	Protocatechuic acid <i>O</i> -hexoside (Cn-3 isomer)	C ₁₃ H ₁₆ O ₉	315, 153	2
Cn-3	6.81	Protocatechuic acid 4- <i>O</i> -glucoside	C ₁₃ H ₁₆ O ₉	315, 153	1b
Cn-4	8.01	4- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Cn-5	9.84	Caffeoyl quinic acid (Cn-4/7/8 isomer)	C ₁₆ H ₁₈ O ₉	353, 179	2
Cn-6	10.12	4- <i>O-p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1a
Cn-7	10.44	5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Cn-8	10.70	3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Cn-9	11.02	4- <i>O</i> -Feruloyl quinic acid	C ₁₇ H ₂₀ O ₉	337, 163	1a
Cn-10	12.53	5- <i>O-p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1b
Cn-11	13.17	3- <i>O-p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1b
Cn-12	13.50	5- <i>O</i> -Feruloyl quinic acid	C ₁₇ H ₂₀ O ₉	337, 163	1b
Cn-13	14.08	3- <i>O</i> -Feruloyl quinic acid	C ₁₇ H ₂₀ O ₉	337, 163	1a
Cn-14	14.83	Luteolin 7- <i>O</i> -(2''- <i>O</i> -glucosyl)-glucoside	C ₂₇ H ₃₀ O ₁₆	609, 447, 285	1b
Cn-15	15.15	Luteolin <i>O</i> -hexoside- <i>O</i> -pentoside	C ₂₆ H ₂₈ O ₁₅	579, 447, 285	2
Cn-16	16.08	Luteolin 7- <i>O</i> -(2''- <i>O</i> -(6'''- <i>O</i> -acetyl)-glucosyl)-glucoside (linariifolioside)	C ₂₉ H ₃₂ O ₁₇	651, 609, 447, 285	1c
Cn-17	16.85	Luteolin di- <i>O</i> -hexoside- <i>O</i> -acetate (Cn-16 isomer)	C ₂₉ H ₃₂ O ₁₇	651, 609, 447, 285	2
Cn-18	17.04	Apigenin di- <i>O</i> -hexoside- <i>O</i> -acetate	C ₂₉ H ₃₂ O ₁₆	635, 593, 431, 269	2
Cn-19	17.18	Apigenin di- <i>O</i> -hexoside- <i>O</i> -acetate	C ₂₉ H ₃₂ O ₁₆	635, 593, 431, 269	2
Cn-20	17.30	Chrysoeriol 7- <i>O</i> -(2''- <i>O</i> -(6'''- <i>O</i> -acetyl)-glucosyl)-glucoside	C ₃₀ H ₃₄ O ₁₇	665, 623, 461, 299	1c

* Identification levels: (1) identified compounds after comparison of retention times, UV and mass-spectral data with previously isolated compounds (1a) or commercial reference standard (1b) or isolated in present work (1c); (2) putatively annotated compounds after comparison of retention times, UV and mass-spectral data with literature data.

Table S9. Extraction/chromatography conditions and spectral data of luteolin 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside and chrysoeriol 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside isolated from *Carduus nutans* subsp. *leiophyllus* leaves.

