

Review

# Functionality and Health-Promoting Properties of Polysaccharide and Plant-Derived Substances from *Mesona chinensis*

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**Abstract:** *Mesona chinensis*, in Thai called Chao Kuay and in Chinese Hsian-tsao, belongs to the Lamiaceae family. This herbal plant grows widely in Southern China, Taiwan (China), Malaysia, the Philippines, Indonesia, Vietnam, and Thailand. The *Mesona* plant is used to make functional products such as drinks and soft textured sweet treats, and also traditional medicine, to treat heat stroke, high blood pressure, heart attack, high blood sugar, hepatic diseases, colon diseases, inflammatory conditions, and to alleviate myalgia. The proximate composition of *M. chinensis* is a mixture of protein, fat, fiber, ash, and minerals. The main biological compounds in *M. chinensis* extracts are polysaccharides, terpenoids, flavonoids, and polyphenols, with wide-ranging pharmacological properties including antioxidant, antidiabetic, antilipidemic, carcinoma-inhibitory, renal-protective, antihypertensive, DNA damage-protective, and anti-inflammatory effects. This review investigated the proximate composition, polysaccharide type, and pharmacological properties of *M. chinensis* extracts. Phytochemical properties enhance the actions of the gut microbiota and improve health benefits. This review assessed the functional and medicinal activities of *M. chinensis* extracts. Future studies should further elucidate the in vitro/in vivo mechanisms of this plant extract and its impact on gut health.

**Keywords:** *Mesona chinensis*; medicinal plant; pharmacological activity; bioactive compound; polysaccharide



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## 1. Introduction

The herbaceous plant *Mesona chinensis* is part of the Lamiaceae family [1]. It grows well in southern China (in Zhejiang, Jiangxi, and Guangdong provinces, as well as Taiwan) [2], India, Malaysia, the Philippines, Indonesia, Vietnam, and Thailand [3,4]. The *Mesona* plant has several names, including *Mesona chinensis* Benth., *Mesona procumbens* Hemsl., *Mesona parviflora* (Benth.) Briq., *Mesona philippinensis* Merr., *Mesona palustris* Blume., *Mesona wallichiana* Benth., *Mesona elegans* Hayata., *Geniosporum parviflorum* Benth., *Platostoma chinense* (Benth.) A. J. Paton., and *Platostoma palustre* (Blume) A. J. Paton [5–7]. *Mesona chinensis*, *Mesona procumbens*, and *Mesona palustris* have been documented in China, Taiwan (China), and Indonesia, respectively. The plant has local names in different countries. For example, in China and Taiwan (China), it is Hsian-tsao or Liangfen Cao; in Indonesia and Malaysia, it is black cincau; in Vietnam, it is Suong Sao; and in Thailand it is Chao Kuay [8–11]. This perennial herb grows 15–100 cm high and its stem is covered with soft hair. The leaves are narrowly ovate and almost circular [12], as demonstrated in Figure 1. The plant grows in ditches, forest slopes, streams, and also on dry sandy land, demonstrating good environmental adaptability [13]. However, drought in summer and cold winters lead to poor plant growth or even death, and impact the yield [14], with a reduced gross weight, number of roots, and aerial parts [15]. This herb is normally digested as a functional beverage and a semi-solid sweet dessert and has been used as a traditional

medicine in China, Vietnam, and Indonesia for thousands of years [2,16,17]. The plant is used to treat heat shock, fever, hypertension, heart attack, diabetes, muscle pains, liver diseases, colon diseases, and inflammatory conditions, and also to alleviate muscle or joint pain [16,18–22], because of the various active phytochemicals it contains, such as flavonoids and phenolic acid [23]. Mesona is also utilized to make a popular jelly dessert, due to its high polysaccharide component [24].

Many researchers have demonstrated the biological activities of phytochemical compounds. However, the nutraceutical and pharmacological properties reported for *M. chinensis* as evidence for its health benefits as a medicine are still limited. This review discussed the recent scientific information on the Mesona plant regarding the composition of its phytochemicals and its alteration of gut microbiota, as well as its health benefits. Several bioactive compounds and polysaccharide components have been reported to have various pharmacological effects including antioxidant activities [25], hypoglycemic and hypolipidemic activities [26,27], antiproliferative activities [28], a growth inhibitory effect on hepatocellular carcinoma (HepG2) cells [15], anti-dyslipidemia activities [29], renal-protective activities [30], antihypertensive [31] and DNA damage-protective activities [32], anti-inflammatory activities [33], antimutagenic effects [34], and antibiosis activities [35].



Figure 1. *Mesona chinensis* leaves.

## 2. Proximate and Mineral Composition

The proximate and mineral components of *M. chinensis* leaves are presented in Table 1. Differences in the environment [36] and climatic conditions impact major plant constituents and chemical components [37]. The mineral contents of *M. chinensis* when extracted with sodium bicarbonate in heated water were  $1420 \pm 10$  µg/g Mg,  $5.8 \pm 1$  µg/g Cu,  $26 \pm 1$  µg/g Zn,  $66 \pm 1$  µg/g Fe,  $290 \pm 5$  µg/g Mn,  $2810 \pm 10$  µg/g Ca,  $10,600 \pm 20$  µg/g K, and  $40,300 \pm 10$  µg/g Na [36]. The major minerals in *M. chinensis* gum (MBG) included Na, K, Ca, and Mg, with Na making up 73% of the total. Similarly, Yuris et al. stated that polysaccharide extracted from plant powder consisted of K, Mg, P, Ca, Na, Fe, Cu, I, Mn, Zn, and Se [38].

Table 1. Proximate composition of *Mesona chinensis* leaves from different regions of South China and Taiwan (China).

Source	Extraction	Proximate Composition (%)				Refs.
		Crude Protein	Crude Fat	Crude Fiber	Ash	
<i>Mesona chinensis</i> leaf powder (China)	With sodium bicarbonate in heated water at 95 °C for 2 h	9.74	-	2.98	30.9	[36]
<i>Mesona chinensis</i> powder (China)	Water extract	9.0	0.1	-	28.2	[38]
<i>Mesona chinensis</i> leaves (farm market in Taiwan)	With sodium bicarbonate in heated water at 95 °C for 4 h	4.56	-	1.07	26.97	[39]

Table 1. Cont.

Source	Extraction	Proximate Composition (%)				Refs.
		Crude Protein	Crude Fat	Crude Fiber	Ash	
<i>Mesona chinensis</i> leaves (contracted farmer, Taiwan)	With sodium bicarbonate in heated water at 95 °C for 4 h	10.04	0.52	1.47	26.2	[40]
<i>Mesona chinensis</i> leaves (contracted farmer in Miao-Li, Taiwan)	With sodium bicarbonate in heated water at 95 °C for 4 h	4.60	0.90	1.10	27.0	[41]

### 3. Polysaccharides and Phytochemicals of *M. chinensis* Extracts

Polymeric macromolecules of carbohydrates or polysaccharides contain long chains of monosaccharide units joined by glycosidic linkages [42,43]. Polysaccharides are essential for all living cells, particularly in plants and microorganisms such as bacteria, yeast, and mold [44,45]. Polysaccharides can be divided into two forms, including homopolysaccharides, which comprise only one type of simple carbohydrate, and heteroglycans, which contain a mixture of two or more different monosaccharides [46].

#### 3.1. Physicochemical Properties

Polysaccharides comprise the main functional components of *M. chinensis* [47], which include structural heteropolysaccharide consisting of an  $\alpha$ -1, 4-linked galacturonan backbone with some  $\alpha$ -1, 2-linked rhap (Rhamnose) residues (Figure 2), and large amounts of uronic acid [20,48], similar to the pectin structure. Wang et al. reported that polysaccharides of the *Mesona* plant are acidic glycoprotein compounds containing uronic acid and some protein [49]. *Mesona* plants can be classified as acidic heteroglycan material containing galacturonic acids and a few glucuronic acids in the polysaccharide backbone [50]. The monosaccharide composition comprised glucose, galactose, galacturonic acid, rhamnose, arabinose, and xylose, while mannose was also reported, as shown in Table 2. Chen et al. listed the different monosaccharide compositions of seven freshly dried and six dried *Mesona* plants after 1 year of storage [51]. Their results indicated that storage time was an important factor affecting monosaccharide quality, while monosaccharide compositions were dependent on the material source, extraction method, growth stage of the plant, and the cultivation environment [25,52,53]. Previous FT-IR spectral band and peak studies determined that the structure of *Mesona* polysaccharides (MPs) consisted of the following three functional groups: carbonyl (C=O), carboxyl (COOH), and hydroxyl (OH) [49,50,54,55]. *M. chinensis* has been widely studied for its chemical composition and various biological activities in China and Taiwan (China), but few studies are documented from the Indonesian region, possibly due to the lack of scientific records, research funding support, and marketing value. Black jelly, as a product, has been mainly produced from *Mesona* plants cultivated in China; however, recently, some large companies producing black jelly (or Chao Kuay in Thailand) have used raw materials from several regions, including Vietnam and Indonesia, due to the better pricing and more consistent quality between each lot (according to survey and interview data from company owners in Thailand and Malaysia).

