



Review Selenium Biofortification: Strategies, Progress and Challenges

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Abstract: Selenium (Se) is an essential trace element for humans and animals. Its necessity for plants is still under examination. Due to the contradictory nature of Se and its significance, it has received much interest in recent years. Se deficiency can be harmful to humans, yet almost a billion people are deficient. Its deficiency has been associated with cancers, impairment of organs, and a number of other ailments. The biofortification of plants and livestock is a guaranteed practice to increase human selenium consumption. Strategies such as foliar spraying, the direct application of Se in plants and Se feed, and injections in livestock have been employed. Se biofortification has been shown to have additional beneficial effects in plants and livestock. In plants, it has been reported to mitigate different types of stress and increase yield. In animal biofortification, Se has been shown to reduce the detrimental effects of ailments and promote healthy growth. Se biofortification, nevertheless, confronts a number of difficulties. For instance, the bulk of biofortified products must be prepared before consumption, lowering the Se concentration. The objective of this review is to convey the current understanding of the Se biofortification of plants and animals, as well as its difficulties, taking into account both the detrimental consequences of Se deficiency and benefits of Se biofortification.

Keywords: selenium; biofortification; plants; livestock; food; humans

1. Introduction

Selenium (Se) is mainly generated as a byproduct of copper mining [1]. Since its industrial use began in the early 1900s, the global output of Se has expanded significantly. Worldwide production in 1910 was around 5000 kg [1]. According to Garside, about 3300 metric tons of Se was produced globally in 2020. China, Japan, and Germany produced the most selenium that year, producing 1120, 740 and 300 metric tons, respectively [2].

Se belongs to a group of elements that cannot be classified distinctly as either metals or non-metals. It is found in the group VIA as a partner to sulfur (S). Se types are determined by the potential of hydrogen (pH) and measurement of electrical potential [3,4]. It exists in nature in four oxidation states: elemental selenium (Se(0)), selenide (Se(II)), selenite (Se(IV)), and selenate (Se(VI)) [5–7]. According to previous studies [8,9], Marco Polo initially described Se poisoning in the 13th century. However, it was not until research by Schwarz and Foltz that Se's essential function in preventing liver damage in rats was recognized [10].

In the human body, Se plays a vital function as a component of enzymes [11]. Selenium's relevance is attributed to its presence in selenoproteins [12,13]. The significance of Se is also demonstrated in its ability to change the expression and activity of over 25 selenoproteins involved in oxidative stress, detoxification, transport processes, metabolism, and inflammatory responses [14,15]. Although essential, Se is termed as a "two-edged



Citation: Danso, O.P.; Asante-Badu, B.; Zhang, Z.; Song, J.; Wang, Z.; Yin, X.; Zhu, R. Selenium Biofortification: Strategies, Progress and Challenges. *Agriculture* 2023, *13*, 416. https:// doi.org/10.3390/agriculture13020416

Academic Editor: Massimiliano Renna

Received: 4 January 2023 Revised: 7 February 2023 Accepted: 8 February 2023 Published: 10 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). sword" [16] because of its ability to be both beneficial and detrimental at different concentrations [17]. Se-deficient diets have been linked to various health problems [18], and they are common in many parts of the world [19]. At least one billion individuals globally are Se deficient [20,21]. For instance, the consumption of Se in China was found to be approximately 26.63 μ g/d, which is very low [22]. Due to the fact that Se is a non-renewable resource, this worrying situation is expected to get worse in the future.

To increase the Se status in humans and reduce the effects of Se deficiency, the production of crops and animals with increased Se levels is vital. Selenium's necessity for plants is still up for contention [23,24]. Nonetheless, several studies have shown that Se is beneficial for plants and animals [8,25–32]. The biofortification of plants and animals with Se has been researched using multiple techniques including genetic biofortification and the application of selenium fertilizers. Although some are already well known, others are constrained by government regulations and other factors. This review's goal is to provide an overview of the state of Se biofortification research, methods, effects, and challenges. The review focuses on (i) Se biofortification of plants and animals; (ii) Se biofortification and human health benefits; and (v) Se biofortification challenges.

2. Methodology

In this study, a comprehensive search was conducted on the World Wide Web for published, peer-reviewed research and review articles utilizing a variety of databases and search engines, including but not limited to Google Scholar, Web of Science, PubMed, Science Direct, Scopus, Directory of Open Access Journals, and MEDLINE. These databases are well-known collections of peer-reviewed articles and widely used. Keywords, index terms, and combinations thereof, such as selenium, selenium biofortification strategies, selenium biofortification, selenium in plants and animals, effects of selenium on plants and animals, selenium overdose, and selenium benefits, were utilized. Over 1000 studies were discovered using the specified keywords and index terms. The list of studies was then scrutinized and duplicates were removed using endnote, leaving 316 articles. To further uncover related studies, relevant papers cited in the selected publications were reviewed.

3. Sources and Pathways of Se

Se sources can be anthropogenic, geogenic, or both [33]. Gypsum, marlstone, volcanic eruptions, sea spray, the weathering of Se-rich rocks, soils, and animal transport are some of the natural sources of Se [34–36]. Atmospheric discharge is one of the most significant sources of Se in different types of soils, as natural resources volatilize Se into the atmosphere [37,38]. In clay soils, Se levels range from 0.8 to 2 mg/kg, whereas tropical soils have a range of 2 to 4.5 mg/kg [39]. In diverse soils, however, Se levels vary from 0.01 to 2 mg/kg [16]. Some studies show that Se accumulates more readily in igneous rocks than in other rock types [35–40].

Se from sediments is transported into rivers and other water bodies by fluctuations in water flow or benthic agitation (Figure 1). In certain areas, Se levels in water from wells and subsurface waters used by humans and livestock for drinking and other activities may surpass $10-20 \ \mu g/L$, with some concentrations reaching hundreds of micrograms per liter [41]. These waters are not often thought of as an excellent source of Se [42]. Nonetheless, their use results in the transfer and transport of the element in the environment. Farming and industrial activities are the main anthropogenic sources and pathways of Se [43]. However, only around 5% of the overall demand for Se is used by agriculture [44], where it is used in producing fertilizers and animal feeds, among other uses. This renders industrial use the primary anthropogenic source and pathway of Se.

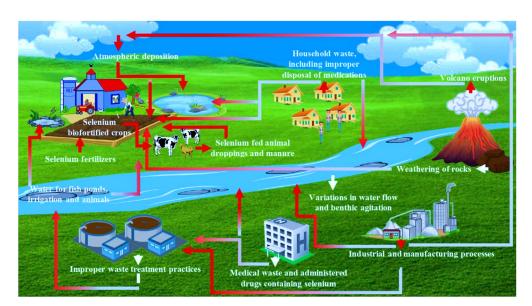


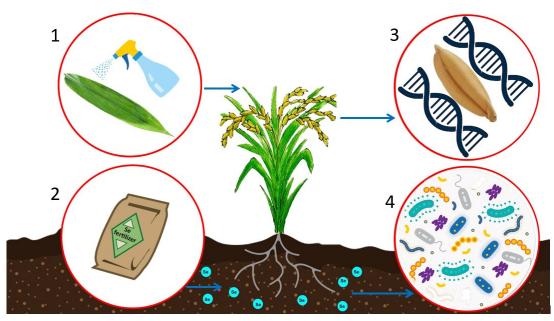
Figure 1. Sources and pathways of selenium in the agro-environment.

4. Se Biofortification Strategies

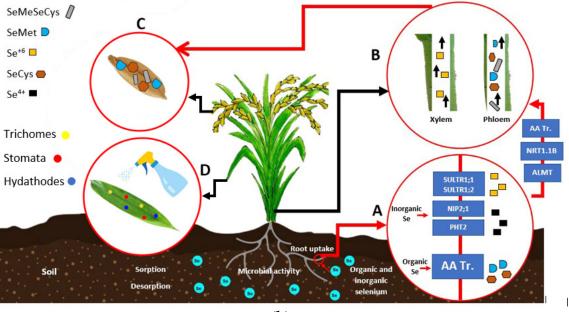
Biofortification is a quick, efficient, and sustainable way to lower micronutrient deficiencies [45]. The strategies of biofortification are the approaches employed to achieve biofortification (Figure 2a). Plants and animal produce are classified as biofortified when there is an increase in the Se content in the edible portions of plants and food animals.