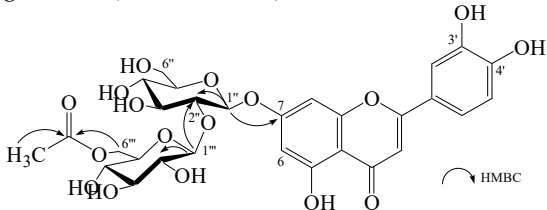
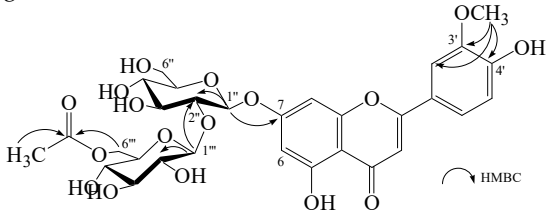
Parameter	Description
Plant extraction / chromatography	Powdered plant material (leaves, sample CNW-1; 5 kg) was extracted with MeOH in a Soxhlet apparatus. The extract was concentrated, which led to the formation of a precipitate (170 g) that was separated over Sephadex LH-20 [column chromatography, eluent EtOH (70%)] and SiO ₂ (column chromatography, eluent hexane–EtOAc, 80:20→60:40) to isolate luteolin 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside (51 g) [69] and chrysoeriol 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside (35 g) [70].
Luteolin 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside (linariifolioside) 	C ₂₉ H ₃₂ O ₁₇ . UV (MeOH, λ _{max} , nm): 252, 268, 348. HR-ESI-MS, <i>m/z</i> 651.3281 [M – H] [–] (calcd. for C ₂₉ H ₃₁ O ₁₇ , 651.5500). ESI-MS, <i>m/z</i> (%): 651 (100) [M – H] [–] , 609 (33) [(M – H) – acetyl] [–] , 447 (25) [(M – H) – acetyl – glucose] [–] , 285 (64) [(M – H) – acetyl – 2 × glucose] [–] . ¹ H NMR (500 MHz, DMSO-d ₆ , 298 K, δ, ppm, J/Hz): luteolin – 6.62 (1H, s; C-3), 6.35 (1H, s; C-6), 6.70 (1H, s; H-8), 7.42 (1H, d, J = 2.0; H-2'), 6.95 (1H, d, J = 8.0; H-5'), 7.45 (1H, dd, J = 2.0, 8.0; H-6'); 7-O-glucose – 5.35 (1H, d, J = 7.0; H-1''), 3.64 (1H, m; H-2''), 3.21–3.48 (6H, m; H-3'', H-4'', H-5'', H-3''', H-4''', H-5'''), 3.52 (1H, m; H-6'' _B), 3.76 (1H, m; H-6'' _A); 2''-O-glucose – 4.73 (1H, d, J = 8.0; H-1'''), 3.01 (1H, m; H-2'''), 3.21–3.48 (6H, m; H-3''', H-4''', H-5''', H-3''''', H-4''''', H-5'''''), 3.80 (1H, m; H-6''' _B), 3.92 (1H, m; H-6''' _A); 6'''-O-acetyl – 1.85 (3H, s; COCH ₃). ¹³ C NMR (125 MHz, DMSO-d ₆ , 298 K, δ, ppm): luteolin – 163.5 (C-2), 104.5 (C-3), 181.7 (C-4), 161.3 (C-5), 99.6 (C-6), 162.5 (C-7), 95.0 (C-8), 153.2 (C-9), 106.3 (C-10), 120.9 (C-1'), 113.0 (C-2'), 146.2 (C-3'), 149.1 (C-4'), 115.6 (C-5'), 119.0 (C-6'); 7-O-glucose – 99.5 (C-1''), 92.5 (C-2''), 75.2 (C-3''), 68.7 (C-4''), 76.9 (C-5''), 60.1 (C-6''); 2''-O-glucose – 104.2 (C-1'''), 74.1 (C-2'''), 75.8 (C-3'''), 69.5 (C-4'''), 73.6 (C-5'''), 62.7 (C-6'''); 6'''-O-acetyl – 20.1 (COCH ₃), 170.5 (COCH ₃).
Chrysoeriol 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside 	C ₃₀ H ₃₄ O ₁₇ . UV (MeOH, λ _{max} , nm): 251, 267, 348. HR-ESI-MS, <i>m/z</i> 665.4217 [M – H] [–] (calcd. for C ₃₀ H ₃₃ O ₁₇ , 665.5770). ESI-MS, <i>m/z</i> (%): 665 (100) [M – H] [–] , 623 (35) [(M – H) – acetyl] [–] , 461 (14) [(M – H) – acetyl – glucose] [–] , 299 (54) [(M – H) – acetyl – 2 × glucose] [–] . ¹ H NMR (500 MHz, DMSO-d ₆ , 298 K, δ, ppm, J/Hz): chrysoeriol – 6.61 (1H, s; C-3), 6.33 (1H, s; C-6), 6.69 (1H, s; H-8), 7.56 (1H, d, J = 2.0; H-2'), 6.92 (1H, d, J = 8.0; H-5'), 7.48 (1H, dd, J = 2.0, 8.0; H-6'), 3.85 (3H, c; 3'-OCH ₃); 7-O-glucose – 5.37 (1H, d, J = 7.0; H-1''), 3.61 (1H, m; H-2''), 3.19–3.49 (6H, m; H-3'', H-4'', H-5'', H-3''', H-4''', H-5'''), 3.53 (1H, m; H-6'' _B), 3.79 (1H, m; H-6'' _A); 2''-O-glucose – 4.78 (1H, d, J = 8.0; H-1'''), 3.03 (1H, m; H-2'''), 3.19–3.49 (6H, m; H-3''', H-4''', H-5''', H-3''''', H-4''''', H-5'''''), 3.81 (1H, m; H-6''' _B), 3.96 (1H, m; H-6''' _A); 6'''-O-acetyl – 1.87 (3H, s; COCH ₃). ¹³ C NMR (125 MHz, DMSO-d ₆ , 298 K, δ, ppm): chrysoeriol – 163.3 (C-2), 104.2 (C-3), 181.5 (C-4), 161.4 (C-5), 99.7 (C-6), 162.6 (C-7), 95.2 (C-8), 153.1 (C-9), 106.2 (C-10), 121.6 (C-1'), 110.9 (C-2'), 148.1 (C-3'), 150.9 (C-4'), 115.8 (C-5'), 120.1 (C-6'); 56.1 (3'-OCH ₃); 7-O-glucose – 99.7 (C-1''), 92.2 (C-2''), 75.1 (C-3''), 68.5 (C-4''), 76.7 (C-5''), 60.2 (C-6''); 2''-O-glucose – 104.0 (C-1'''), 74.0 (C-2'''), 75.5 (C-3'''), 69.6 (C-4'''), 73.4 (C-5'''), 62.9 (C-6'''); 6'''-O-acetyl – 20.2 (COCH ₃), 170.3 (COCH ₃).