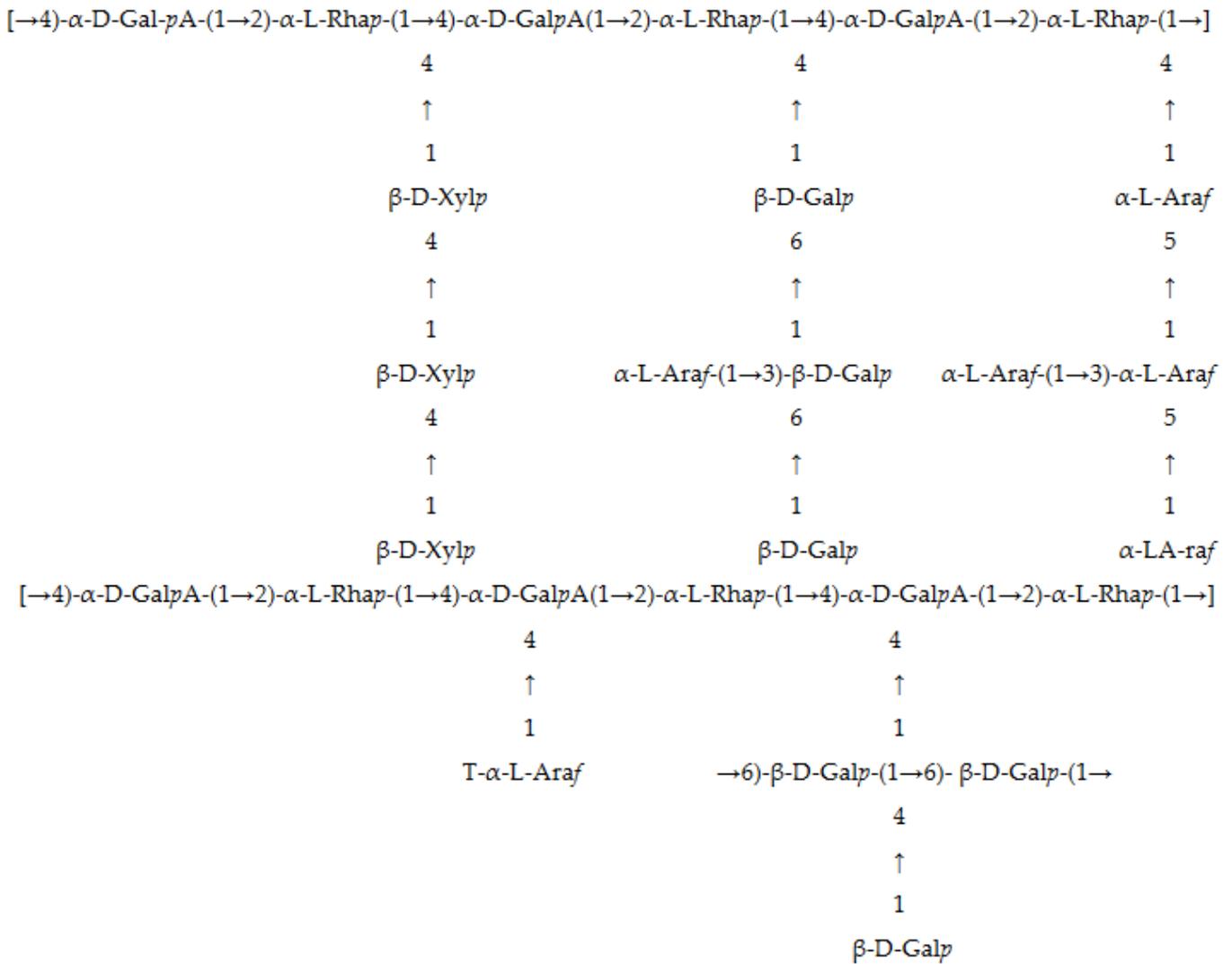


Figure 2. A possible acidic polysaccharide structural model of *M. chinensis*.

**Table 2.** Physicochemical properties of *Mesona chinensis* polysaccharides.

Collected Region	Extraction	Yield (%)	Molecular Weight (kDa)	Chemical Composition (%)				Monosaccharide Composition (Mole Ratios)						Refs.
				Total Sugar	Uronic Acid	Protein	Glu	Gal	Gala	Rha	Ara	Man	Xyl	
Xiaoshicheng, Ganzhou, Jiangxi, China	Boil in hot water 95 °C for 2 h with Na <sub>2</sub> CO <sub>3</sub>	-	158	29.03	17.06	22.64	N.D.	2.80	2.40	N.D.	N.D.	N.D.	5.50	[4]
China	Boil in hot water 95 °C for 2 h with Na <sub>2</sub> HCO <sub>3</sub>	29.36	16.26	42.20	13.80	9.74	2.30	3.10	1.40	1.20	2.30	0.20	1.00	[36]
Xiaoshicheng, Jiangxi, China	Boil in hot water 95 °C for 3 h with Na <sub>2</sub> CO <sub>3</sub>	-	141.6	16.88	36.91	-	1.36	3.76	17.5	0.87	0.14	-	2.00	[48]
Ganzhou, Jiangxi, China.	Boil in hot water 95 °C for 2.5 h	-	375	81.12	-	14.07	4.90	2.16	6.75	1.38	1.64	-	0.42	[49]
Xiaoshicheng, Jiangxi, China	Boil in hot water 90 °C for 2 h with Na <sub>2</sub> CO <sub>3</sub>	7.05	1450	-	29.30	10.40	1.38	1.0	-	-	-	-	-	[50]
Yichun, Jiangxi, China	Boil in hot water 100 °C	0.84	44.39	30.69	20.86	25.30	1.12	1.97	1.69	0.42	0.30	0.50	N.D.	[51]
Xiaoshicheng, Ganzhou, Jiangxi, China	Boil in hot water 95 °C for 2.5 h with Na <sub>2</sub> CO <sub>3</sub>	11.14	195	34.40	24.30	17.30	1.00	1.34	0.25	-	N.D.	-	N.D.	[52]
Xiaoshicheng, Ganzhou, Jiangxi, China	Boil in hot water 95 °C for 2.5 h with Na <sub>2</sub> CO <sub>3</sub>	-	204	32.28	29.52	31.35	6.24	0.82	-	0.11	0.32	-	0.34	[54]
Ganzhou, Jiangxi, China.	Boil in hot water 100 °C for 2 h	1.68	157	39.01	29.30	27.52	1.49	0.68	6.33	-	-	-	2.54	[55]

N.D.: Not detectable or lower than the limit of determination. Glu: glucose, Gal: galactose, Gala: galacturonic acid, Rha: rhamnose, Ara: arabinose, Man: mannose, and Xyl: xylose.