4.1. Se Forms

As noted earlier, Se exists in soils in four different oxidation states that determine its behavior, specifically its mobility and bioavailability in the natural environment. Some common organic Se compounds include selenomethionine (SeMet), selenocysteine (SeCys), dimethylselenide, selenium methylselenocysteine, dimethyldiselenide, dimethylselenone, methane selenol, and dimethylselenyl sulfide [46]. Se(IV) and Se(VI) are the dominant forms of Se and are often considered the only abundant forms for plant uptake in many studies [47,48]. Se(VI) is water soluble, while Se(IV) is less water soluble and more attached to soil minerals and organic matter [47]. Metallic Se(II) and Se(0) are generally not watersoluble [48]. Se(IV) and Se(VI) are considered the most bioavailable forms for plant uptake due to their solubility [49]. Se(VI) has a higher rate of translocation from roots to shoots compared with Se(IV) [50]. This is because Se(IV) is quickly converted into organic forms like SeCys or SeMet in roots [51]. In anaerobic soils, Se(0) and organic Se(II) are the dominant forms, while Se(IV) and Se(VI) are common in aerobic soils [52]. Se(0) and metallic Se(II) are not water-soluble and, therefore, not bioavailable for plant uptake [52]. Under low redox potential conditions, Se(IV) and Se(VI) can be reduced to Se(II) and Se(0) [53]. Se(0) can also be oxidized into bioavailable inorganic Se compounds through microbial oxidation and hydrolysis [54]. The uptake and transport of Se by plants varies among species and genotypes. The mobility of Se in wheat and canola plants is in the following order: selenate > SeMet > selenite/SeCys [55]. Studies show that rice grains contain higher amounts of Se compared with maize and wheat grains [56]. This may be due to the existence of high-Se and low-Se varieties of rice [57]. The use of Nano-Se to increase the Se content in food is considered a potential solution due to its high biological activity, bioavailability, low toxicity, and large surface area [58]. The use of Se nanoparticles is a promising alternative to other forms of Se as it simplifies application and leads to improved antioxidant metabolism, agronomic sustainability, and waste reduction [59].



(a)



(**b**)

Figure 2. (a) Selenium biofortification strategies. Note: 1. Foliar application; 2. Soil application; 3. Genetic biofortification; 4. Microbial-assisted biofortification. (b) Selenium uptake and transport. (A) Amino acid permeases (AA Tr.) and SULTR1;1, SULTR1;2, NIP2;1, PT2, and PT8 transporters aid organic and inorganic Se absorption by the roots. (B) The shoots receive organic Se forms from the AA Tr. transporter. (C) Selenate is transported by both the xylem and phloem; concomitantly, organic Se compounds enter the seed via the phloem. (D) Upon foliar application, Se enters the plants via trichomes, stomata, and hydathodes.

In livestock production, Se is added to animal feed in both organic and inorganic forms [60]. Ruminants absorb and retain organic forms of Se more effectively than inorganic forms. A common way to enhance animal diets with Se is through in-feed administration of Se-enriched yeast, which has a moderate to high Se content and is a source of SeMet [61]. A safe and natural way to provide animals with Se is by offering feed with optimal Se content, as long as the level of Se in the dry matter is carefully monitored. Plants accumulate Se

primarily in the inorganic form and then synthesize seleno-amino acids in SeMet, becoming a source of organic Se for animals [62].

4.2. Se Biofortification Strategies in Plants

4.2.1. Foliar Application

Foliar application appears to be the most popular method of applying selenium among all methods because of its simplicity and preferable outcomes. The danger of environmental contamination also seems to be lower. Studies have demonstrated that foliar spray entails minimal use of Se salts [63,64]. This technique entails spraying a crop's leaf surface with a Se-containing solution. Selenium enters the plant through the leaf cuticles. Particles can also enter plants through trichomes, stomata, stigma, and hydathodes [65]. In this respect, soil chemistry and microbiological processes have less of an impact on Se, resulting in a higher absorption rate with modest quantities of administered Se solution. With this strategy, there are changes in plant-specific parameters that must be taken into account, including the quantity of Se applied, leaf area and surface structure, and leaf structure. Wang et al. [66] recently found that applying Se as Se(IV) or Se(VI) using foliar spray during the prefilling stage has a substantial influence on Se concentration in wheat grains. In a previous rice study, foliar application of Se (30–300 μ g Se/ha as SeO₃^{2–} or SeO₄^{2–}) raised the Se concentration and other bioactive molecules in rice grains [67]. Per Lidon's recent report, foliar SeO_3^{2-} fertilization caused a 427–884-fold increase in grain Se content in four rice genotypes, while SeO_4^{2-} application led to a 128–347-fold increase in grain Se concentrations [68]. Pannico et al. [69] found that foliar spraying of Se (0–40 μ M) enhanced leaf Se content in two lettuce cultivars with varying pigmentation, with the red cultivar storing 57% more Se than the green cultivar. However, all treatments decreased the fresh weight of green lettuce by 9%, whereas 32 and 40 μ M lowered the fresh weight of red lettuce by 11% and 22%, respectively.

4.2.2. Soil Application

This technique involves amending the soil with Se to raise the amount of overall or bioaccessible Se, enhance the rhizosphere conditions for soil crops, and raise the Se content of produce. With this approach, Se is applied either as Se salts, Se solution, or Secontaining fertilizers. Soil chemistry and microbial activities affect whether Se administered with this technique will result in a desired effect. This strategy is said to have been employed by the Finnish government to boost the population's daily consumption of selenium [70,71]. Soil Se application has been shown to have a favorable influence on various plant physiological systems. Plants absorb Se in the form of organic Se (SeCys and SeMet), Se(IV), and Se(VI) [72,73]. Although plant roots cannot absorb Se(II), they may do so for organic Se species such as SeCys and SeMet and inorganic Se species such as Se(0), Se(IV), and Se(VI) [74,75]. Se (VI) has been shown to enter plants through the sulfate transporters SULTR1; 2 and SULTR1 [72], while Se(IV) enters plants via phosphate transporter transport [76,77]. OsPT2, a phosphate transporter, has been demonstrated to be involved in plant uptake of Se(IV) [78]. Se in the form of Se(VI) applied via soil was the best strategy for increasing Se content in the radish without causing damage to biomass growth. The researchers found that the accumulation of Se in the leaf, root, and whole plants was higher when Se was applied via soil compared with the foliar application [79]. This result is consistent with the findings of a recent study with the same Se application strategy, which found that Se considerably enhanced the Se concentration in mushroom fruit bodies (p < 0.05) [80]. However, there was no significant increase in fruit production. Consistent with that study is one that reported a rise in Se concentrations with the same Se application strategy but also no increase in yield [81].

4.2.3. Microbial-Assisted Biofortification

Agronomic biofortification strategies are not always successful due to a number of factors including impromptu rainfall in the case of foliar applications and pH, and heavy

metals in soil applications. Plant growth characteristics and yield have been documented to be influenced by microorganisms located in the rhizosphere via a range of processes. These processes include the release of hormones, nutrient transformations, and stress mitigation [82,83]. The roles played by microorganisms in this respect may be species and/or Se species and bioavailability dependent. Bacterial species such as Bacillus, Entrobacter, *Paenibacillus*, and *Pseudomonas* have been shown to be capable of Se transformations via methylation and oxidation reduction processes [52,84]. Previous studies found that inoculating wheat with Se-tolerant bacteria derived from Se-deficient soils increased tissue Se accumulation [85]. Researchers demonstrated in 2015 that various bacterial consortia increased Se concentrations in Indian mustard growing in seleniferous soil [86]. A study in 2019 indicated that in a test of two Se forms (SeCys and SeO_4^{2-}) in shallots with and without inoculation of arbuscular mycorrhizal fungi, inoculation increased the concentration of Se in the bulb by more than five times [87]. Recently, Enterobacter sp. EG16 (7.65107 CFU/mL) was observed to promote the growth and development of pak choi. Chlorophyll concentration, SOD, CAT, and POD activity were likewise increased by the same Se and EG16 doses [88]. These investigations demonstrate the essential roles microbes play in Se biofortification. The study of bioreduction of selenate or selenite using microorganisms such as bacteria, fungi, and plant extracts has become a popular area of interest for scientists [89]. Microbes have been shown to produce the purest form of Se, with previous research demonstrating the production of Se(0) by the anaerobic bacterium Bacillus selenireducens [90]. Yeast and other microbes play a vital role in synthesizing Se-containing compounds such as SeCys and SeMet [91]. According to several authors' studies on SeMet determination, it may make up to 90% of the total Se content in yeast cells [92,93].

4.2.4. Genetic Biofortification

Genetic engineering has the capability to enhance the capacity of plants to accumulate selenium as an alternative to agronomic approaches. Nevertheless, due to the stringent limitations on the usage of transgenics that are still present in several nations, genetic engineering is still not as prevalent and recognized as agronomic biofortification [94]. However, various chromosomal loci linked to elevated Se accumulation in a number of crops have been reported [74,95,96]. Marker-assisted breeding can be employed to transfer high-Se chromosomal loci from high-yielding, low-Se edible plant varieties into the breeding population [75]. According to Schiavon et al., a significant drawback of conventional or marker-assisted plant breeding is that it must be supplemented with agronomic biofortification treatments employing Se fertilizers when crops are grown in low Se regions [97]. The majority of plants targeted by this method are staple crops [98]. According to a previous study, double-transgenic crops produced from crossed-transgenic mustard greens absorbed up to nine times more selenium than wild-type plants [99]. By using the low Cd replacement line CSSL^{GCC7} as the breeding material, researchers at the China National Rice Research Institute recently reported that CSSL^{GCC7+GSC5} demonstrated increased Se concentrations in grains when crossed with CSSLs containing other major quantitative trait loci for essential mineral elements [100].