Table S10. Retention times (t_R), molecular formulas (MF), mass-spectral data for negative ionization (MSD), and identification level (IL) of compounds Ch-1–Ch-24 found in *Cirsium heterophyllum* herb.

No	t_R , min	Compound	MF	MSD, m/z	IL
Ch-1	6.41	Protocatechuic acid <i>O</i> -hexoside (Cn-3 isomer)	C ₁₃ H ₁₆ O ₉	315, 153	2
Ch-2	6.81	Protocatechuic acid 4- <i>O</i> -glucoside	C ₁₃ H ₁₆ O ₉	315, 153	1b
Ch-3	8.01	4- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ch-4	9.84	4- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ch-5	10.12	4- <i>O</i> - <i>p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1a
Ch-6	10.44	5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ch-7	10.70	3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ch-8	12.16	5- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ch-9	12.53	5- <i>O</i> - <i>p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1b
Ch-10	13.64	Schaftoside (apigenin 6- <i>C</i> -glucoside-8- <i>C</i> -arabinoside)	C ₂₆ H ₂₈ O ₁₄	563	1b
Ch-11	14.73	3- <i>O</i> - <i>p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1b
Ch-12	15.43	Scolymoside (veronicastroside, luteolin 7- <i>O</i> -rutinoside)	C ₂₇ H ₃₀ O ₁₅	593, 447, 285	1b
Ch-13	15.98	Cynaroside (luteolin 7- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₁	447, 285	1a
Ch-14	16.24	Chrysoeriol 7- <i>O</i> -glucoside	C ₂₂ H ₂₂ O ₁₁	461, 299	1b
Ch-15	16.39	Isorhoifolin (apigenin 7- <i>O</i> -rutinoside)	C ₂₇ H ₃₀ O ₁₄	577, 431, 269	1b
Ch-16	16.67	Chrysoeriol 7- <i>O</i> -(2''- <i>O</i> -acetyl)-glucoside	C ₂₄ H ₂₄ O ₁₂	503, 461, 299	2
Ch-17	16.75	3,4-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Ch-18	16.93	Dracocephaloside (luteolin 3'- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₁	447, 285	1a
Ch-19	17.02	Chrysoeriol 7- <i>O</i> -(6''- <i>O</i> -acetyl)-glucoside	C ₂₄ H ₂₄ O ₁₂	503, 461, 299	1a
Ch-20	17.04	3,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Ch-21	17.16	Cosmosiin (apigenin 7- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₀	431, 269	1b
Ch-22	17.43	Chrysoeriol 4'- <i>O</i> -glucoside	C ₂₂ H ₂₂ O ₁₁	461, 299	1a
Ch-23	17.55	4,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Ch-24	17.57	Acacetin 7- <i>O</i> -glucoside	C ₂₂ H ₂₂ O ₁₀	445, 283	1b
Ch-25	17.60	Luteolin 4'- <i>O</i> -glucoside	C ₂₁ H ₂₀ O ₁₁	447, 285	1b
Ch-26	17.85	Pectolinarigenin 7- <i>O</i> -glucoside	C ₂₂ H ₂₄ O ₁₁	475, 313	1b

* Identification levels: (1) identified compounds after comparison of retention times, UV and mass-spectral data with previously isolated compounds (1a) or commercial reference standard (1b) or isolated in present work (1c); (2) putatively annotated compounds after comparison of retention times, UV and mass-spectral data with literature data.