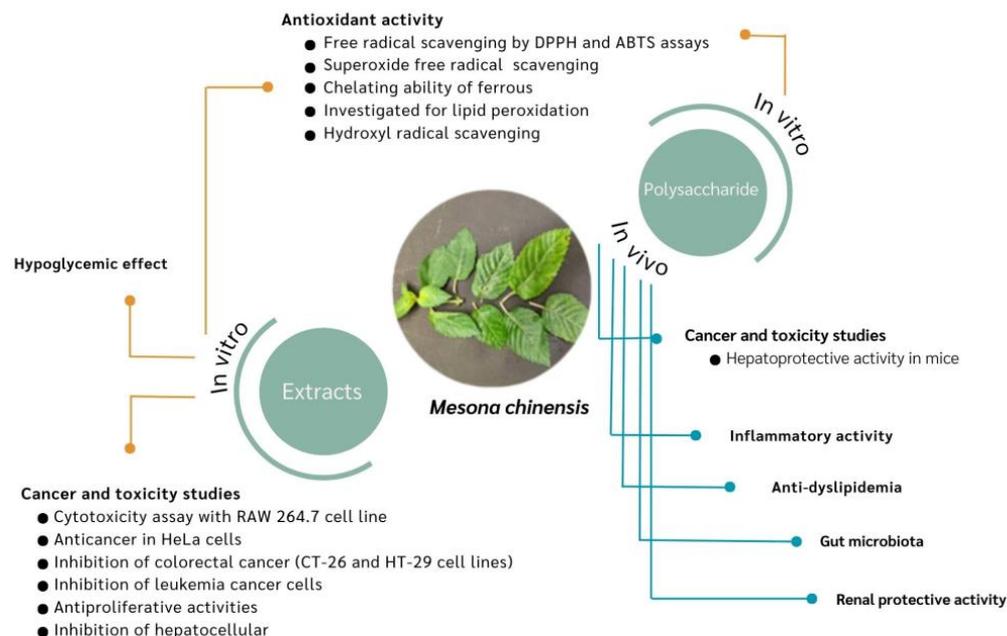
### 3.2. Phytochemicals

The powder extraction of *M. chinensis*, using methanol to separate it into acidic ethyl acetate fractions, was evaluated for polyphenolics. The results showed that the polyphenolic compounds found included protocatechuic acid, *p*-hydroxybenzoic acid, vanillic acid, caffeic acid, and syringic acid [18]. The ethanolic *M. chinensis* extraction yielded rosmarinic acid, apigenin, 7-hydroxycoumarin, ferulic acid, and rutin [16]. Eleven novel diterpenoids were reported as resulting from the methanolic extraction of *M. chinensis* after isolation with *n*-hexane and CH<sub>2</sub>Cl<sub>2</sub>, including seven *ent*-kauranes, three *ent*-atisanes, and one sarcopetalane [28]. Bioactive compounds of *M. chinensis* ethanolic extracts and their fractions, named as F0, F10, F20, F30, F40, F50, and MTFs (*Mesona* total flavonoids) (including aqueous extract (AE)) and detected using high performance liquid chromatography with mass spectrometry (HPLC-MS) analysis, mainly contained caffeic acid, quercetin 3-O-galactoside, isoquercetin, astragaln, rosmarinic acid, aromadendrin-3-O-rutinoside, rosmarinic acid-3-O-glucoside, and kaempferol-7-O-glucoside. MTFs, prepared using an ethanolic extract and X-5 macroporous resin as purification for flavonoid enrichment, exhibited the highest peaks of these compounds [27]. *M. chinensis* extracted in boiled water provided crude polysaccharides (301.7 mg/g) and β-1,3-glucan (68.9 mg/g). The functional groups of the crude polysaccharide extract were confirmed, using FT-IR spectrophotometry, to be hydroxyl and carbonyl [11]. Another study of *Mesona* plants, gathered in Southern China and extracted using deionized water at 80 °C for 2 h before ultra-high performance liquid chromatography with quadrupole time-of-flight-mass spectrometry (UPLC-Q-TOF-MS/MS) analysis, found 5757 compounds including 45 polyphenols, 6 terpenoids, and 6 other unknown compounds [56]. Water extracts of *M. chinensis* recorded seven phenolic compounds, namely kaempferol, apigenin, caffeic acid, protocatechuic acid, syringic acid, vanillic acid, and *p*-hydroxybenzoic acid. Interestingly, caffeic acid and kaempferol gave the highest values of phenolic constituents in the extracts [57]. The results indicated that extraction media with different polarities identified bioactive compounds by following the “like dissolves like” rule. However, raw material types, planting area and harvesting time, preparation method, extraction condition, and different determination assays can also give diverse results.

## 4. Pharmacological Properties

### 4.1. Pharmacological Properties

Consumers now prefer to eat natural plant food to promote their nutrition, as this contains pharmaceuticals, fibers, pigments, minerals, vitamins, and unsaturated fatty acids and is free of manufactured food additives [58]. *M. chinensis* polysaccharides (MCPs) are attracting interest, due to their potential biological activities in food and their pharmacological properties [59], as shown in Figure 3. *Mesona* has been applied into extruded rice products [60] and effervescent powder [29], encapsulated in alginate beads [61], and used for its antitumor [62], anticoagulant [63], antioxidant [64], antidiabetic [65,66], and immunomodulatory activities [67–69]. Most extracted polysaccharides from medicinal plants are nonpoisonous and have no adverse issues [70,71]. Natural products are preferred to synthetic agents, which typically exhibit negative side effects.



**Figure 3.** Summary of proposed pharmacological properties of Mesona polysaccharides (MPs).

#### 4.2. Antioxidant Activity

Free radicals, high-energy particles that ricochet widely and damage cells, can be created in living cells as highly unstable molecules, such as reactive oxygen species (ROS), comprising superoxides, hydroxyls, peroxy, and alkoxy [72]. ROS induce various chronic and degenerative diseases, including dementia and shaking palsy [73], respiratory, neurodegenerative, and digestive diseases [74], cancer, diabetes mellitus, insulin resistance, cardiovascular diseases, atherosclerosis, and aging [75]. MCPs exhibited high potential antioxidant activities. Lai et al. reported that polysaccharide gum from Mesona leaf was strongly concentration-dependent on free radical scavenging activities [40]. The 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging  $IC_{50}$  value was 68.6% at 1250  $\mu\text{g}/\text{mL}$ . Superoxide free radical scavenging activities increased with the extract concentration (86.5% at a dose level of 1250  $\mu\text{g}/\text{mL}$ ), while the chelating ability of ferrous ion also increased to 74.4% at 1.5 mg/mL, and the reducing power increased in a dose-dependent manner.  $\text{FeSO}_4\text{-H}_2\text{O}_2$  was used to induce malondialdehyde (MDA) in rat histology and was evaluated for lipid peroxidation. MDA formation in rat tissue homogenate (brain, liver, and heart tissue) significantly decreased by 15.86–83.68% when adding 5–40 mg/mL of polysaccharide concentrate. Interestingly, water-soluble polysaccharides showed higher potential in hepatic and heart organs than the brain [40], while MCPs provided strong hydroxyl radicals in a concentration-dependent manner, with the highest scavenging rate being  $54.36 \pm 1.56\%$  at 1600  $\mu\text{g}/\text{mL}$ . By contrast, the superoxide anion scavenging activity reached  $58.42 \pm 1.17\%$ , when the value of MCPs was 1600  $\mu\text{g}/\text{mL}$ . The scavenging activities of MCPs on DPPH free radicals gradually increased ( $55.59 \pm 0.69\%$ ) as MCP concentration increased to 1600  $\mu\text{g}/\text{mL}$  [50].

Mesona polysaccharides (MPs) at 1000  $\mu\text{g}/\text{mL}$  showed high DPPH free radical scavenging activity of  $75.11 \pm 0.31\%$ . Similarly, MPs also showed scavenging effects ( $63.26 \pm 0.28\%$ ) against hydroxyl radicals and demonstrated significant dose-dependent defense against  $\text{H}_2\text{O}_2$ -promoted injury to RAW 264.7 macrophage cell line at 100  $\mu\text{g}/\text{mL}$ , measured as  $78.58 \pm 0.11\%$ . Oxidative damage caused by lipid peroxidation to RAW 264.7 cells, measured using the MDA assay gradually decreased to  $96.88 \pm 2.52 \mu\text{mol}/\text{mL}$  at a 100  $\mu\text{g}/\text{mL}$  MP concentration [55]. Chen et al. studied the ability of MPs to scavenge free radicals using the DPPH and ABTS (2,2'-azino-bis (3-ethylbenzthiazoline-6-sulphonic acid)) assays. Their results demonstrated DPPH antioxidant activities of  $75.59 \pm 0.13\%$  at 1000  $\mu\text{g}/\text{mL}$ , while the ABTS free radical scavenging  $IC_{50}$  value was 332.34  $\mu\text{g}/\text{mL}$  [51].

The antioxidant activities of aqueous extracts of *M. chinensis* from various areas in Southern China, including Guangdong Meizhou, Guangdong Raoping, Guangdong Shaoguan, Fujian Longyan, Fujian Zhangzhou, Jiangxi Jian, Jiangxi Ganzhou, Guangxi Chongzuo, and Guangxi Yulin, were evaluated. The results showed that samples obtained from Guangdong Raoping provided the strongest antioxidant ability (as measured using DPPH and ABTS assays) with  $IC_{50}$  values of  $0.00076 \pm 0.00006$  mg/mL and  $0.00383 \pm 0.00017$  mg/mL, respectively [56], indicating that planting area or physiological race significantly impacted biological activity, possibly due to the quality and quantity of the phytochemical compounds. Recently, MCP aqueous and ethanolic extracts and their fractions (F0, F10, F20, F30, F40, and F50) and MTFs were intensively evaluated using DPPH free radical scavenging and FRAP (Ferric reducing antioxidant power) assays. MTFs and F30 exhibited higher free radical scavenging abilities and reducing power than the control (vitamin C), with  $IC_{50}$  values of 0.005323 mg/mL and 0.005278 mg/mL, respectively [27].

Polyphenols and antioxidant activities were further evaluated through gastrointestinal digestion experiments. An aqueous dried *M. chinensis* extract was encapsulated in alginate beads to study the polyphenol and antioxidant activities under a gastrointestinal digestion experiment. The release of total phenolic content (TPC) from the beads was low, at only 8.9%, after soaking in water for 4 h. TPC and FRAP activities of the encapsulated beads were higher than the control, because the gastric pH was lower than the pKa value of the bead material as an alginate monomer; therefore, the protective effect was strong [61]. The results indicated that the Mesona extract improved health through antioxidation activity, particularly when encapsulation was applied before digestion.

#### 4.3. Cancer and Toxicity Studies

Cancer is the most common cause of death in Thailand. Natural products have attracted an increasing interest as novel anticancer drugs, with potential biological activities associated with therapies that have fewer lower side effects [76,77]. The cytotoxic effects of different extractions (water, ethanol, and ethyl acetate) from *M. chinensis* were tested for anticancer activity against Hela cells in an in vitro culture. The results indicated that the water extract induced higher anticancer activity than the other two organic extracts (ethanol and ethyl acetate), with cytotoxicity against Hela cells giving  $IC_{50}$  values of 0.1326, 0.146, and 0.18296 mg/mL, respectively [78]. In an in vivo study, MCPs acted in a dose-dependent manner (0.10, 0.20, and 0.30 g/kg body weight) and provided hepatoprotective activity against acute liver damage induced by  $CCl_4$ . A significant reduction in serum markers was found in the livers of mice after treatment with MCPs at medium and high doses (0.20 and 0.30 mg/kg body weight), when assayed through aspartate aminotransferase (AST) and alanine aminotransferase (ALT) parameters. A reduced impact of  $CCl_4$  toxicity on the serum markers (aspartate aminotransferase, AST, and alanine aminotransferase, ALT) of liver damage in mice was noticed at medium and high doses of MCPs. MCPs also increased levels of antioxidant enzymes (superoxide dismutase, SOD) and non-enzyme antioxidants (glutathione, GSH), while lipid peroxidation levels of liver tissues, evaluated by MDA, significantly declined. Serum levels of IL-1 $\beta$  and TNF- $\alpha$  increased, indicating that MCPs had hepatoprotective activity against acute injured liver induced by  $CCl_4$  [49].