4.2.5. Crop Breeding

Some researchers believe that conventional crop breeding may be a sustainable and long-term approach to crop biofortification with Se [101]. However, compared to genetic biofortification crop breeding is a slower and less accurate method as this procedure is generally executed by hand. For instance, one study lasted for about five years [102]. Additionally, establishing appropriate and viable genotypic variation may be difficult [103]. Nevertheless, it can be utilized to create new plant types with enhanced features. Crop breeding for Se biofortification uses the conventional procedure of cross-pollinating two separate plants to develop a new hybrid plant with a mix of features from both parents to promote Se absorption and translocation to edible portions of the crop. The researchers' goal in the above-mentioned study was to breed Se-rich red glutinous rice and evaluate the

concentration of Se and protein in various parts of the rice. The red glutinous rice attained an Se concentration of 121.75 ng/g (\pm 3.01 ng/g) after five years of breeding. According to the results, upwards of 80% of the Se in the grain was organic Se, and over half of the total Se was in the endosperm [102]. The study suggests that plant breeding for Se biofortification was successful and supplementary research is needed in the future.

4.3. Se Biofortification Strategies in Livestock

In livestock, biofortification comprises employing agronomic or biotechnological techniques to increase the quantity of vital nutrients in edible sections of animals [104]. Se fertilization of farmlands, dietary supplementation via feed concentrate rations, and direct administration, including injections, are viable supplementation techniques. High Se feed concentrations are expected to raise Se concentrations in livestock. Animals can generate a variety of selenoproteins, including glutathione peroxidase, selenoprotein P, selenoprotein W, thioredoxin reductase, and other iodothyronine deiodinases, using absorbed selenium forms [105]. However, excessive Se ingestion in livestock (5–50 mg per kg of mass) may cause alkali disease, characterized by hoof deformities, a lack of vitality, anemia, and stiffness [106].

5. Se Biofortification in Plants

Even though livestock products contain significantly higher amounts of Se, crops are relatively good sources of Se due to their greater bioavailability [107]. While organically cultivated foods are regarded as safe and wholesome, their low Se content may have an unfavorable effect on their appearance [108]. Se supplementation of diets is essential to control Se deficiency [109]. Se fertilization should be performed once every planting season since plant supply can only be maintained for one growing season during crop production for biofortification [110,111]. Sorption, desorption, precipitation, dissolution, production of inorganic and organic complexes, methylation to volatile Se compounds [112,113], and microbiological activity [114] all influence Se mobility and availability to plants (Figure 2b). The amount of Se in a plant is determined mainly by the plant species; soil type in which the plant is produced; use of herbicides, manure, and fertilizers; and agro-ecological management practices [93,115]. Some researchers found that vegetables contain approximately 6 mg/g Se when grown in seleniferous soil; however, asparagus and onions can accumulate up to 17 mg/g Se when grown in similar soils [116].

Se is a valuable element for plants because it stimulates plant development (Table 1) [23,31,32,117]. A recent study [118] discovered that Se improved the plants' agronomic parameters when applied alone. Se application techniques influence whether the element impacts plant development and biofortification. Selecting the appropriate chemical form (Na₂SeO₄ or Na₂SeO₃) [119] and application approach is a crucial step for achieving desirable effects and efficient biofortification.

5.1. Se Biofortification Effects in Plants

Plants have evolved a system in response to oxidative stress. It is an enzymatic antioxidant complex that uses antioxidant enzymes such as superoxide dismutase (SOD), peroxidase (POD), and catalase (CAT) to control oxidative stress [120]. For instance, malondialdehyde (MDA), an oxidized output of membrane lipids, is an indicator of the degree of oxidative stress and lipid peroxidation. Se was shown to modulate antioxidant enzyme activity [120,121]. The primary mechanisms used by Se include the prevention of plant peroxidation, restoration of cell membrane integrity and function, modification of antioxidant enzyme activity, and repair and rebuilding of chloroplast [122].

5.1.1. Se and Salinity

When plants are grown in soils with high levels of salt, their development and growth are stifled. However, the specific mechanism of Se-mediated salinity tolerance is not fully understood. Nonetheless, scientists frequently employ the usage of mineral elements to

enhance crop tolerance to salt-induced stress [122]. Selenium efficacy in preventing such stress has been documented in a number of publications. Onions grown on silt loam soil with a salinity of 8 dS/m were less affected by salt stress after receiving an application of Se in the form of sodium selenite (0.5–1 kg/ha). Improvements in qualitative and physiological markers were also noted by the authors [123]. Se was shown to reduce the negative effects of salt stress in a separate experiment using sunflower plants treated with Na₂SeO₄ (5 mg/kg). Treatment with Se resulted in increased glutathione peroxidase activity and a decrease in MDA levels in plant tissues [124]. Numerous studies have shown similar results, indicating that the MDA content in various plant species under different circumstances might be reduced by Se [125–127]. Recently, the use of Se was found to boost antioxidant enzyme activities in grapes, leading to a reduction in salt stress [128]. According to their research, foliar applications of Se treatment (Na₂OSe₄: 5–10 mg/L) boosted CAT activity in both groups of plants (0 or 75 mM NaCl), and leaf ascorbate peroxidase activity also increased.

5.1.2. Se and Heavy Metals

Plants are capable of absorbing nutrients from the soil for use in their vital metabolic activities. Meanwhile, because there is no particular preference for this mechanism, plants face the possibility of absorbing substances that are detrimental to their physiology. Cadmium (Cd) is one of the most hazardous heavy metals. Previous studies have shown that it has no identified benefits to the environment, and its introduction even in minute concentrations may have negative consequences [129–131]. One recent study showed that POD increased by 28%, 32%, and 27% in the presence of 0.0125, 0.025, and 0.05 mM foliar nano-Se, whereas MDA fell by 8%, 18%, and 4%, respectively. The authors claim that this improved rice's ability to endure Cd stress [132]. Another recent finding suggested that Se enhanced the antioxidant system in tall fescue plants to reduce the negative impacts of Cd and enhance Cd resistance. According to the researchers, Cd treatment (30 mg/L, as $CdSO_4 \cdot 8/3 H_2O$ increased MDA content by 63% and the relative electrolyte leakage value by approximately 166% higher than that of the control in tall fescue plants. However, Se supplementation $(0.1 \text{ mg/L}, \text{ as Na}_2\text{SeO}_3)$ reduced the MDA content by 52% and the relative EL value by approximately 29%. Additionally, Se treatment considerably increased the CAT activity by 40% and SOD activity by 30% compared with the Cd stress and control, respectively [133]. A study on the interactive effects of Cd and Se on the growth of rice plants found that at a constant Cd content of 4.16 mg/kg, dry grain weight rose considerably with increasing soil Se concentration [134].

In a different study, the researchers found that the root As content decreased by 7 to 55% in the Se treatments (Se-yeast and Se-malt); additionally, the stem As content decreased by 35 to 50%, and the leaf As content decreased by 0.1 to 33%. When Se malt was used, the results showed a similar pattern [135]. The result of Se and heavy metal interaction is thought to rely on Se speciation, relative dosage, application time, and application manner [136,137].

5.1.3. Se and Extreme Temperatures

Despite extensive research into how plants react to extreme temperatures, the mechanism underlying the tolerance capacity brought on by Se biofortification remains not fully understood. Under extremely cold conditions, Se (foliar, Na₂SeO₃, 5 mg/L) decreased the net photosynthetic rate and chlorophyll content and increased the MDA and hydrogen peroxide contents of strawberry seedling leaves. Se also increased the activities of catalase and superoxide dismutase [138].

Hasanuzzaman et al. looked at the protective function of Se (25μ M Na₂SeO₄) in reducing the harm that high temperature ($38 \,^{\circ}$ C) caused to rapeseed. They claim that heat-treated seedlings supplemented with Se experienced a considerable reduction in lipid peroxidation as well as an increase in chlorophyll content and antioxidant activity [139]. The development and physiological tolerance of lamb's lettuce cultivated under heat stress ($35/22 \,^{\circ}$ C; day/night) by biofortification with Se (foliar (50 mg Se/dm^3) and soil (Na₂SeO₄)

improved plant growth, reduced oxidative stress due to increased guaiacol peroxidase and catalase activity, and increased levels of GSH, and showed no change in the concentration of phenolic compounds [140].

5.1.4. Se and Photosynthesis

The impact of Se on plant leaf anatomy has been explored, with results varying based on soil Se concentration, plant species, and growth stage. Se can impact various physio-biochemical processes, increasing photosynthesis efficiency (Fv/Fm) in chlorophylls and activating the antioxidant system, and improving photosynthesis in stressed plants [141,142]. Wheat Fv/Fm was significantly reduced under salinity stress, but Se and Se + Si application reduced the negative effects on photosynthesis by decreasing the production of ROS that inhibit photosynthetic pigments [142]. In a study on Se's impact on tomato plant photosynthetic attributes such as leaf transpiration and CO₂ assimilation rate compared to the control plant [141]. Se application was also found to increase photosynthetic attributes in sorghum, likely due to its ability to reduce ROS production, repair damaged chloroplasts, and stimulate the production of other vital metabolites [120,143]. A recent study also revealed that both forms of Se (1 μ M Na₂SeO₃ and Na₂SeO₄) led to an increase in mesophyll intercellular spaces and thicker leaves [141].