Table S11. Retention times (t_R), molecular formulas (MF), mass-spectral data for negative ionization (MSD), and identification level (IL) of compounds El-1–El-19 found in *Echinops davuricus* herb.

No	t_R , min	Compound	MF	MSD, m/z	IL
El-1	6.81	Protocatechuic acid 4- <i>O</i> -glucoside	C ₁₃ H ₁₆ O ₉	315, 153	1b
El-2	8.01	4- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
El-3	10.44	5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
El-4	10.70	3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
El-5	12.16	5- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
El-6	12.53	5- <i>O-p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1b
El-7	14.73	3- <i>O-p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	337, 163	1b
El-8	15.97	Isoquercitrin (quercetin 3- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₂	463, 301	1b
El-9	16.00	Hyperoside (quercetin 3- <i>O</i> -galactoside)	C ₂₁ H ₂₀ O ₁₂	463, 301	1b
El-10	16.02	Cynaroside (luteolin 7- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₁	447, 285	1b
El-11	16.24	Chrysoeriol 7- <i>O</i> -glucoside	C ₂₂ H ₂₂ O ₁₁	461, 299	1b
El-12	16.64	3,4-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
El-13	16.65	Luteolin 5- <i>O</i> -glucoside	C ₂₁ H ₂₀ O ₁₁	447, 285	1b
El-14	16.93	Dracocephaloside (luteolin 3'- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₁	447, 285	1a
El-15	17.04	3,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
El-16	17.55	4,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
El-17	23.06	Echitin (apigenin 7- <i>O</i> -(2''- <i>O-p</i> -coumaroyl)-glucoside)	C ₃₀ H ₂₆ O ₁₂	577, 431, 269	1a
El-18	23.29	N ¹ ,N ⁵ ,N ¹⁰ -Tri- <i>O</i> -(<i>EEE</i>)- <i>p</i> -coumaroyl-spermidine	C ₃₄ H ₃₇ N ₃ O ₆	582, 436, 290, 163, 144	1b
El-19	24.22	Echinacin (apigenin 7- <i>O</i> -(6''- <i>O-p</i> -coumaroyl)-glucoside)	C ₃₀ H ₂₆ O ₁₂	577, 431, 269	1b

* Identification levels: (1) identified compounds after comparison of retention times, UV and mass-spectral data with previously isolated compounds (1a) or commercial reference standard (1b) or isolated in present work (1c); (2) putatively annotated compounds after comparison of retention times, UV and mass-spectral data with literature data.

Table S12. Retention times (t_R), molecular formulas (MF), mass-spectral data for negative ionization (MSD), and identification level (IL) of compounds Ic-1–Ic-25 found in *Ixeris chinensis* subsp. *versicolor* herb.