Ethanolic extracts of *M. chinensis*, at concentrations of 1560–100,000  $\mu$ g/mL and 390–3130  $\mu$ g/mL, showed significant inhibition effects against the viability of CT-26 (colorectal cancer cell line) and HT-29 (Human colon cancer cells) colorectal cancer cell lines, respectively [79]. The protective effect of a methanolic extract of *M. chinensis* on human leukemia cancer cells was reported by Huang et al. [28]. The methanolic extract revealed 11 new diterpenoid compounds, mesonols 1–11, and mesonols 1–4 compounds, which provided antiproliferative activities against five cancer cell lines (human lung carcinoma (A549), human liver carcinoma (Hep-3B), human prostate carcinoma (PC-3), human colon carcinoma (HT29), and human monoblastic leukemia (U937)), with reduced toxicity against the RAW 264.7 cell line. Mesonols 1–2 showed higher antiproliferative activities against U937 cells than the standard drug Camptothecin (CPT-11; irinotecan), with  $IC_{50}$  values of

2.66, 1.97, and 4.95  $\mu\text{M}$ , respectively. The results suggested that Mesona plants could be used as alternative active ingredients and food for medicinal and nutraceutical purposes. However, further clinical trials are needed.

#### 4.4. Hypolipidemic Effect

Hyperlipidemia or dyslipidemia is the primary cause of heart attack, resulting from increased serum levels of TC (total cholesterol), LDL (low-density lipoprotein), or TG (triglycerides), or from decreased amounts of HDL (high-density lipoprotein) [80,81]. Pharmaceutical herbs are commonly utilized in various congestive heart failure treatments [82]. Handayani et al. stated that a semisolid product from *M. chinensis* exhibited antihyperlipidemic activity in rats fed a high-density lipoprotein diet. Lipid profiles, including plasma cholesterol and triacylglycerol, were assessed using the enzymatic cholesterol oxidase-*p*-aminophenazone (CHOD-PAP) assay in Wistar rats induced with a hypercholesterol diet. *M. chinensis*, as an effervescent powder, was introduced to the treated rats for 3 weeks. The results showed that increasing doses of *M. chinensis* as an effervescent powder significantly influenced plasma lipid reduction [29]. The antihyperlipidemic activity of a Mesona ethanolic extract was tested on overweight mice fed with greasy food. The mice were separated into two groups. The first group (normal weight mice) was fed with an ethanolic extract of *M. chinensis* (0.40 g/kg body weight) and high-fat feed for 4 weeks, while the second (obese mice) was fed with an ethanolic extract of *M. chinensis* (0.40 g/kg body weight) and then reared on a high-fat feed for 15 days. Mice receiving NaCl 0.9% were used as the positive control, with mice receiving fenofibrate comprising the negative control group. After 4 weeks of plant extract treatment, the prevention mice group showed significantly lower TG concentration and total cholesterol/high density lipoprotein cholesterol (TC/HDL-C) levels than the control group. However, the mice receiving the plant extract did not show any significant differences in blood fat composition, compared with mice receiving fenofibrate and NaCl. The ethanolic extraction from *M. chinensis* gave protection to high-cholesterol mice [83].

Isolated flavonoid compounds, from an *M. chinensis* ethanolic extract at a concentration of 200  $\mu\text{g}/\text{mL}$ , decreased the fat increment in oleic acid (OA)-induced HepG2 cells and inhibited compound C on 5' adenosine monophosphate-activated protein kinase (AMPK). The glucose utilization of insulin induced HepG2 cells was significantly increased by MTFs and F30, compared to Metformin, which was used as the positive control [27]. The results indicated that the Mesona extract provided overweight control in a similar way to the mechanism of Metformin, which is currently used as the standard drug to treat obesity.

#### 4.5. Hypoglycemic Effect

Diabetes mellitus (DM) is a chronic disease that causes excessive blood sugar levels [84] caused by insulin secretion, insulin dysfunction, and/or both [85]. DM can be classified as insulin-dependent diabetes mellitus (IDDM) or type 1, which caused by autoimmune  $\beta$ -cell damage in the pancreas that leads to absolute insulin deficiency [86], or as non-insulin-dependent diabetes mellitus (NIDDM) or type 2, which is a metabolic disorder with variable phenotypic expressions, including  $\beta$ -cell insufficiency and insulin resistance [87]. The hypoglycemic effects and antioxidant activities generated by a diet of excessive calories in obese subjects were examined. The results indicated that *M. chinensis* (MC) extract, produced by boiling in distilled water, suppressed intestinal maltase and sucrase, with  $\text{IC}_{50}$  values of  $4660 \pm 220 \mu\text{g}/\text{mL}$  and  $1300 \pm 430 \mu\text{g}/\text{mL}$ , respectively. In contrast, the plant extract did not show inhibitory activity against pancreatic  $\alpha$ -amylase. Interestingly, 1000 mg of MC extract with a HC (high carbohydrate) diet reduced post-prandial plasma glucose, triglyceride, and MDA levels, while an increase in the plasma antioxidant capacity (FRAP and oxygen radical absorbance capacity, ORAC) of overweight subjects was noticed after treatment with MC [88]. Lim et al. examined the effect of MC on healthy Chinese men. They found that both gel and solution forms significantly reduced

glycemic and insulinemic properties, compared to the control group (who received only glucose without MC extract) [23].

An extraction of MC collected from Guangdong Raoping in Southern China exhibited the highest inhibitory effect on the  $\alpha$ -glucosidase enzyme, with an  $IC_{50}$  value of  $35.05 \pm 2.16 \mu\text{g/mL}$  [56]. The inhibition of the  $\alpha$ -glucosidase enzyme enhanced carbohydrate digestion abilities, due to decreased blood glucose levels. Compounds with lower  $IC_{50}$  values resulted in higher anticarbohydrate digestibility. The results indicated that the topography of each area impacted the biological activities of the phytochemicals and bioactive compounds.

#### 4.6. Renal Protective Activity

Hsian-tiao or *M. chinensis* water extract was given to ten Sprague Dawley female rats, together with an injection of streptozotocin (STZ) to induce diabetes. The expression of thrombospondin-1 (TSP-1) in the kidney was measured using immunohistochemistry, with significantly lower results in the plant-treated group than in the diabetic group. Kidney ultrastructural changes, assayed by transmission electron microscopy, were significantly less severe in the plant-treated group compared with the diabetic group, indicating that MC protected the kidneys of diabetic rats [21].

#### 4.7. Inflammatory Activity

Inflammation contributes to several illnesses, including rheumatoid arthritis, atherosclerosis, and asthma, by stimulating the immune system, circulation system, and various organelles within the injured cells [89]. The inflammatory mechanisms induce the up-regulation of a series of pro-inflammatory cytokines, including interleukin IL-1, tumor necrosis factor (TNF), interferon (INF)- $\gamma$ , IL-6, IL-12, IL-18, and the granulocyte-macrophage colony-activating factor [90]. The inhibitory effects of ethanolic and water extracts of *M. chinensis* on methylglyoxal (MG)-induced glycation in mice were investigated, using Western blot to analyze IL-8, MIP-2, and MCP-1 inflammation-related factors of RAW 264.7 cells. The MG-induced protein expression of IL-8, MIP-2, and MCP-1 in RAW 264.7 cells were notably reduced with ethanolic and water *M. chinensis* extracts (0.0125–0.025 mg/mL), compared with the control. Interestingly, the water extract exhibited higher anti-inflammatory activity against MG-induced inflammation [11]. The biological actions of specific MCP water extracts may play more important roles than phenolic compounds of ethanolic extracts, due to the structure and chemical composition of polysaccharides, which have at least two functional groups, such as C=O, -OH, COOH, -NR<sub>2</sub>, and -SH [91,92]. MCPs also had high uronic acid and protein contents, which may increase antioxidant activity and inhibit liver inflammation [49,93]. However, animal and clinical trials must be further performed, to gain strong evidence and scientific support.

MC polysaccharides (MCPs), extracted with boiled water at 100 °C for 2 h, alleviated pathological signs of DSS (dextran sulfate sodium)-induced colitis in mice, based on a reduction in body weight, an increase in colon length, and reduced disease activity index (DAI) scores. MCPs also improved inflammatory cell infiltration, disorganized glandular arrangement, and disrupted the intestinal structure in colonic tissues. Inflammatory cytokines showed significant increases in TNF- $\alpha$  and IL-1 $\beta$  levels and a decrease in IL-10 content. After DSS supplementation in the model control group, MCP administration reversed these expressions. MCP doses of over 0.2 g/kg/day significantly reduced the expression of phosphorylated proteins caused by MAPK/NF- $\kappa$ B signaling pathways, as analyzed by Western blot analysis [94].