Plant	Se Forms and Dosage	Se Effects	Reference
	Na2SeO4 20 and 40 mg/L Foliar Spray	Increased plant development due to higher salt tolerance during the reproductive stage by reducing oxidative damage and enhancing the activity of antioxidant enzymes.	[122]
	Na2SeO4 40 mg/L Foliar Spray	Increased fodder yield by 15%	[144]
Maize	Na ₂ SeO ₄ 0.8–1.0 g/L Foliar Spray	At the jointing stage, fresh ear yield went up by 2.3%; at the large bell stage, it went up by 2%.	[145]
	Na2SeO3 1, 5 and 25 μM Addition to nutrient solution	Enhanced salt resistance via changes in photosynthetic capacity, antioxidant activity, and Na ⁺ homeostasis	[146]
	Na ₂ SeO ₃ 5–15 μM Addition to nutrient solution.	Improved the activity of antioxidant system components	[147]
	Na2SeO4 0.4 mg Na2SeO4/kg soil Direct soil application	Height and weight of the plant increased	
Wheat	Na ₂ SeO ₄ 5 μM Direct addition to soil	In normal and NaCl-stressed seedlings, Se increased proline and sugar build-up and supplied additional osmolarity to preserve relative water content and safeguard photosynthesis.	[148]
	Na ₂ SeO ₄ 10 mL/pot Foliar spray	Enhanced antioxidant enzyme activity; improved plant growth, photosynthetic capacity, relative water content, and chlorophyll content	[149]
	Na2SeO3 25µM Addition to nutrient solution.	Increased phenolic chemicals and decreased arsenic accumulation	
Rice	Na_2SeO_4 10 μM Addition to nutrient solution.	Increased plant growth and biomass, and increased protein content. The activities of MDA, H_2O_2 , APX, CAT, and SOD reduced in the shoots.	
	Na ₂ SeO ₃ 0.8 and 1.0 mg/L	As-induced toxicity significantly decreased germination by 70%, and Se supplementation by seed priming increased germination by 9% and root, shoot, and seedling biomass accumulation by 1.3, 1.6, and 1.4 folds, respectively.	[152]

Table 1. Se biofortification effects in plants.

Plant	Se Forms and Dosage	Se Effects	Reference
Tomato	Na ₂ SeO ₃ or Na ₂ SeO ₄ 1 μM Addition to nutrient solution	Enhanced photosynthesis and increased root and shoot dry weight	
	Na ₂ SeO ₃ ·5H ₂ O 10 μM Direct soil application	Increased the levels of stomatal conductance, chlorophyll and carotene, transpiration rate and net photosynthesis rate	
	Se nanoparticles 10 mg/L Foliar Spray	Increased the yield by 21%	
P	Na2SeO3 5 μM Addition to nutrient solution	Increased root development, membrane stability index, chlorophyll concentration, and starch content in leaves	[154]
Pepper	Na ₂ SeO ₃ 3 and 7 μM Direct addition to soil	Plants cultivated in the medium containing 0.25 mM Cd had higher mean productivity, a greater capacity to withstand stress, and a higher yield stability index when the Se doses were added.	[155]
Onion	Na2SeO3 0.5 and 1 kg/ha Foliar spraying	Improvements in both qualitative and physiological markers. Maximum production at 1 kg/ha of foliar Se supplementation	[123]
Garlic	Na2SeO4 4, 8 and 16 mg/L Addition to nutrient solution.	Se improved salt tolerance and decreased oxidative damage by boosting the activity of antioxidant enzymes.	
Cucumber	Na2SeO3 2 g/L Addition to nutrient solution	Increased root and shoot biomass, as well as chlorophyll content	[157]
Mustard greens	Na2SeO4 4 μM/kg Addition to nutrient solution	Improved growth, increased chlorophyll and carotene content, net photosynthesis rate, stomatal conductance, and transpiration rate	[158]
Broad Beans	Na_2SeO_3 $1.5 \ \mu M$ Addition to nutrient solution	Decreased MDA content; and H ₂ O ₂ buildup, increased chlorophyll content shoot elongation and shoot fresh weight	[159]
Lemon balm	Na ₂ SeO ₃ .5H2O 0.2 μM Addition to nutrient solution	Enhanced growth	[160]
Strawberry	Na ₂ SeO ₄ nanoparticles, 10 and 20 mg/L Foliar Spray	Increased number of fruit plants—1 by 21.22 and 12.54%, and yield by 21 and 14%, respectively, in two growing seasons	[161]
Pomegranate	Na2SeO4 and Se-nanoparticles, 1 and 2 μM Foliar Spray	In two growing seasons, the number of fruits per tree grew by 1.35 and 1.28 times, and the yield grew by 1.17 and 1.16 times.	
Cowpea	Na2SeO4 5 and 10 μM Foliar application	Enhanced yield-related indicators, growth, and protein levels	[163]
Sunflower	Na2SeO4 5 mg/kg Direct soil application	Increased antioxidant enzyme activity	[124]
Tobacco	Na2SeO3 0.1 mg/L Addition to nutrient solution.	Se reduced the toxicity of the high As dosage (5 mg/L) and stimulated the development of the plant by increasing antioxidative stress resistance and decreasing MDA levels.	[164]

Table 1. Cont.

6. Se Biofortification in Livestock

Rapid livestock production increases the necessity for Se [165,166]. In animals, the liver, heart, and skeletal muscle are the first organs to become deficient in Se [114]. Inadequate levels of Se have been linked to a variety of illnesses in livestock, including nutritional myopathy and ill-thrifting [167,168] as well as white muscle disease [169], which is primarily unnoticeable in older animals [170]. White muscle disease is a severe degenerative condition marked by rigidity, exhaustion, limb trembling, and inflamed muscles [171]. It has been labeled the animal variant of Keshan disease [172]. Ruminants' absorption of Se is less efficient and complicated than that of non-ruminants since rumen microbial populations may convert Se to inaccessible forms [173], and only one-third of inorganic Se is absorbed. Some studies suggest that Se supplementation in animal feeds should be between 0.05 and 0.1 mg/kg dry matter (DM) to satisfy Se requirements [174,175]. Freshwater creatures such as crayfish, crabs, and carp [176] may deposit Se in their tissues, even in rare Se geographic locations. However, it is uncertain whether eating seafood has a role in their Se buildup.

Se Biofortification Effects in Livestock

One of the most critical factors recognized in the pathological course of many illnesses and cancers is oxidative stress and the unregulated creation of reactive oxygen species [177]. Total superoxide dismutase (T-SOD), GSH-Px, and catalase are intricate defense and repair mechanisms that may protect animals from oxidative damage [178]. Livestock research has shown that Se may reduce heavy metal toxicity and alter the degree of heavy metal exposure and illness (Table 2). For instance, in a previous study, Se reduced Cd-induced hepatotoxicity [179]. The researchers discovered that in cocks, a 10 mg/kg Se(IV) diet decreased Cd buildup and boosted antioxidant resistance in hepatic tissue, as well as Cdinduced morphological alterations and oxidative stress. According to the researchers, the results can be explained by Se's crucial role in avoiding lipid peroxidation and preserving the structural and functional integrity of tissues. Selenium supplementation protected the chicken brain against chromium damage by blocking adverse effects [180]. They further found that Se supplementation reduced MDA activity and that the supplementation at 5 mg/kg BW greatly boosted SOD activity. Furthermore, Se supplementation reduced MDA activity because Se inhibits hydroxyl radical production and maintains tissue function. Additionally, Se is acknowledged to be a crucial component for reproductive function in poultry. Dietary Se supplementation at varying concentrations (0.10-1.00 mg/kg) and sources enhanced laying performance and egg quality [181]. Se administration using either organic (Se-enriched yeast) or inorganic (sodium selenite) forms of Se at levels of 0.3 and 0.15 mg/kg against the H9N2 virus dramatically reduced viral shedding within the chicken, with the organic form proving more efficient [182]. However, according to an experiment on hens augmented with Se orally at doses of 5, 10, and 15 mg/kg for 15, 30, and 45 days, high intakes of Se caused a significant decrease in the levels of the cytokines IFN- γ and IL-2 in both serum and the thymus, as well as a low-to-moderate incidence of pathological changes in the thymus tissue, which suggests a decrease in protection and an upsurge in oxidative damage [183].

A study by Li et al. [176] revealed that nano-Se may counteract the oxidative stress and inflammation in chickens caused by di-(2-ethylhexyl)phthalate (DEHP), a plasticizer extensively used in the food sector. According to their study, the SOD, T-AOC, GSH-PX, and CAT activities of the DEHP group were considerably reduced (p < 0.05) compared with those of the control group, whereas the SOD, T-AOC, and CAT activities of the nano Se group were significantly increased (p < 0.05).