No	t_R , min	Compound	MF	MSD, m/z	IL
Ic-1	8.01	4- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ic-2	8.87	Cichoriin (esculetin 7- <i>O</i> -glucoside)	C ₁₅ H ₁₆ O ₉	339, 177	1b
Ic-3	9.15	Caftaric acid isomer	C ₁₃ H ₁₂ O ₉	311, 179	1b
Ic-4	9.48	Caftaric acid	C ₁₃ H ₁₂ O ₉	311, 179	1b
Ic-5	9.52	Baimaside (quercetin 3- <i>O</i> -sophoroside)	C ₂₇ H ₃₀ O ₁₇	625, 463, 301	1b
Ic-6	10.44	5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ic-7	10.70	3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ic-8	10.93	Quercetin 3- <i>O</i> -gentiobioside	C ₂₇ H ₃₀ O ₁₇	625, 463, 301	1b
Ic-9	10.98	Quercetin 3- <i>O</i> -(2''- <i>O</i> -arabinosyl)-glucoside	C ₂₆ H ₂₈ O ₁₆	595, 463, 301	2
Ic-10	11.08	Kaempferol di- <i>O</i> -hexoside	C ₂₇ H ₃₀ O ₁₆	609, 447, 285	2
Ic-11	12.31	Peltatoside (quercetin 3- <i>O</i> -(6''- <i>O</i> -arabinosyl)-glucoside)	C ₂₆ H ₂₈ O ₁₆	595, 463, 301	1b
Ic-12	12.39	5- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ic-13	12.46	Sophoraflavonolside (kaempferol 3- <i>O</i> -sophoroside)	C ₂₇ H ₃₀ O ₁₆	609, 447, 285	1b
Ic-14	13.22	Kaempferol 3- <i>O</i> -gentiobioside	C ₂₇ H ₃₀ O ₁₆	609, 447, 285	1b
Ic-15	14.32	Coutaric acid	C ₁₃ H ₁₂ O ₈	295, 163	1b
Ic-16	14.66	Calendoflavobioside (quercetin 3- <i>O</i> -neohesperidoside)	C ₂₇ H ₃₀ O ₁₆	609, 463, 285	1a
Ic-17	15.57	Rutin (quercetin 3- <i>O</i> -rutinoside)	C ₂₇ H ₃₀ O ₁₆	609, 463, 285	1b
Ic-18	16.07	Populnin (kaempferol 7- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₁	447, 285	1b
Ic-19	16.24	Chrysoeriol 7- <i>O</i> -glucoside	C ₂₂ H ₂₂ O ₁₁	461, 299	1b
Ic-20	17.30	Cichoric acid (di- <i>trans</i> isomer)	C ₂₂ H ₁₈ O ₁₂	473, 311, 179	1b
Ic-21	18.01	Cichoric acid (<i>cis-trans</i> isomer)	C ₂₂ H ₁₈ O ₁₂	473, 311, 179	2
Ic-22	19.29	<i>p</i> -Coumaroyl-caffeoyl-tartaric acid	C ₂₂ H ₁₈ O ₁₁	457, 311, 295, 179, 163	2
Ic-23	19.91	Feruloyl-caffeoyl-tartaric acid	C ₂₃ H ₂₀ O ₁₂	587, 425, 411, 193, 179	2

* Identification levels: (1) identified compounds after comparison of retention times, UV and mass-spectral data with previously isolated compounds (1a) or commercial reference standard (1b) or isolated in present work (1c); (2) putatively annotated compounds after comparison of retention times, UV and mass-spectral data with literature data.

Table S13. Retention times (t_R), molecular formulas (MF), mass-spectral data for negative ionization (MSD), and identification level (IL) of compounds Ls-1–Ls-41 found in *Lactuca sibirica* herb.