#### 4.8. Gut Microbiota

The human body is lacking in gastrointestinal enzymes, and most polysaccharides cannot be directly digested by the stomach and small intestine [95,96]. Compounds that reach the large intestine are fermented by intestinal microbiota to generate short-chain fatty acids (SCFAs) and other metabolites, which play a key role in well-being [97–99].

MCPs administrated to mice at doses of 0.30 g/kg/day decreased the abundance of *Firmicutes*, *Verrucomicrobiota*, and *Proteobacteria* microbiota, but the incidence of *Bacteroidetes* increased. MCPs at high-dose levels increased *Bifidobacterium*, *Lactobacillus*, *Coprococcus*, and *Oscillospira*, while *Akkermansia*, *Clostridiaceae*, *Clostridium*, *Helicobacter*, and *Prevotella* were decreased. DSS-induced colitis in mice was improved compared with the NC (normal control) group without DSS-induced colitis. The dominant bacterial composition of the intestinal microbiota was identified using the linear discriminant analysis effect size (LEfSe) of *M. chinensis* Benth polysaccharides (MBP) treatment in a dose-dependent manner as follows: *Coprococcus* (with low dose, LD), *Akkermansia* and *Sphingomonas* (with medium dose, MD), and *Blautia* and *Dysgonomonas* (with high dose, HD) [94]. After injecting mice with CTX (cyclophosphamide) to induce liver damage and treating them with MCP concentrations ranging from 0.05 to 0.20 g/kg bw for 7 days, the results showed that, at high dosage levels (0.2 g/kg bw), serum aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were controlled, while antioxidant activity, repaired liver damage, the inhibition of inflammatory cytokines in the liver, and the concentrations of lipopolysaccharides (LPS) in the serum improved. Also, an increasing abundance of *Ruminococcaceae* and a decreasing abundance of *Bacteroidaceae* was noticed. Interestingly, concentrations of acetic acid, propionic acid, butyric acid, valeric acid, and total SCFAs caused by CTX decreased. SCFAs increased with MP treatment in a dose-dependent manner, providing evidence of the prebiotic ability of MPs to prevent liver disease [93,100]. Hong et al. confirmed that MPs treatment could promote gut microbiota and reduce liver injury caused by CTX [100]. The results identified 18 and 29 endogenous metabolites in hepatic organs and feces, respectively, after mice were treated with MPs. At least eight metabolic pathways were involved with gut health and liver improvement, including taurine and hypotaurine metabolism, phenylalanine metabolism,  $\alpha$ -linolenic acid metabolism, the tricarboxylic acid (TCA) cycle, phenylalanine, tyrosine, and tryptophan biosynthesis, arachidonic acid metabolism, and sphingolipid metabolism, as determined by ultra-high performance liquid chromatography with quadrupole time-of-flight mass spectrometry (UPLC-Q-TOF/MS).

### 5. Application of Mesona Polysaccharides as Biomaterials for Medicine

Plant polysaccharides are nontoxic, highly stable, hydrophilic, biodegradable, and biocompatible, and they pose no significant negative side effects [70,71,101]. The beneficial improvements of MCPs and other polyphenolics were investigated for their bioavailability in drug carrier systems. Quercetin was loaded into smooth spherical nanoparticles, made from zein and Z-M NPs (zein-Mesona nanoparticles) under hydrophobic, hydrogen-bonding, and electrostatic interaction. The Z-M NPs showed a higher activity with regard to in vitro anti-inflammatory activity, NO (nitric oxide), TFN- $\alpha$ , IL-1 $\beta$ , and IL-6 in RAW 264.7 cells, compared with free quercetin [101]. Curcumin was loaded into zein-MCP nanoparticles (ZMC NPs) and measured under a simulated gastrointestinal environment. The results showed higher antioxidant activity and enhanced antitumor activity, by inducing cell apoptosis, against hepatocellular carcinoma cells (HepG-2), compared with free curcumin not encapsulated by nanoparticles [102]. Interestingly, MCPs and chitosan maintained the encapsulation curcumin effect. Curcumin was released at 7% after 2 h under simulated gastrointestinal tract conditions [103]. The results showed that using MPs to encapsulate the nanoparticles improved the retention of the bioactive compounds. Therefore, the Mesona plant showed promise as an alternative material for utilization in other fields, as well as in the food industry.

### 6. Conclusions and Future Prospects

*Mesona chinensis* possesses a proximate composition, physicochemical properties, nutraceuticals, phytochemistry, and pharmacological properties. Bioactive compounds contained in this plant have been widely used in Southern China and Southeast Asia as folk medicine and indigenous food. The Mesona plant has recently attracted interest due to its phytochemical and biological activities. High variations in chemical composition

and functional properties are still challenging issues, which result from different varieties, origins, climate, harvesting times, storage, extraction processes, and determination methods. In the near future, the plant industry or smart farming will need to better understand and control this plant's growth, harvesting time, and growing conditions. Research on the Mesona plant is mainly focused on *M. chinensis* from China. The geography of each growing area may alter plant characteristics and compositions; therefore, the uniqueness of Mesona varieties from different regions must be studied in detail, to enable its ultimate utilization in functional ingredients, pharmaceuticals, and other fields, according to matched proximate compositions. The link between polysaccharide extracts and gut health is interesting and has implications for corresponding physical and mental disorders. However, meta-analysis data and safety concerns must be further investigated before therapeutic applications and innovation processes are introduced.

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## References

1. Tang, D.; Wei, F.; Cai, Z.; Wei, Y.; Khan, A.; Miao, J.; Wei, K. Analysis of codon usage bias and evolution in the chloroplast genome of *Mesona chinensis* Benth. *Dev. Genes Evol.* **2021**, *231*, 1–9. [[CrossRef](#)]
2. Huang, H.C.; Chuang, S.H.; Wu, Y.C.; Chao, P.M. Hypolipidaemic function of Hsian-tsao tea (*Mesona procumbens* Hemsl.): Working mechanisms and active components. *J. Funct. Foods* **2016**, *26*, 217–227. [[CrossRef](#)]
3. Wang, H.; Qin, L. Determination of natural benzoic acid in different *Mesona Chinensis* Benth. *China Pharm.* **2014**, *12*, 1493–1495.
4. Huang, L.; Shen, M.; Zhang, X.; Jiang, L.; Song, Q.; Xie, J. Effect of high-pressure microfluidization treatment on the physicochemical properties and antioxidant activities of polysaccharide from *Mesona chinensis* Benth. *Carbohydr. Polym.* **2018**, *200*, 191–199. [[CrossRef](#)]
5. Govaerts, R. *World Checklist of Selected Plant Families Database in ACCESS: 1-216203*; The Board of Trustees of the Royal Botanic Gardens: London, UK, 2003.
6. Suddee, S.; Paton, A.J.; Parnell, J.A.N. Taxonomic Revision of the tribe *Ocimeae* Dumort. (*Lamiaceae*) in continental South East Asia III. *Ociminae*. *Kew Bull.* **2005**, *60*, 3–75.
7. Bramley, G.L.C. *Flora Malesiana*; Noordhoff-Kolff N.V.: Djakarta, Indonesia, 2019; Volume 23, pp. 1–444.
8. Sasmita, A.O.; Ling, A.P.K. Bioactivity of *Mesona palustris* (Black Cincau) as a Nutraceutical Agent. *J. Eng. Sci. Res.* **2017**, *1*, 47–53. [[CrossRef](#)]
9. Widyaningsih, T.D.; Widjanarko, S.B.; Waziroh, E.; Wijayanti, N.; Maslukhah, Y.L. Pilot plant scale extraction of black cincau (*Mesona palustris* BL) using historical data response surface methodology. *Int. Food Res. J.* **2018**, *25*, 712–719.
10. Leelawat, B.; Permpoonchokkana, P.; Jirapornsirikun, T. Development of grass jelly processing using modified starches and higher efficient extraction method. *Int. J. Agric. Technol.* **2020**, *16*, 297–308.
11. Fan, S.L.; Lin, J.A.; Chen, S.Y.; Lin, J.H.; Lin, H.T.; Chen, Y.Y.; Yen, G.C. Effects of Hsian-tsao (*Mesona procumbens* Hemsl.) extracts and its polysaccharides on the promotion of wound healing under diabetes-like conditions. *Food Funct.* **2021**, *12*, 119–132. [[CrossRef](#)]
12. Tang, W.; Chen, X.; Liu, D.; Xie, J. Bioactive components of *Mesona Blume* and their potential health benefits. *Food Rev. Int.* **2020**, *26*, 70–85. [[CrossRef](#)]