A previous study in pigs found that 6 mM Se Met suppressed PCV2, a postweaning multisystemic wasting syndrome, and that 2 or 4 mM Se Met prevented the increase in PCV2 replication induced by oxidative stress [184]. Some studies have speculated that the underlying mechanism of Se Met inhibition of PCV2 replication is mediated by increased activity of GSH-Px, which shields the cell from free-radical oxidant harm [185]. The effects of dietary Se on parainfluenza virus infection in lambs have also been studied, focusing on the innate and adaptive immune responses to virus infection. When the parainfluenza virus was injected into lambs that had been fed Se, they exhibited increased immunological activity [186]. A recent study found that health and reproduction parameters improved for cows fed with organic Se forms [187]. Sun and coworkers [188] found that Se-yeast treatment boosts antioxidant capacity in dairy cows. They showed an increase in serum

glutathione peroxidase activity (p < 0.05) and total antioxidant capacity (p = 0.08), as well as a reduction in MDA content (p < 0.05).

Table 2. Se biofortification effects in livestock.

Animal	Se Form and Dosage	Se Effects	Reference
Cow	Se yeast supplement	Enhanced antioxidant levels and immunological responses following calving	
	Se-enriched alfalfa hay	Supplemental selenium increased immunization responses against Escherichia coli during the weaning transition phase and subsequent growth and survival in the feedlot.	
	DL-selenomethionine 2–16 µmol/L	Significant inhibitive effect on Porcine circovirus type 2 replication	
Pig	SeMet 2–6 µM	Inhibited porcine circovirus type 2 replication and its related oxidative stress	
	Se yeast diet	Piglets given selenium yeast showed greater digestibility of DM, crude protein, and crude fat; which impacted the production of inflammatory cytokines, and decreased the quantity of Escherichia coli in feces.	
	SeMet	Increased immune function and selenoprotein expression, and reduced the inflammation generated by lipopolysaccharides.	
	0.3 mg/kg Se yeast 0.3 mg/kg of organic Se from <i>Stenotrophomonas maltophilia</i> (bacterial organic Se, ADS18).	Bacterial selenoprotein or Se-yeast improved the performance index, egg quality features, egg yolk and tissue of Se concentrations and intestinal villus.	
Chicken	Se Enriched Yeast Na ₂ SeO ₃ (High—0.30 mg/kg of feed; Low—0.15 mg/kg of feed)	Virus shedding from the cloaca was substantially reduced in all selenium-supplemented groups compared with non-supplemented control groups.	[182]
	sodium selenite 10 or 20 µg	Se injection enhanced immune and antioxidant responses	[194]
	Probiotics as (P, 0.11 mg Se/kg) Na ₂ SeO ₃ (SS, 0.41 mg Se/kg) and (SP, 0.41 mg Se/kg)	In groups supplemented with selenium, oocyst shedding and cecal lesion scores were reduced.	
Sheep	Se yeast supplementation >4.9 mg Se/week	Supplementation with Se-yeast enhanced the Se status of sheep and the expression of genes involved in innate immunity in whole blood neutrophils.	
	Se yeast 0.5–1.0 mg/kg	Drip loss of muscle decreased significantly with an increase in dietary selenium yeast Supplementation.	[197]
D 11 ''	Se yeast 0.3 mg Se/kg diet	Positive effect on growth performance of rabbits. Se increased daily gain and the final body weight. Supplementation with Se increased muscle Se content to 559% of the control level.	[198]
Rabbit	Sodium selenate solution 10% of Se-fortified olive leaves (2.10 mg/kg)	Meat exhibited better oxidative status and a 5-fold higher Se content compared to that of the other treatments.	[199]

7. Se Biofortification and Humans

Se deficiency in humans can be linked to lower levels in plant and livestock produce. Deficiency in humans can be associated with numerous ailments that denote a significant impact on the socioeconomic development of individuals. However, due to changes in laboratory methodologies throughout the world and the lack of specific biofortification values and guidelines for application and intake, there is a significant variability globally (Table 3). Some regulatory agencies recommend daily Se consumption of 30–85 μ g/d for males and 30–70 μ g/d for females to meet dietary requirements [200–202].

On average, the concentration of Se in cereals produced in Europe reportedly varies from 0.02 to 0.05 mg/kg DM, whereas in North America, it is 0.2 to 0.5 mg/kg (Table 3) [45,203]. Previous studies [204,205] demonstrated that the highest mean and median amounts of Se in rice grown in the United States were 176 and 180 ng/g, respectively. In contrast, the lowest mean and median concentrations of Se in Egyptian rice were 9.0 and

6.0 ng/g, respectively. The researchers also reported that the average value of rice in India was 152 ng/g; however, 5% of Chinese rice had an Se content of more than 200 ng/g.

Se consumption levels equivalent to the required dietary amount have been reported in some countries. For instance, Belgium and France [41,206] have met the required dietary quota. The present Se intake in Finland is also reported to be in accordance with the Nordic, European Union, and United States standards [207–209]. However, studies from several parts of the world show that the required dietary amount is not met. Places such as Italy and Slovenia showed intakes that were below the recommended dietary limit [210–212]. Previous studies in Kuwait, Saudi Arabia, and Turkey that looked at Se concentration in blood plasma and serum recorded that breast milk and umbilical cord blood were low [213–215]. In Turkey, the Se content in breast milk was below the international standard range (18.5 μ g/L) during the breastfeeding period [216]. In Britain, a longitudinal study of British individuals in good health revealed poor Se intake [217]. Se intake in Poles compared to that of Spaniards was reported to be four times lower [218–221].

Table 3. Se intake status in different countries.

Country	Se Intake	References
Russia	35.5 μg	[222]
Brazil	84.3–105.9 μg	[223-225]
United States of America	60–220 μg	[209,224,226–228]
Turkey	20–138 µg	[215,229–239]
Slovakia	27–43 μg	[240]
Saudi Arabia	34–121.65 μg	[241,242]
Venezuela	200–350 μg	[21,243]
Czech Republic	10–25 μg	[243]
Canada	98–224 μg	[225,228]
England	12–43 μg	[228]
Belgium	28–61 µg	[224]
Germany	35–47 μg	[224,225]
Mexico	61–73 μg	[224,228]
Venezuela	200–350 µg	[224,228]
Australia	57–87 μg	[209,228]
Japan	104–127 µg	[228]
Greece	110 μg	[228]
China	3–6690 µg	[22,224,243]
Poland	30–40 μg	[244]
Finland	70–80 µg	[71]
Spain	44–50 μg	[117,245]
Austria	48 μg	[21,117]
Slovenia	87 μg	[246]
Slovakia	27–43 µg	[240]
Jordan	59.26 μg	[247]
Greenland	193–5885 µg	[248]

7.1. Se in Humans

7.1.1. Se Intake

Mehdi et al. [116] found that between inorganic and organic Se, the former is more harmful than the latter. Additionally, Vinceti et al. [42] discovered that inorganic Se was 40 times more dangerous than organic Se. However, several studies on humans have revealed that doses of up to 800 μ g Se/day provided as Se-yeast did not result in any harmful effects [105]. Several variables influence Se intake, including nutritional habits and geographical location, as well as food imports and sources of food. Reduced Se intake and levels in the United Kingdom and other northern European nations are thought to have originated since the mid-20th century due to changing trade, which resulted in lower wheat imports from the US and Canada [172]. In New Zealand, the process that was used to remove arsenic from superphosphate fertilizers also removed Se, decreasing the Se status in plants and livestock [44].

Se deficiency is linked to a variety of cancerous diseases [249], renal impairment [250], type 1 diabetes [251], epilepsy, and cardiovascular diseases [93]. Keshan illness [252] was the first human disease associated with Se deficit. Kashin–Beck disease, a bone and cartilage disease described in China, Tibet, North Korea, and Siberia, was the second [249]. Thyroid hormone metabolism problems and selenoprotein N-related myopathy, both of which are congenital muscle disorders, have also been linked to decreased selenoprotein expression [253]. Other variables besides Se insufficiency may be the main cause of the conditions of various diseases and increased oxidative stress. However, a strong Se status along with a sufficient intake of other antioxidative nutrients may assist cells and tissues better withstand the detrimental oxidative stress caused [44]. For example, hyperglycemia [231] or the immune system's response to infection [254].

7.1.2. Health Benefits of Se

Se has been shown to aid muscular function by improving endurance and recuperation, as well as slowing the aging process [116]. Cengiz et al. [255] discovered a significant link between low Se levels in expectant women's blood and the occurrence of neural tube abnormalities, particularly anencephaly and rachis. Hence, a reduction in reproduction in females is usually related to Se-insufficient body saturation [256,257]. However, the role of Se in this cycle is unclear.