No	t_R , min	Compound	MF	MSD, m/z	IL
Ls-1	3.09	Vanilloyl quinic acid (probably 1- <i>O</i> -isomer)	C ₁₅ H ₁₈ O ₉	341, 167	2
Ls-2	5.32	Protocatechuic acid <i>O</i> -hexoside (Ls-4 isomer)	C ₁₃ H ₁₆ O ₉	315, 153	2
Ls-3	6.78	Protocatechuic acid <i>O</i> -hexoside (Ls-4 isomer)	C ₁₃ H ₁₆ O ₉	315, 153	2
Ls-4	7.41	Protocatechuic acid 4- <i>O</i> -glucoside	C ₁₃ H ₁₆ O ₉	315, 153	1b
Ls-5	7.99	Vanilloyl quinic acid (probably 4- <i>O</i> -isomer)	C ₁₅ H ₁₈ O ₉	341, 167	2
Ls-6	8.01	4- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ls-7	8.10	Vanilloyl quinic acid (probably 5- <i>O</i> -isomer)	C ₁₅ H ₁₈ O ₉	341, 167	2
Ls-8	9.48	Caftaric acid	C ₁₃ H ₁₂ O ₉	311, 179	1b
Ls-9	9.91	Vanilloyl quinic acid (probably 5- <i>O</i> -isomer)	C ₁₅ H ₁₈ O ₉	341, 167	2
Ls-10	10.44	5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ls-11	10.70	3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ls-12	12.29	Luteolin tri- <i>O</i> -hexoside	C ₃₃ H ₄₀ O ₂₁	771, 609, 447, 285	2
Ls-13	12.39	5- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	C ₁₆ H ₁₈ O ₉	353, 179	1b
Ls-14	13.15	Luteolin tri- <i>O</i> -hexoside	C ₃₃ H ₄₀ O ₂₁	771, 609, 447, 285	2
Ls-15	14.63	Divanilloyl quinic acid (probably 3,4-isomer)	C ₂₃ H ₂₄ O ₁₂	491, 341, 167	2
Ls-16	15.05	Divanilloyl quinic acid (probably 3,5-isomer)	C ₂₃ H ₂₄ O ₁₂	491, 341, 167	2
Ls-17	15.53	Kaempferol 3- <i>O</i> -neohesperidoside	C ₂₇ H ₃₀ O ₁₅	593, 447, 285	1b
Ls-18	15.58	Rutin (quercetin 3- <i>O</i> -rutinoside)	C ₂₇ H ₃₀ O ₁₆	609, 463, 301	1b
Ls-19	15.61	Divanilloyl quinic acid (probably 4,5-isomer)	C ₂₃ H ₂₄ O ₁₂	491, 341, 167	2
Ls-20	16.08	Cynaroside (luteolin 7- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₁	447, 285	1b
Ls-21	16.31	Nicotiflorin (kaempferol 3- <i>O</i> -rutinoside)	C ₂₇ H ₃₀ O ₁₅	593, 447, 285	1b
Ls-22	16.35	Chrysoeriol 7- <i>O</i> -glucoside	C ₂₂ H ₂₂ O ₁₁	461, 299	1b
Ls-23	16.70	3,4-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Ls-24	16.73	Quercetin 3- <i>O</i> -(6''- <i>O</i> -acetyl)-glucoside	C ₂₃ H ₂₂ O ₁₃	505, 463, 301	1a
Ls-25	16.97	Dracocephaloside (luteolin 3'- <i>O</i> -glucoside)	C ₂₁ H ₂₀ O ₁₁	447, 285	1a
Ls-26	17.11	3,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Ls-27	17.30	Cichoric acid (<i>di-trans</i> isomer)	C ₂₂ H ₁₈ O ₁₂	473, 311, 179	1b
Ls-28	17.32	Quercetin 3- <i>O</i> -(3''- <i>O</i> -acetyl)-glucoside	C ₂₃ H ₂₂ O ₁₃	505, 463, 301	1a
Ls-29	17.60	4,5-Di- <i>O</i> -caffeoyl quinic acid	C ₂₅ H ₂₄ O ₁₂	515, 353, 179	1b
Ls-30	17.64	Apigenin 7- <i>O</i> -glucuronide	C ₂₁ H ₁₈ O ₁₁	445, 269	1b
Ls-31	17.69	Luteolin 4'- <i>O</i> -glucoside	C ₂₁ H ₂₀ O ₁₁	447, 285	1b
Ls-32	17.85	Quercetin 3- <i>O</i> -(4''- <i>O</i> -acetyl)-glucoside	C ₂₃ H ₂₂ O ₁₃	505, 463, 301	1a
Ls-33	18.01	Cichoric acid (<i>cis-trans</i> isomer)	C ₂₂ H ₁₈ O ₁₂	473, 311, 179	2
Ls-34	19.02	Cichoric acid (<i>cis-cis</i> isomer)	C ₂₂ H ₁₈ O ₁₂	473, 311, 179	2
Ls-35	19.29	<i>p</i> -Coumaroyl-caffeoyl-tartaric acid	C ₂₂ H ₁₈ O ₁₁	457, 311, 295, 179, 163	2
Ls-36	19.91	Feruloyl-caffeoyl-tartaric acid	C ₂₃ H ₂₀ O ₁₂	587, 425, 411, 193, 179	2
Ls-37	20.08	<i>p</i> -Coumaroyl-caffeoyl-tartaric acid	C ₂₂ H ₁₈ O ₁₁	457, 311, 295, 179, 163	2
Ls-38	22.52	Luteolin	C ₁₅ H ₁₀ O ₆	285	1b
Ls-39	23.81	Tetra- <i>O-p</i> -coumaroyl spermine	C ₄₆ H ₅₀ N ₄ O ₈	785, 639, 493, 347, 163	2
Ls-40	23.87	Tetra- <i>O-p</i> -coumaroyl spermine	C ₄₆ H ₅₀ N ₄ O ₈	785, 639, 493, 347, 163	2
Ls-41	24.03	Tetra- <i>O-p</i> -coumaroyl spermine	C ₄₆ H ₅₀ N ₄ O ₈	785, 639, 493, 347, 163	2
Ls-42	24.17	Tetra- <i>O-p</i> -coumaroyl spermine	C ₄₆ H ₅₀ N ₄ O ₈	785, 639, 493, 347, 163	2
Ls-43	24.63	Apigenin	C ₁₅ H ₁₀ O ₅	269	1b

* Identification levels: (1) identified compounds after comparison of retention times, UV and mass-spectral data with previously isolated compounds (1a) or commercial reference standard (1b) or isolated in present work (1c); (2) putatively annotated compounds after comparison of retention times, UV and mass-spectral data with literature data.