13. Zhao, Z.G.; Shi, Y.P.; Huang, N.Z.; Fu, C.M.; Tang, F.L.; Jiang, Q.Y. The research advances on *Mesona chinensis* Benth in China. *J. South. Agric.* **2011**, *42*, 657–660.
14. Su, H.L.; Li, S.; Chen, J.Y. Research progress of *Mesona chinensis* Benth. *Res. Pract. Chin. Med.* **2008**, *22*, 79–81.
15. Tang, D.; Quan, C.; Lin, Y.; Wei, K.; Qin, S.; Liang, Y.; Wei, F.; Miao, J. Physio-Morphological, Biochemical and Transcriptomic Analyses Provide Insights into Drought Stress Responses in *Mesona chinensis* Benth. *Front. Plant. Sci.* **2022**, *13*, 809723. [[CrossRef](#)] [[PubMed](#)]
16. Le, Q.U.; Lay, H.L.; Wu, M.C. Antioxidant activities and HepG2 cells growth inhibitory capacity of whole plant ethanol extracts (*Eclipta alba* Hassk and *Mesona procumbens* Hemsl.). *J. Food Biochem.* **2018**, *42*, e12454. [[CrossRef](#)]
17. Rahmah, R.; Astuti, Y.; Salimo, H.; Pamungkasari, E.P.; Wasita, B. Beneficial Effect of *Mesona palustris* BL: A Review on Human and Animal Intervention. *J. Med. Sci.* **2022**, *10*, 171–174. [[CrossRef](#)]
18. Huang, C.Y.; Yen, G.C. Antioxidant activity of phenolic compounds isolated from *Mesona procumbens* Hemsl. *Agric. Food Chem.* **2002**, *50*, 2993–2997. [[CrossRef](#)] [[PubMed](#)]
19. Chau, C.F.; Wu, S.H. The development of regulations of Chinese herbal medicines for both medicinal and food uses. *Trends Food Sci. Technol.* **2006**, *17*, 313–323. [[CrossRef](#)]
20. Feng, T.; Biao, G.Z.; Jin, Z.Y.; Zhuang, H.N. Isolation and characterization of an acidic polysaccharide from *Mesona Blumes* gum. *Carbohydr. Polym.* **2008**, *71*, 159–169. [[CrossRef](#)]
21. Yang, M.; Xu, Z.P.; Xu, C.J.; Meng, J.; Ding, G.Q.; Zhang, X.M.; Weng, Y. Renal Protective Activity of Hsian-tsoo Extracts in Diabetic Rats. *Biomed. Environ. Sci.* **2008**, *21*, 222–227. [[CrossRef](#)]
22. Lin, L.H.; Shen, M.Y.; Liu, S.C.; Tang, W.; Wang, Z.J.; Xie, M.Y.; Xie, J.H. An acidic hetero polysaccharide from *Mesona chinensis*: Rheological properties, gelling behavior and texture characteristics. *Int. J. Biol. Macromol.* **2018**, *107*, 1591–1598. [[CrossRef](#)]
23. Lim, J.; Adisakwattana, S.; Henry, C.J. Effects of grass jelly on glycemic control: Hydrocolloids may inhibit gut carbohydrase. *Asia Pac. J. Clin. Nutr.* **2018**, *27*, 336–340. [[CrossRef](#)]
24. Xiao, Y.; Liu, S.; Shen, M.; Jiang, L.; Ren, Y.; Luo, Y.; Xie, J. Effect of different *Mesona chinensis* polysaccharides on pasting, gelation, structural properties and in vitro digestibility of tapioca starch-*Mesona chinensis* polysaccharides gels. *Food Hydrocoll.* **2020**, *99*, 105327. [[CrossRef](#)]
25. Li, Q.; Li, Y.; Li, Q.; Chen, Z.; Chen, J.; Geng, S. Evaluation of morphological and phytochemical characteristics of *Mesona chinensis* populations in southern China. *J. Plant Prod. Sci.* **2021**, *24*, 374–387. [[CrossRef](#)]
26. Li, D.Y.; Lu, G.; Wang, D.D.; Wang, M. The influence of Xiancao hypolipidemic tea on the TC and TG metabolism of the experimental rabbits. *Chin. Gen. Pract.* **2010**, *13*, 9–10.
27. Xiao, L.; Lu, X.; Yang, H.; Lin, C.; Li, L.; Ni, C.; Fang, Y.; Mo, S.; Zhan, R.; Yan, P. The Antioxidant and Hypolipidemic Effects of *Mesona Chinensis* Benth Extracts. *Molecules* **2022**, *27*, 3423. [[CrossRef](#)] [[PubMed](#)]
28. Huang, H.T.; Liaw, C.C.; Lin, Y.C.; Liao, G.Y.; Chao, C.H.; Chiou, C.T.; Kuo, Y.H.; Lee, K.T. New Diterpenoids from *Mesona procumbens* with Antiproliferative Activities Modulate Cell Cycle Arrest and Apoptosis in Human Leukemia Cancer Cells. *Pharmaceuticals* **2021**, *14*, 1108. [[CrossRef](#)] [[PubMed](#)]
29. Handayani, D.; TriDewanti, W.; Novita, W.; Mey, E.; Hanifa, H. Black Grass Jelly (*Mesona Palustris* Bl) Effervescent Powder has Anti-Dyslipidemia in High Cholesterol Diet-Fed Rats and Antioxidant Activity. *Res. J. Life. Sci.* **2017**, *4*, 159–167. [[CrossRef](#)]
30. Yang, M.; Xu, Z.; Zhang, R.; Zhan, P.; Wen, Y.; Shen, Y.; Zhang, X. Protection of myocardium in streptozotocin-induced diabetic rats by water extracts of Hsian-tsoo (*Mesona procumbens* Hemsl.). *Asia. Pac. J. Clin. Nutr.* **2008**, *17*, 23–29. [[PubMed](#)]
31. Yeh, C.T.; Huang, W.H.; Yen, G.C. Antihypertensive effects of Hsian-tsoo and its active compound in spontaneously hypertensive rats. *J. Nutr. Biochem.* **2009**, *20*, 866–875. [[CrossRef](#)]
32. Yen, G.C.; Hung, Y.L.; Hsieh, C.L. Protective effect of extracts of *Mesona procumbens* Hemsl. on DNA damage in human lymphocytes exposed to hydrogen peroxide and UV irradiation. *Food Chem. Toxicol.* **2000**, *38*, 747–754. [[CrossRef](#)]
33. Huang, G.J.; Liao, J.C.; Chiu, C.S.; Huang, S.S.; Lin, T.H.; Deng, J.S. Anti-inflammatory activities of aqueous extract of *Mesona procumbens* in experimental mice. *Sci. Food Agric.* **2012**, *92*, 1186–1193. [[CrossRef](#)] [[PubMed](#)]
34. Yen, G.; Duh, P.; Hung, Y. Contributions of Major Components to the Antimutagenic Effect of Hsian-tsoo (*Mesona procumbens* Hemsl.). *J. Agric. Food Chem.* **2001**, *49*, 5000–5004. [[CrossRef](#)] [[PubMed](#)]
35. Liu, F.L.; Feng, C.L. In vitro antibacterial test of Hsian-tsoo (*Mesona chinensis* Benth) against avian *Escherichia coli*. *Guangdong. J. Anim. Vet. Sci.* **2008**, *33*, 17–43.
36. Feng, T.; Gu, Z.B.; Jin, Z.Y. Chemical Composition and Some Rheological Properties of *Mesona Blumes* Gum. *Food Sci. Technol. Int.* **2007**, *13*, 55–61. [[CrossRef](#)]
37. Nikolopoulou, D.; Grigorakis, K.; Stasini, M.; Alexis, M.N.; Iliadis, K. Differences in chemical composition of field pea (*Pisum sativum*) cultivars: Effects of cultivation area and year. *Food Chem.* **2007**, *103*, 847–852. [[CrossRef](#)]
38. Yuris, A.; Merino, L.M.; Hardacre, A.K.; Hindmarsh, J.; Goh, K.K.T. Molecular interactions in composite wheat starch-*Mesona chinensis* polysaccharide gels: Rheological, textural, microstructural and retrogradation properties. *Food Hydrocoll.* **2018**, *79*, 1–12. [[CrossRef](#)]
39. Lai, L.S.; Tung, J.; Lin, P.S. Solution properties of hsian-tsoo (*Mesona procumbens* Hemsl) leaf gum. *Food Hydrocoll.* **2000**, *14*, 287–294. [[CrossRef](#)]
40. Lai, L.S.; Chou, S.T.; Chao, W.W. Studies on the Antioxidative Activities of Hsian-tsoo (*Mesona procumbens* Hemsl) Leaf Gum. *Agric. Food Chem.* **2001**, *49*, 963–968. [[CrossRef](#)]