In 1960, the US states with higher intakes of Se reported lower rates of cancer deaths than states with lower intakes [258,259]. The socioeconomic impact of cancers is significantly increasing. In 2010, the global economic cost of cancer was estimated to be over \$1.2 trillion [238], and it was €199 billion in Europe in 2018 [260]. The ingestion of Se-supplemented foods has been researched as a potential remedy. For instance, the consumption of Se-fortified garlic and broccoli produces Se-methyl–Se-cysteine, which is converted to methyl selenol, a powerful cancer-fighting compound [261,262]. According to a study on 18 individuals given 200 μ g Se-enriched broccoli daily for three days, Se intake led to noticeably greater levels of both Th1 and Th2 cytokines released by peripheral blood mononuclear cells. The researchers found that supplementation raised plasma Se levels [263]. The contribution of Se to tumor cell invasion, cell proliferation, and apoptosis has been investigated [264,265]. Se contained in a fraction of selenoproteins, has been found to promote anti-carcinogenic factors and have anti-proliferative and anti-inflammatory properties [266,267]. It has also been shown to lessen the severe side effects of several chemotherapeutic drugs while maintaining their anticancer properties [268,269].

In the treatment of viral infections, 200 μ g of selenium per day reduced HIV patients' hospital admissions and infection-related admissions [270]. Similar research showed that higher Se content in serum was associated with lower viral load, even after adjusting for antiretroviral therapy regimen and adherence [271]. However, some researchers have disputed the data analysis approach [272]. In patients who received a live attenuated poliovirus vaccination, treatment using 100 μ g Se/d increased the number of total T cells and Th cells and improved virus clearance [273].

Se is reportedly used in a number of health materials. It suppresses bone cancer in a localized location without damaging healthy tissue in the surrounding area, indicating tremendous potential for novel bone cancer therapy options. Depending on the kind of Se used, the frequent technique for treating bone cancer is incorporating it into ceramic substrates [274]. Moreover, Se has been used as an anticancer material in breast, lung [275], and prostate [276] applications. This indicates Se antioxidant capabilities, immunological protection, carcinogen detoxification, cell proliferation modulation, and suppression of cancer cell invasion and angiogenesis, among other things [277]. Selenium has also been shown to lessen the severe side effects of several chemotherapeutic drugs while maintaining their anticancer properties [268,269].

8. Se Biofortification Challenges

8.1. Influence of Soil Characeristics

Direct soil application has been shown to enhance the biofortification of crops with Se. However, studies highlight issues with Se biofortification through soil, owing to the lower uptake by plants and the probable economic losses associated with this strategy [278]. Previous research, for example, found that improving Se status in grain to 100 μ g Se/kg requires almost six times more fertilizer for soil treatment than foliar spray [279]. Plants absorb most of their nutrients in the rhizosphere, and the conditions there can affect how bioavailable Se is to plants [280,281].

Ions present in soils may hinder the uptake of selenium by plants (Figure 3). The anions sulfate and phosphate may compete with Se for absorption by plants [282]. Se remains bound in phosphate precipitates when phosphate fertilizers are applied to the soil, rendering it unavailable for uptake [44]. Selenium also shares similar characteristics to S [283,284], and it is documented that adding S to the soil inhibits plant Se absorption [285,286], as they share the same metabolic pathway during translocation [287].

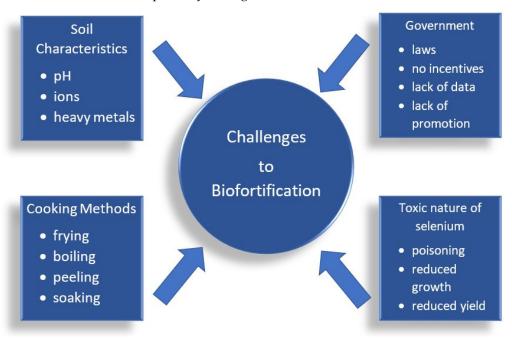


Figure 3. Challenges to biofortification.

Additionally, metal and metalloid bioavailability and speciation in soils are affected by soil clay [288,289]. Se is poorly bioavailable in clayey soils as a result of its attraction to clay minerals [290]. According to studies, Se can adsorb on positively charged sites of Al-octahedral sheets in clay minerals such as kaolinite and changes significantly with soil pH [291].

Soil pH also affects the adsorption of Se [282]. Selenite is insoluble in acidic and neutral soils while Se(IV) is more readily absorbed by plants in neutral and alkaline soils [292]. This has been demonstrated in previous studies in which Se-Se(VI)-enriched soils were noted to absorb Se 10 times more readily than Se-Se(IV) [293]. Additionally, according to thermodynamic calculations, Se(IV) should predominate in mineral soils with pH values between acidic and neutral (7.5 < pe + pH < 15), and selenate in alkaline and well-oxidized soils (pe + pH > 15) [294–296].

8.2. Food Processing Methods

Consuming Se-fortified foods is critical for improving Se status in humans. The major goal of Se biofortification of plants and animals is to raise the Se status in humans. The concentration of Se in a living organism is heavily influenced by Se consumption [20,297].

However, Se-fortified products such as grains, vegetables, and meat are usually ingested after cooking practices such as roasting, frying, and boiling. Se content in foods is reported to be affected by cooking and preservation methods. Dong et al. [298] found that the overall Se content was reduced by about 43.3% after boiling (10 min, 100 degrees Celsius), 38.5% was lost to the water used, and 31.7% was lost after frying (10 min, 180 degrees Celsius). They also observed that Se in tubers was reduced by 53.4–69.9% when peeled. Some researchers observed that among the five typical processing techniques, frying recorded the highest Se loss (64%) in garlic, and Se volatilization caused by high-temperature heating was the primary factor [299]. Correspondingly, other food processing techniques, such as soaking, have recorded a loss in the concentration of Se content by 2.6% to 7.2% [300]. Boiling, steaming, and frying were found to negatively affect the total Se concentration in wheat (5.6–13.6%) [301]. A recent publication found that the latter had a more significant decreasing effect on the total Se concentration in *Pleurotus eryngii* fruit bodies than in boiled fruit bodies [80]. Additionally, food preservation methods have been observed to cause Se to be embedded in the food medium and not freed in the intestines [302]; as a result, absorption and usage of the element are affected [303]. Matos and coworkers found that while the Se content in blue shark rose after steaming and grilling, its bioaccessibility decreased considerably (p < 0.05) [304]. Zhou et al. [304] discovered that 35.3% of Se in boiled tubers was not bioavailable, but the figure for fried samples rose to 76.6% following oral and gastrointestinal digestion.

8.3. Toxic Nature of Se

Despite the importance of Se, its toxic nature past a certain level is a factor that hinders its biofortification. The transition between biofortification and toxicity of Se is narrow. Considering the very small range between nutritional quantities that are deadly and inadequate, it is easy for selenium supplementation in animals to lead to toxic or even fatal doses. Treatments for Se biofortification in animals can lead to poisoning as a result of either unintentional or deliberate dosages [305]. In plants, while some can accumulate high concentrations of the element and are termed hyperaccumulators [292], most plants that are consumed, such as wheat, rice, maize and barley, which cannot accumulate such higher amounts and are termed non-accumulators [72]. Selenium toxicity has been reported to cause a negative impact on plant physiology, growth and development [50,306–313], nutrient content [50,312,314,315], and yield [306,316,317].

Selenium (SeO₃²⁻; 50 or 100 μ M) produced secondary nitrooxidative stress, lowered root development and yield, lowered cell viability, affected cell wall structure by altering pectin and callose, and lowered stomatal density in a study using thale cress [306]. Sefortified (SeO₄²⁻ 80 μ M and SeO₃²⁻ 20 μ M) cucumbers showed reduced biomass, shoot growth, root growth, and leaf area. Additionally, it worsened nutritional content, decreased the formation of photosynthetic pigments, increased lipid peroxidation, and decreased chlorophyll fluorescence [50].

8.4. Government Support

Considering that Se deposits are rare, finite, and potentially susceptible to depletion by improper or inefficient use, they must be conserved [279]. Support from government in the type of funds for research and laws will be essential to ensure the proper utilization of the resource. Most importantly, as the greatest advantage of Se enrichment is to improve public wellbeing by lowering illness costs [318], policymakers must ensure that scientists, food producers, and health providers have enough data on the population and environment to aid them in better assessing the situation and providing the necessary and required duties. Education programs about Se biofortification, Se biofortified products and why they are needed will also go a long way to help the social acceptance and patronage of Se-biofortified produce. A higher patronage will likely cause a more affordable price for the general public. According to Bouis et al., [319], if provided biofortified produce is comparatively less expensive than the competition and has similar quality, individuals from developed and

underdeveloped countries will patronize them. However, most developing economies do not have the resources needed to ensure the needed measures for biofortification and put to action.

9. Conclusions

Inadequate Se status in plants and livestock appears to be the most prevalent cause of Se deficiency. Se-deficient crops may stem from either a low Se concentration in the soil, a limited availability of soil Se for absorption by plant roots, or both. In food animals, low levels of Se in their diets seem to be the primary cause. Research trends suggest Se supplementation's efficacy in maintaining homeostasis of several metabolic processes. Researchers are utilizing various techniques to increase Se content in edible sections of crops and livestock to address both the issue of low Se intake and its impacts. However, biofortification of agricultural products should be thoroughly researched and assessed in view of Se resource planning and conservation. Additionally, further study is required to confirm the safety and appropriateness of present Se levels to define optimal levels. Modern biological and analytical technologies must also be progressively developed and utilized to assess the imprints and pathways of selenium in a wide variety of foods and geographic locations to find other Se-deficient regions and combat the existing global Se shortfall. Additionally, international support systems must be recognized for developing economies.