Table S14. Synopsis of compounds found in six Asteraceae species.

Compound	Aj	Cn	Ch	Ed	Ic	Ls
1. Benzoates						
<u>1.1. Hydroxybenzoic acid glucosides</u>						
Protocatechuic acid <i>O</i> -hexoside		✓				✓
Protocatechuic acid <i>O</i> -hexoside		✓	✓			✓
Protocatechuic acid 4- <i>O</i> -glucoside		✓	✓	✓		✓
<u>1.2. Hydroxybenzoyl quinic acids</u>						
Vanilloyl quinic acid (probably 1- <i>O</i> -isomer)						✓
Vanilloyl quinic acid (probably 4- <i>O</i> -isomer)						✓
Vanilloyl quinic acid (probably 5- <i>O</i> -isomer)						✓
Vanilloyl quinic acid (probably 5- <i>O</i> -isomer)						✓
Divanilloyl quinic acid (probably 3,4- <i>O</i> -isomer)						✓
Divanilloyl quinic acid (probably 3,5- <i>O</i> -isomer)						✓
Divanilloyl quinic acid (probably 4,5- <i>O</i> -isomer)						✓
2. Coumarins						
Esculetin 7- <i>O</i> -glucoside					✓	
3. Hydroxycinnamates						
<u>3.1. Caffeoyl glucaric acids</u>						
3- <i>O</i> -Caffeoyl glucaric acid	✓					
4- <i>O</i> -Caffeoyl glucaric acid	✓	✓				
2- <i>O</i> -Caffeoyl glucaric acid	✓					
5- <i>O</i> -Caffeoyl glucaric acid	✓					
<u>3.2. Caffeoyl tartaric acids</u>						
Caftaric acid isomer					✓	
Caftaric acid					✓	✓
Cichoric acid (<i>di-trans</i> isomer)					✓	✓
Cichoric acid (<i>cis-trans</i> isomer)					✓	✓
Cichoric acid (<i>cis-cis</i> isomer)						✓
<u>3.3. <i>p</i>-Coumaroyl tartaric acids</u>						
Coutaric acid					✓	
<u>3.4. Mixed tartaric acids</u>						
<i>p</i> -Coumaroyl-caffeoyl-tartaric acid					✓	✓
Feruloyl-caffeoyl-tartaric acid					✓	✓
<i>p</i> -Coumaroyl-caffeoyl-tartaric acid						✓
<u>3.4. <i>p</i>-Coumaroyl quinic acids</u>						
4- <i>O-p</i> -Coumaroyl quinic acid		✓	✓			
5- <i>O-p</i> -Coumaroyl quinic acid		✓	✓	✓		
3- <i>O-p</i> -Coumaroyl quinic acid		✓	✓	✓		
<u>3.5. Caffeoyl quinic acids</u>						
4- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	✓		✓	✓	✓	✓
4- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	✓		✓	✓		
5- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	✓	✓	✓	✓	✓	✓
3- <i>O</i> -Caffeoyl quinic acid (<i>trans</i> -)	✓		✓	✓	✓	✓
5- <i>O</i> -Caffeoyl quinic acid (<i>cis</i> -)	✓			✓	✓	✓
3,4-Di- <i>O</i> -caffeoyl quinic acid	✓		✓	✓		✓
3,5-Di- <i>O</i> -caffeoyl quinic acid	✓		✓	✓		✓
4,5-Di- <i>O</i> -caffeoyl quinic acid	✓		✓	✓		✓
<u>3.6. Feruloyl quinic acids</u>						
4- <i>O</i> -Feruloyl quinic acid		✓			✓	
5- <i>O</i> -Feruloyl quinic acid		✓				
3- <i>O</i> -Feruloyl quinic acid		✓				