41. Lai, L.S.; Liu, Y.L.; Lin, P.H. Rheological/textural properties of starch and crude hsian-tsao leaf gum mixed systems. *J. Sci. Food Agric.* **2003**, *83*, 1051–1058. [[CrossRef](#)]
42. Delattre, C.; Fenoradosa, T.A.; Michaud, P. Galactans: An overview of their most important sourcing and applications as natural polysaccharides. *Braz. Arch. Biol. Technol.* **2011**, *54*, 1075–1092. [[CrossRef](#)]
43. Elboutachfai, R.; Delattre, C.; Petit, E.; Michaud, P. Polyglucuronic acids: Structures, functions and degrading enzymes. *Carbohydr. Polym.* **2011**, *84*, 1–13. [[CrossRef](#)]
44. Singh, V.; Kumar, P.; Sanghi, R. Use of microwave irradiation in the grafting modification of the polysaccharides—A review. *Prog. Polym. Sci.* **2012**, *37*, 340–364. [[CrossRef](#)]
45. Xie, J.H.; Tang, W.; Jin, M.L.; Li, J.E.; Xie, M.Y. Recent advances in bioactive polysaccharides from *Lycium barbarum* L., *Zizyphus jujuba* Mill, *Plantago* spp., and *Morus* spp.: Structures and functionalities. *Food Hydrocoll.* **2016**, *60*, 148–160. [[CrossRef](#)]
46. Gan, L.; Wang, J.; Guo, Y. Polysaccharides influence human health via microbiota-dependent and independent pathways. *Front. Nutr.* **2022**, *9*, 1030065. [[CrossRef](#)] [[PubMed](#)]
47. Liu, S.; Xiao, Y.; Shen, M.; Zhang, X.; Wang, W.; Xie, J. Effect of sodium carbonate on the gelation, rheology, texture and structural properties of maize starch-*Mesona chinensis* polysaccharide gel. *Food Hydrocoll.* **2019**, *87*, 943–951. [[CrossRef](#)]
48. Yang, J.; Shen, M.; Wu, T.; Luo, Y.; Li, M.; Wen, K.; Xie, J. Role of salt ions and molecular weights on the formation of *Mesona chinensis* polysaccharide-chitosan polyelectrolyte complex hydrogel. *Food Chem.* **2020**, *333*, 127493. [[CrossRef](#)] [[PubMed](#)]
49. Wang, W.; Jiang, L.; Ren, Y.; Shen, M.; Xie, J. Characterizations and hepatoprotective effect of polysaccharides from *Mesona blumes* against tetrachloride-induced acute liver injury in mice. *Int. J. Biol. Macromol.* **2019**, *124*, 788–795. [[CrossRef](#)] [[PubMed](#)]
50. Lin, L.; Xie, J.; Liu, S.; Shen, M.; Tang, W.; Xie, M. Polysaccharide from *Mesona chinensis*: Extraction optimization, physicochemical characterizations and antioxidant activities. *Int. J. Biol. Macromol.* **2017**, *99*, 665–673. [[CrossRef](#)] [[PubMed](#)]
51. Chen, X.; Xiao, W.; Shen, M.; Yu, Q.; Chen, Y.; Yang, J.; Xie, J. Changes in polysaccharides structure and bioactivity during *Mesona chinensis* Benth storage. *Curr. Res. Food Sci.* **2022**, *5*, 392–400. [[CrossRef](#)]
52. Tang, W.; Shen, M.; Xie, J.; Liu, D.; Du, M.; Lin, L.; Gao, H.; Hamake, B.R.; Xie, M.Y. Physicochemical Characterization, Antioxidant Activity of Polysaccharides from *Mesona Chinensis* Benth and Their Protective Effect on Injured NCTC-1469 Cells Induced by H<sub>2</sub>O<sub>2</sub>. *Carbohydr. Polym.* **2017**, *175*, 538–546. [[CrossRef](#)]
53. Yan, L.; Xiong, C.; Xu, P.; Zhu, J.; Yang, Z.; Ren, H.; Luo, Q. Structural characterization and in vitro antitumor activity of A polysaccharide from *Artemisia annua* L. (Huang Huahao). *Carbohydr. Polym.* **2019**, *213*, 361–369. [[CrossRef](#)]
54. Xiao, Y.; Liu, S.; Shen, M.; Jiang, L.; Ren, Y.; Luo, Y.; Wen, H.; Xie, J. Physicochemical, rheological and thermal properties of *Mesona chinensis* polysaccharides obtained by sodium carbonate assisted and cellulase assisted extraction. *Int. J. Biol. Macromol.* **2018**, *126*, 30–36. [[CrossRef](#)]
55. Huang, L.; Huang, M.; Shen, M.; Wen, P.; Wu, T.; Hong, Y.; Yu, Q.; Chen, Y.; Xie, J. Sulfated modification enhanced the antioxidant activity of *Mesona chinensis* Benth polysaccharide and its protective effect on cellular oxidative stress. *Int. J. Biol. Macromol.* **2019**, *136*, 1000–1006. [[CrossRef](#)]
56. Huang, J.; Ding, L.; Tian, W.; Zhi, H.; Chen, J.; Wu, L.; Wang, L.; Xie, J.; Bai, J.; Fan, H.; et al. Polyphenolic profiling, antioxidant properties, and inhibition of  $\alpha$ -glucosidase of *Mesona chinensis* benth from Southern China. *J. Microchem.* **2021**, *168*, 106399. [[CrossRef](#)]
57. Yen, G.C.; Hung, C.Y.; Chen, Y.J. Antioxidant Properties of Hsian-tsao (*Mesona procumbens* Hemsl.). *Orient. Food Herb.* **2003**, *859*, 202–214. [[CrossRef](#)]
58. ElSamahy, S.K.; Abd El-Hady, E.A.; Habiba, R.A.; Moussa-Ayoub, T.E. Some Functional, Chemical, and Sensory Characteristics of Cactus Pear Rice Based Extrudates. *J. Prof. Assoc. Cactus.* **2007**, *9*, 136–147.
59. Ren, Y.; Jiang, L.; Wang, W.; Xiao, Y.; Liu, S.; Luo, Y.; Shen, M.; Xie, J. Effects of *Mesona chinensis* Benth polysaccharide on physicochemical and rheological properties of sweet potato starch and its interactions. *Food Hydrocoll.* **2020**, *99*, 105371. [[CrossRef](#)]
60. Zhuang, H.; Feng, T.; Xie, Z.; Toure, A.; Xu, X.; Jin, Z.; Su, Q. Effect of *Mesona Blumes* gum on physicochemical and sensory characteristics of rice extrudates. *Int. J. Food Sci. Technol.* **2010**, *45*, 2415–2424. [[CrossRef](#)]
61. Wongverawattanakul, C.; Suklaew, P.; Chusak, C.; Adisakwattana, S.; Thilavech, T. Encapsulation of *Mesona chinensis* Benth Extract in Alginate Beads Enhances the Stability and Antioxidant Activity of Polyphenols under Simulated Gastrointestinal Digestion. *Foods* **2022**, *11*, 2378. [[CrossRef](#)]
62. Iguchi, C.; Nio, Y.; Takeda, H.; Yamasawa, K.; Hirahara, N.; Toga, T.; Tamura, K. Plant polysaccharide PSK: Cytostatic effects on growth and invasion; modulating effect on the expression of HLA and adhesion molecules on human gastric and colonic tumor cell surface. *Anticancer Res.* **2000**, *21*, 1007–1013.
63. Cai, W.; Xie, L.; Chen, Y.; Zhang, H. Purification, characterization and anticoagulant activity of the polysaccharides from green tea. *Carbohydr. Polym.* **2013**, *92*, 1086–1090. [[CrossRef](#)]
64. Xie, J.H.; Xie, M.Y.; Nie, S.P.; Shen, M.Y.; Wang, Y.X.; Li, C. Isolation, chemical composition and antioxidant activities of a water-soluble polysaccharide from *Cyclocarya paliurus* (Batal.) Iljinskaja. *Food Chem.* **2010**, *119*, 1626–1632. [[CrossRef](#)]
65. Simpson, R.; Morris, G.A. The anti-diabetic potential of polysaccharides extracted from members of the cucurbit family: A review. *Bioact. Carbohydr. Diet. Fibre* **2014**, *3*, 106–114. [[CrossRef](#)]
66. Wang, C.; Li, W.; Chen, Z.; Gao, X.; Yuan, G.; Pan, Y.; Chen, H. Effects of simulated gastrointestinal digestion in vitro on the chemical properties, antioxidant activity,  $\alpha$ -amylase and  $\alpha$ -glucosidase inhibitory activity of polysaccharides from *Inonotus obliquus*. *Food Res. Int.* **2018**, *103*, 280–288. [[CrossRef](#)]