Author Contributions: Conceptualization, O.P.D., X.Y. and R.Z.; writing—original draft preparation, O.P.D.; writing—review and editing, O.P.D., B.A.-B., Z.Z., J.S., Z.W., X.Y. and R.Z.; visualization, O.P.D. and B.A.-B.; supervision, Z.Z., J.S., Z.W., X.Y. and R.Z.; funding acquisition, X.Y. and R.Z. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Special Fund for Functional Agricultural Development of National Agricultural Parks (No. NJGJNCY-FAST01) and the National Natural Science Foundation of China (No. 41976220).

Institutional Review Board Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: We highly acknowledge the support of the Alliance of International Science Organizations (ANSO) and the Nanjing Institute for Functional Agriculture Science and Technology (iFAST). We are thankful to the National Natural Science Foundation of China and the Functional Agricultural Development of National Agricultural Parks.

Conflicts of Interest: The authors declare that they have no conflict of interest.

References

- 1. Oldfield, J. Se World Atlas; Se-Tellurium Development Association (STDA): Grimbergen, Belgium, 1999.
- Garside, M. Production Volume of Selenium Worldwide in 2020, by Country. 2022. Available online: https://www.statista. com/statistics/1312522/selenium-production-volume-worldwide-by-country/#:~{}:text=The%20country%20with%20the%20 largest,3.33%20thousand%20metric%20tons%20worldwide. (accessed on 13 December 2022).
- Basu, A.; Schilling, K.; Brown, S.T.; Johnson, T.M.; Christensen, J.N.; Hartmann, M.; Reimus, P.W.; Heikoop, J.M.; Woldegabriel, G.; DePaolo, D.J. Se Isotopes as Groundwater Redox Indicators: Detecting Natural Attenuation of Se at an in Situ Recovery U Mine. *Environ. Sci. Technol.* 2016, *50*, 10833–10842. [CrossRef]
- Stroud, J.L.; McGrath, S.P.; Zhao, F.J. Selenium speciation in soil extracts using LC-ICP-MS. Int. J. Environ. Anal. Chem. 2012, 92, 222–236. [CrossRef]
- 5. Pettine, M.; Gennari, F.; Campanella, L. The reaction of selenium (IV) with ascorbic acid: Its relevance in aqueous and soil systems. *Chemosphere* **2013**, *90*, 245–250. [CrossRef] [PubMed]
- 6. Reich, H.J.; Hondal, R.J. Why Nature Chose Selenium. ACS Chem. Biol. 2016, 11, 821–841. [CrossRef] [PubMed]
- Sharma, V.K.; McDonald, T.J.; Sohn, M.; Anquandah, G.A.; Pettine, M.; Zboril, R. Biogeochemistry of selenium. A review. *Environ. Chem. Lett.* 2015, 13, 49–58. [CrossRef]
- 8. Oldfield, J.E.; Muth, O.H.; Schubert, J.R. Selenium and vit. E as related to growth and white muscle disease in lambs. *Proc. Soc. Exp. Biol. Med.* **1960**, *103*, 799–800. [CrossRef]
- 9. Pilarczyk, B.; Tomza-Marciniak, A.; Pilarczyk, R.; Kuba, J.; Hendzel, D.; Udała, J.; Tarasewicz, Z. Eggs as a source of selenium in the human diet. *J. Food Compos. Anal.* 2019, *78*, 19–23. [CrossRef]

- Schwarz, K.; Foltz, C.M. Selenium as an Integral Part of Factor 3 against Dietary Necrotic Liver Degeneration. J. Am. Chem. Soc. 1957, 79, 3292–3293. [CrossRef]
- Collery, P. Strategies for the development of selenium-based anticancer drugs. J. Trace Elements Med. Biol. 2018, 50, 498–507. [CrossRef]
- 12. Hesketh, J. Nutrigenomics and selenium: Gene expression patterns, physiological targets, and genetics. *Annu. Rev. Nutr.* 2008, 28, 157–177. [CrossRef]
- Saito, Y. Selenium Transport Mechanism via Selenoprotein P—Its Physiological Role and Related Diseases. Front. Nutr. 2021, 8, 685517. [CrossRef] [PubMed]
- Guo, Y.; Guo, X.; Yan, S.; Zhang, B.; Shi, B. Mechanism Underlying the Protective Effect of Selenium on NO-Induced Oxidative Damage in Bovine Mammary Epithelial Cells. *Biol. Trace Element Res.* 2019, 191, 104–114. [CrossRef] [PubMed]
- 15. Hall, J.A.; Bobe, G.; Vorachek, W.R.; Hugejiletu; Gorman, M.E.; Mosher, W.D.; Pirelli, G.J. Effects of Feeding Selenium-Enriched Alfalfa Hay on Immunity and Health of Weaned Beef Calves. *Biol. Trace Element Res.* **2013**, *156*, 96–110. [CrossRef]
- 16. Hartikainen, H. Biogeochemistry of selenium and its impact on food chain quality and human health. *J. Trace Elements Med. Biol.* **2005**, *18*, 309–318. [CrossRef] [PubMed]
- 17. Rayman, M.P. Selenium intake, status, and health: A complex relationship. Hormones 2019, 19, 9–14. [CrossRef] [PubMed]
- 18. Minich, W.B. Selenium Metabolism and Biosynthesis of Selenoproteins in the Human Body. *Biochemistry* **2022**, *87*, S168–S177. [CrossRef]
- 19. Martinez, S.S.; Huang, Y.; Acuna, L.; Laverde, E.; Trujillo, D.; Barbieri, M.A.; Baum, M.K. Role of selenium in viral infections with a major focus on SARS-CoV-2. *Int. J. Mol. Sci.* 2022, *23*, 280. [CrossRef]
- 20. Adadi, P.; Barakova, N.V.; Muravyov, K.Y.; Krivoshapkina, E.F. Designing selenium functional foods and beverages: A review. *Food Res. Int.* **2019**, *120*, 708–725. [CrossRef]
- 21. Combs, G.F., Jr. Selenium in global food systems. Br. J. Nutr. 2001, 85, 517–547. [CrossRef]
- 22. Zhang, B.; Wei, Y.; Yan, S.; Shi, H.; Nie, Y.; Zou, G.; Zhang, X.; Luo, L. Characterization of selenium accumulation of different rice genotypes in Chinese natural seleniferous soil. *Plant Soil Environ.* **2019**, *65*, 15–20. [CrossRef]
- Pilon-Smits, E.A.; LeDuc, D.L. Phytoremediation of selenium using transgenic plants. *Curr. Opin. Biotechnol.* 2009, 20, 207–212. [CrossRef]
- 24. Pilon-Smits, E.A.; Quinn, C.F. Selenium metabolism in plants. In *Cell Biology of Metals and Nutrients*; Springer: Berlin/Heidelberg, Germany, 2010; pp. 225–241.
- 25. Andrews, E.; Hartley, W.; Grant, A. Selenium-responsive diseases of animals in New Zealand. N. Z. Vet.- J. 1968, 16, 3–17. [CrossRef]
- Abdel-Moneim, A.-M.E.; Shehata, A.M.; Mohamed, N.G.; Elbaz, A.M.; Ibrahim, N.S. Synergistic effect of Spirulina platensis and selenium nanoparticles on growth performance, serum metabolites, immune responses, and antioxidant capacity of heat-stressed broiler chickens. *Biol. Trace Element Res.* 2021, 200, 768–779. [CrossRef] [PubMed]
- 27. Libera, K.; Konieczny, K.; Witkowska, K.; Żurek, K.; Szumacher-Strabel, M.; Cieslak, A.; Smulski, S. The Association between Selected Dietary Minerals and Mastitis in Dairy Cows—A Review. *Animals* **2021**, *11*, 2330. [CrossRef] [PubMed]
- Rashnoo, M.; Rahmati, Z.; Azarfar, A.; Fadayifar, A. The effects of maternal supplementation of selenium and iodine via slow-release blouses in late pregnancy on milk production of goats and performance of their kids. *Ital. J. Anim. Sci.* 2020, 19, 502–513. [CrossRef]
- 29. Skrajnowska, D.; Jagielska, A.; Ruszczyńska, A.; Idkowiak, J.; Bobrowska-Korczak, B. Effect of Copper and Selenium Supplementation on the Level of Elements in Rats' Femurs under Neoplastic Conditions. *Nutrients* **2022**, *14*, 1285. [CrossRef]
- Safonov, V. Comparison of LPO-AOS Indices and Biochemical Composition of Animal Blood in Biogeochemical Provinces with Different Levels of Selenium. *Biol. Trace Element Res.* 2021, 200, 2055–2061. [CrossRef]
- Hemmati, M.; Delkhosh, B.; Rad, A.H.S.; Mohammadi, G.N. Effect of the Application of Foliar Selenium on Canola Cultivars as Influenced by Different Irrigation Regimes. J. Agric. Sci. 2019, 25, 309–318. [CrossRef]
- Kleiber, T.; Krzesiński, W.; Przygocka-Cyna, K.; Spiżewski, T. Alleviation Effect of Selenium on Manganese Stress of Plants. *Ecol. Chem. Eng. S* 2018, 25, 143–152. [CrossRef]
- 33. Asante-Badu, B.; Kgorutla, L.; Li, S.; Danso, P.; Xue, Z.; Qiang, G. Phytoremediation of organic and inorganic compounds in a natural and an agricultural environment: A review. *Appl. Ecol. Environ. Res.* **2020**, *18*, 6875–6904. [CrossRef]
- 34. Fairweather-Tait, S.J.; Bao, Y.; Broadley, M.; Collings, R.; Ford, D.; Hesketh, J.E.; Hurst, R. Selenium in Human Health and Disease. *Antioxid. Redox Signal.* 2011, 14, 1337–1383. [CrossRef] [PubMed]
- 35. Rodriguez, M.M.; Rivero, V.C.; Ballesta, R.J.; Rivero, M.V.C. Selenium Distribution in Topsoils and Plants of a Semi-arid Mediterranean Environment. *Environ. Geochem. Health* **2005**, *27*, 513–519. [CrossRef] [PubMed]
- 36. Ye, W.; Zhu, R.; Yuan, L.; Zhang, W.; Zang, H.; Jiao, Y.; Yin, X. The influence of sea animals on selenium distribution in tundra soils and lake sediments in maritime Antarctica. *Chemosphere* **2021**, *291*, 132748. [CrossRef]
- 37. Feinberg, A.; Stenke, A.; Peter, T.; Hinckley, E.L.S.; Driscoll, C.T.; Winkel, L.H. Reductions in the deposition of sulfur and selenium to agricultural soils pose risk of future nutrient deficiencies. *Commun. Earth Environ.* **2021**, *2*, 101. [CrossRef]
- Ye, W.; Yuan, L.; Zhu, R.; Yin, X.; Bañuelos, G. Selenium volatilization from tundra soils in maritime Antarctica. *Environ. Int.* 2021, 146, 106189. [CrossRef] [PubMed]
- 39. Hoet, P. Sélénium et ses Composés; Elsevier Masson SAS: Paris, France, 2013.