4. Flavonoids						
4.1. Apigenin O-glucosides						
Apigenin 7-O-glucoside			✓			
Apigenin 7-O-glucuronide						✓
Apigenin 7-O-rutinoside			✓			
Apigenin 7-O-(2''-O-p-coumaroyl)-glucoside				✓		
Apigenin 7-O-(6''-O-p-coumaroyl)-glucoside				✓		
Apigenin di-O-hexoside-O-acetate		✓				
Apigenin di-O-hexoside-O-acetate		✓				
4.2. Apigenin C-glucosides						
Apigenin 6-C-glucoside-8-C-arabinoside	✓		✓			
4.3. Acacetin O-glucosides						
Acacetin 7-O-glucoside			✓			
4.4. Luteolin O-glucosides						
Luteolin 5-O-glucoside				✓		
Luteolin 7-O-glucoside			✓	✓		✓
Luteolin 3'-O-glucoside			✓	✓		✓
Luteolin 4'-O-glucoside			✓			✓
Luteolin 7-O-rutinoside			✓			
Luteolin 7-O-(2''-O-glucosyl)-glucoside		✓				
Luteolin 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside		✓				
Luteolin O-hexoside-O-pentoside		✓				
Luteolin di-O-hexoside-O-acetate		✓				
Luteolin tri-O-hexoside						✓
Luteolin tri-O-hexoside						✓
4.5. Chrysoeriol O-glucosides						
Chrysoeriol 7-O-glucoside			✓	✓	✓	✓
Chrysoeriol 4'-O-glucoside			✓			
Chrysoeriol 7-O-(2''-O-acetyl)-glucoside			✓			
Chrysoeriol 7-O-(6''-O-acetyl)-glucoside			✓			
Chrysoeriol 7-O-(2''-O-(6'''-O-acetyl)-glucosyl)-glucoside		✓				
4.6. 6-Hydroxyluteolin O-glucosides						
6-Hydroxyluteolin di-O-hexoside	✓					
4.7. Pectolinarigenin O-glucosides						
Pectolinarigenin 7-O-glucoside			✓			
4.8. Kaempferol O-glucosides						
Kaempferol 7-O-glucoside					✓	
Kaempferol 3-O-neohesperidoside						✓
Kaempferol 3-O-rutinoside						✓
Kaempferol 3-O-sophoroside					✓	
Kaempferol 3-O-gentiobioside					✓	
Kaempferol di-O-hexoside					✓	
4.9. Quercetin O-glucosides						
Quercetin 3-O-glucoside				✓		
Quercetin 3-O-galactoside				✓		
Quercetin 3-O-(2''-O-arabinosyl)-glucoside					✓	
Quercetin 3-O-(6''-O-arabinosyl)-glucoside					✓	
Quercetin 3-O-neohesperidoside	✓				✓	
Quercetin 3-O-rungioside	✓					
Quercetin 3-O-(4''-O-rhamnosyl)-glucoside	✓					
Quercetin 3-O-rutinoside	✓				✓	✓
Quercetin 3-O-sophoroside					✓	
Quercetin 3-O-gentiobioside	✓				✓	

Quercetin 3-O-(2''-O-acetyl)-glucoside	√					
Quercetin 3-O-(3''-O-acetyl)-glucoside						√
Quercetin 3-O-(4''-O-acetyl)-glucoside	√					√
Quercetin 3-O-(6''-O-acetyl)-glucoside	√					√
<u>4.10. Isorhamnetin O-glucosides</u>						
Isorhamnetin 3-O-(2''-O-acetyl)-glucoside	√					
Isorhamnetin 3-O-(6''-O-acetyl)-glucoside	√					
<u>4.11. Flavone aglycones</u>						
Apigenin						√
Luteolin						√
Cirsiliol	√					
Cirsilineol	√					
<u>4.12. Flavonol aglycones</u>						
Axillarin	√					
Chrysosplenetin	√					
5. Phenylamines						
<u>5.1. Tri-O-p-coumaroyl spermines</u>						
Tri-O-p-coumaroyl spermine	√					
Tri-O-p-coumaroyl spermine	√					
Tri-O-p-coumaroyl spermine	√					
<u>5.2. Tetra-O-p-coumaroyl spermines</u>						
Tetra-O-p-coumaroyl spermines						√
Tetra-O-p-coumaroyl spermine						√
Tetra-O-p-coumaroyl spermine						√
Tetra-O-p-coumaroyl spermine	√					√
<u>5.3. Tri-O-p-coumaroyl-spermidine</u>						
N ¹ ,N ⁵ ,N ¹⁰ -Tri-O-(EEE)-p-coumaroyl-spermidine				√		