67. Tzianabos, A.O. Polysaccharide immunomodulators as therapeutic agents: Structural aspects and biological function. *Clin. Microbiol. Rev.* **2000**, *13*, 523–533. [[CrossRef](#)]
68. Lee, J.S.; Synytsya, A.; Kim, H.B.; Choi, D.; Lee, S.; Lee, J.; Kim, W.J.; Jang, S.; Park, Y. Purification, characterization and immunomodulating activity of a pectic polysaccharide isolated from Korean mulberry fruit Oddi (*Morus alba* L.). *Int. Immunopharmacol.* **2013**, *17*, 858–866. [[CrossRef](#)]
69. Li, C.; Dong, Z.; Zhang, B.; Huang, Q.; Liu, G.; Fu, X. Structural characterization and immune enhancement activity of a novel polysaccharide from *Moringa oleifera* leaves. *Carbohydr. Polym.* **2020**, *234*, 115897. [[CrossRef](#)]
70. Xu, H.S.; Wu, Y.W.; Xu, S.F.; Sun, H.X.; Chen, F.Y.; Yao, L. Antitumor and immunomodulatory activity of polysaccharides from the roots of *Actinidia eriantha*. *J. Ethnopharmacol.* **2009**, *125*, 310–317. [[CrossRef](#)]
71. Xie, J.H.; Shen, M.Y.; Nie, S.P.; Zhao, Q.; Li, C.; Xie, M.Y. Separation of water-soluble polysaccharides from *Cyclocarya paliurus* by ultrafiltration process. *Carbohydr. Polym.* **2014**, *101*, 479–483. [[CrossRef](#)]
72. Aruoma, I.O. Free radicals, oxidative stress and antioxidants in human health and disease. *J. Am. Oil. Chem. Soc.* **1998**, *75*, 199–212. [[CrossRef](#)]
73. Uttara, B.; Singh, A.V.; Zamboni, P.A.; Mahajan, R.T. Oxidative stress and neurodegenerative diseases: A review of upstream and downstream antioxidant therapeutic options. *Curr. Neuropharmacol.* **2009**, *7*, 65–74. [[CrossRef](#)]
74. Liu, Z.; Ren, Z.; Zhang, J.; Chuang, C.C.; Kandaswamy, E.; Zhou, T.; Zuo, L. Role of ROS and Nutritional Antioxidants in Human Diseases. *Front. Physiol.* **2018**, *9*, 477. [[CrossRef](#)]
75. Alfadda, A.A.; Sallam, R.M. Reactive Oxygen Species in Health and Disease. *J. Biomed. Biotechnol.* **2012**, *2012*, 936486. [[CrossRef](#)]
76. Newman, D.J.; Cragg, G.M. Natural products as sources of new drugs over the 30 years from 1981 to 2010. *Nat. Prod.* **2012**, *75*, 311–335. [[CrossRef](#)]
77. Bon, R.S.; Waldmann, H. Bioactivity-guided navigation of chemical space. *Acc. Chem. Res.* **2010**, *43*, 1103–1114. [[CrossRef](#)]
78. Widyaningsih, T.D. Cytotoxic Effect of Water, Ethanol and Ethyl Acetate Extract of Black Cincau (*Mesona Palustris* BL) against HeLa Cell Culture. *APCBEE Procedia* **2012**, *2*, 110–114. [[CrossRef](#)]
79. Lin, Y.H.; Chang, Y.X.; Chen, C.C.; Chen, H.Y.; Hung, Y.C.; Chi, T.Y.; Lin, C.Y.; Chen, G.H.; Huang, P.M.; Wang, Y.P.; et al. Effects of *Mesona chinensis* ethanolic extracts and commercial herbal tea on the cell viability of colorectal cancer cells. *GSC. Biol. Pharm. Sci.* **2022**, *18*, 326–330. [[CrossRef](#)]
80. Chandra, K.S.; Bansal, M.; Nair, T.; Iyengar, S.S.; Gupta, R.; Manchanda, S.C.; Mohanan, P.P.; Rao, V.D.; Manjunath, C.N.; Sawhney, J.P.S.; et al. Consensus statement on management of dyslipidemia in Indian subjects. *Indian Heart J.* **2014**, *66*, S1–S51. [[CrossRef](#)]
81. Rauf, A.; Akram, M.; Anwar, H.; Daniyal, M.; Munir, N.; Bawazeer, S.; Bawazeer, S.; Rebezov, M.; Bouyahya, A.; Ali Shariati, M.; et al. Therapeutic potential of herbal medicine for the management of hyperlipidemia: Latest updates. *Environ. Sci. Pollut. Res. Int.* **2022**, *29*, 40281–40301. [[CrossRef](#)]
82. Orekhov, A.N.; Ivanova, E.A. Conventional, traditional and alternative therapies for cardiovascular disorders. Part 2: Traditional therapy. *Curr. Pharm. Des.* **2017**, *23*, 967–968. [[CrossRef](#)]
83. Thao, N.T.P.; Thu, N.T.; Hanh, N.T.H. Hypolipidemic effect of ethanol extract from *Mesona chinensis* Benth. in high fat diet-induced obesity mice. *VNU J. Sci. Med. Pharm. Sci.* **2019**, *35*, 37–43. [[CrossRef](#)]
84. Lin, Y.; Sun, Z. Current views on type 2 diabetes. A review. *J. Endocrinol.* **2010**, *204*, 1–11. [[CrossRef](#)]
85. American Diabetes Association. Diagnosis and classification of diabetes mellitus. *Diabetes Care* **2013**, *36* (Suppl. 1), S67–S74. [[CrossRef](#)]
86. Eizirik, D.L.; Colli, M.L.; Ortis, F. The role of inflammation in insulinitis and  $\beta$ -cell loss in type 1 diabetes. *Nat. Rev. Endocrinol.* **2009**, *5*, 219–226. [[CrossRef](#)]
87. Reinehr, T. Type 2 diabetes mellitus in children and adolescents. *World J. Diabetes* **2013**, *4*, 270. [[CrossRef](#)]
88. Chusak, C.; Thilavech, T.; Adisakwattana, S. Consumption of *Mesona chinensis* Attenuates Postprandial Glucose and Improves Antioxidant Status Induced by a High Carbohydrate Meal in Overweight Subjects. *Am. J. Chin. Med.* **2014**, *42*, 315–336. [[CrossRef](#)]
89. Pan, M.H.; Chiou, Y.S.; Tsai, M.L.; Ho, C.T. Anti-inflammatory activity of traditional Chinese medicinal herbs. *Tradit. Complement. Med.* **2011**, *1*, 8–24. [[CrossRef](#)]
90. Mueller, M.; Hobiger, S.; Jungbauer, A. Anti-inflammatory activity of extracts from fruits, herbs and spices. *Food Chem.* **2010**, *122*, 987–996. [[CrossRef](#)]
91. Yuan, Y.V.; Bone, D.E.; Carrington, M.F. Antioxidant activity of dulse (*Palmaria palmate*) extract evaluated in vitro. *Food Chem.* **2005**, *91*, 485–494. [[CrossRef](#)]
92. Zeng, B.Y.; Su, M.H.; Chen, Q.X.; Chang, Q.; Wang, W.; Li, H.H. Protective effect of a polysaccharide from *Anoectochilus roxburghii* against carbon tetrachloride induced acute liver injury in mice. *J. Ethnopharmacol.* **2017**, *200*, 124–135. [[CrossRef](#)]
93. Hong, Y.; Shen, M.; Huang, L.; Wu, T.; Xie, J. *Mesona chinensis* Benth. Polysaccharides alleviates liver injury by beneficial regulation of gut microbiota in cyclophosphamide-induced mice. *Food Sci. Hum. Wellness* **2022**, *11*, 74–84. [[CrossRef](#)]
94. Lu, H.; Shen, M.; Chen, T.; Yu, Y.; Chen, Y.; Yu, Q.; Chen, X.; Xie, J. *Mesona chinensis* Benth Polysaccharides Alleviate DSS-Induced Ulcerative Colitis via Inhibiting of TLR4/MAPK/NF- $\kappa$ B Signaling Pathways and Modulating Intestinal Microbiota. *Mol. Nutr. Food Res.* **2022**, *66*, e2200047. [[CrossRef](#)]
95. Chen, G.; Xie, M.; Wan, P.; Chen, D.; Ye, H.; Chen, L.; Zeng, X.; Liu, Z. Digestion under saliva, simulated gastric and small intestinal conditions and fermentation in vitro by human intestinal microbiota of polysaccharides from Fuzhuan brick tea. *Food Chem.* **2018**, *244*, 331–339. [[CrossRef](#)]

96. Shang, Q.; Jiang, H.; Cai, C.; Hao, J.; Li, G.; Yu, G. Gut microbiota fermentation of marine polysaccharides and its effects on intestinal ecology: An overview. *Carbohydr. Polym.* **2018**, *179*, 173–185. [[CrossRef](#)]
97. Jha, R.; Berrocoso, J.D. Dietary fiber utilization and its effects on physiological functions and gut health of swine. *Animal* **2015**, *9*, 1441–1452. [[CrossRef](#)]
98. Krautkramer, K.A.; Fan, J.; Backhed, F. Gut microbial metabolites as multikingdom intermediates. *Nat. Rev. Microbiol.* **2020**, *19*, 77–94. [[CrossRef](#)]
99. Song, Q.; Wang, Y.; Huang, L.; Shen, M.; Yu, Y.; Yu, Q.; Chen, Y.; Xie, J. Review of the relationships among polysaccharides, gut microbiota, and human health. *Food Res. Int.* **2021**, *140*, 109858. [[CrossRef](#)]
100. Hong, Y.; Shen, M.; Yu, Q.; Chen, Y.; Xie, J. UPLC-Q-TOF/MS-based metabolomics reveals modulatory effects of *Mesona chinensis* Benth polysaccharide in liver injury mice induced by cyclophosphamide. *Food Sci. Hum. Wellness* **2023**, *12*, 584–595. [[CrossRef](#)]
101. Yang, J.; Lin, J.; Zhang, J.; Chen, X.; Wang, Y.; Shen, M.; Xie, J. Fabrication of Zein/*Mesona chinensis* Polysaccharide Nanoparticles: Physical Characteristics and Delivery of Quercetin. *ACS Appl. Bio Mater.* **2022**, *5*, 1817–1828. [[CrossRef](#)]
102. Yang, J.; Lin, J.; Chen, X.; Rong, L.; Shen, M.; Wang, Y. *Mesona chinensis* polysaccharide/zein nanoparticles to improve the bioaccessibility and in vitro bioactivities of curcumin. *Carbohydr. Polym.* **2022**, *295*, 119875. [[CrossRef](#)] [[PubMed](#)]
103. Yang, J.; Chen, X.; Wen, H.; Chen, Y.; Yu, Q.; Shen, M.; Xie, J. Curcumin-Loaded pH-Sensitive Biopolymer Hydrogels: Fabrication, Characterization, and Release Properties. *ASC Food Sci. Technol.* **2022**, *2*, 512–520. [[CrossRef](#)]

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