- 40. Dhillon, K.S.; Dhillon, S.K.; Singh, B. Genesis of seleniferous soils and associated animal and human health problems. *Adv. Agron.* 2018, 154, 1–80. [CrossRef]
- 41. Emmanuelle, B.; Virginie, M.; Fabienne, S.; Isabelle, I.; Martine, P.-G.; Bernard, L.; Sylvie, R. Selenium exposure in subjects living in areas with high selenium concentrated drinking water: Results of a French integrated exposure assessment survey. *Environ. Int.* **2012**, *40*, 155–161. [CrossRef]
- 42. Vinceti, M.; Crespi, C.M.; Bonvicini, F.; Malagoli, C.; Ferrante, M.; Marmiroli, S.; Stranges, S. The need for a reassessment of the safe upper limit of selenium in drinking water. *Sci. Total. Environ.* **2013**, *443*, 633–642. [CrossRef]
- Santos, S.; Ungureanu, G.; Boaventura, R.; Botelho, C. Selenium contaminated waters: An overview of analytical methods, treatment options and recent advances in sorption methods. *Sci. Total. Environ.* 2015, 521–522, 246–260. [CrossRef]
- 44. Haug, A.; Graham, R.D.; Christophersen, O.A.; Lyons, G.H. How to use the world's scarce selenium resources efficiently to increase the selenium concentration in food. *Microb. Ecol. Health Dis.* **2007**, *19*, 209–228.
- 45. Natasha; Shahid, M.; Niazi, N.K.; Khalid, S.; Murtaza, B.; Bibi, I.; Rashid, M.I. A critical review of selenium biogeochemical behavior in soil-plant system with an inference to human health. *Environ. Pollut.* **2018**, 234, 915–934. [CrossRef]
- 46. Dungan, R.S.; Frankenberger, W.T. Microbial Transformations of Selenium and the Bioremediation of Seleniferous Environments. *Bioremediation J.* **1999**, *3*, 171–188. [CrossRef]
- 47. Besser, J.M.; Canfield, T.J.; La Point, T.W. Bioaccumulation of organic and inorganic selenium in a laboratory food chain. *Environ. Toxicol. Chem. Int. J.* **1993**, *12*, 57–72. [CrossRef]
- 48. Elrashidi, M.A.; Adriano, D.C.; Lindsay, W.L. Solubility, speciation, and transformations of selenium in soils. *Selenium Agric. Environ.* **1989**, *23*, 51–63.
- Freeman, J.L.; Bañuelos, G.S. Selection of Salt and Boron Tolerant Selenium Hyperaccumulator *Stanleya pinnata* Genotypes and Characterization of Se Phytoremediation from Agricultural Drainage Sediments. *Environ. Sci. Technol.* 2011, 45, 9703–9710. [CrossRef]
- 50. Hawrylak-Nowak, B.; Matraszek, R.; Pogorzelec, M. The dual effects of two inorganic selenium forms on the growth, selected physiological parameters and macronutrients accumulation in cucumber plants. *Acta Physiol. Plant.* **2015**, *37*, 41. [CrossRef]
- Mikkelsen, R.L.; Page, A.L.; Bingham, F.T.; Jacobs, L.W. Factors Affecting Selenium Accumulation by Agricultural Crops. Selenium Agric. Environ. 2015, 23, 65–94. [CrossRef]
- 52. Fernandez-Martinez, A.; Charlet, L. Selenium environmental cycling and bioavailability: A structural chemist point of view. *Rev. Environ. Sci. Bio/Technol.* 2009, *8*, 81–110. [CrossRef]
- 53. Masscheleyn, P.H.; Delaune, R.D.; Patrick, W.H. Arsenic and Selenium Chemistry as Affected by Sediment Redox Potential and pH. *J. Environ. Qual.* **1991**, *20*, 522–527. [CrossRef]
- McNeal, J.M.; Balistrieri, L.S.; Jacobs, L.W. Geochemistry and Occurrence of Selenium: An Overview. *Selenium Agric. Environ*. 2015, 23, 1–13. [CrossRef]
- 55. Kikkert, J.; Berkelaar, E. Plant Uptake and Translocation of Inorganic and Organic Forms of Selenium. *Arch. Environ. Contam. Toxicol.* **2013**, *65*, 458–465. [CrossRef] [PubMed]
- 56. Dinh, Q.T.; Cui, Z.; Huang, J.; Tran, T.A.T.; Wang, D.; Yang, W.; Zhou, F.; Wang, M.; Yu, D.; Liang, D. Selenium distribution in the Chinese environment and its relationship with human health: A review. *Environ. Int.* **2018**, *112*, 294–309. [CrossRef] [PubMed]
- 57. Zhang, M.; Xing, G.; Tang, S.; Pang, Y.; Yi, Q.; Huang, Q.; Huang, X.; Huang, J.; Li, P.; Fu, H. Improving soil selenium availability as a strategy to promote selenium uptake by high-Se rice cultivar. *Environ. Exp. Bot.* **2019**, *163*, 45–54. [CrossRef]
- Kumar, A.; Prasad, K.S. Role of nano-selenium in health and environment. *J. Biotechnol.* 2020, 325, 152–163. [CrossRef] [PubMed]
 Kápolna, E.; Hillestrøm, P.R.; Laursen, K.H.; Husted, S.; Larsen, E.H. Effect of foliar application of selenium on its uptake and speciation in carrot. *Food Chem.* 2009, 115, 1357–1363. [CrossRef]
- 60. Qin, S.; Gao, J.; Huang, K. Effects of different selenium sources on tissue selenium concentrations, blood GSH-Px activities and plasma interleukin levels in finishing lambs. *Biol. Trace Elem. Res.* **2007**, *116*, 91–102. [CrossRef]
- Kruzhel, B.; Bakowska, M.; Vovk, S.; Nowakowska, E.; Sergei, P. Selenium in the diet of ruminants. Acta Sci. Polonorum. Zootech. 2014, 13, 5–16.
- 62. Lyons, M.P.; Papazyan, T.T.; Surai, P.F. Selenium in Food Chain and Animal Nutrition: Lessons from Nature -Review-. Asian-Australas. J. Anim. Sci. 2007, 20, 1135–1155. [CrossRef]
- 63. Hawrylak-Nowak, B.; Hasanuzzaman, M.; Matraszek-Gawron, R. Mechanisms of Selenium-Induced Enhancement of Abiotic Stress Tolerance in Plants. In *Plant Nutrients and Abiotic Stress Tolerance*; Springer: Singapore, 2018; pp. 269–295.
- 64. Puccinelli, M.; Malorgio, F.; Pezzarossa, B. Selenium Enrichment of Horticultural Crops. Molecules 2017, 22, 933. [CrossRef]
- 65. Wang, P.; Lombi, E.; Zhao, F.-J.; Kopittke, P.M. Nanotechnology: A New Opportunity in Plant Sciences. *Trends Plant Sci.* 2016, 21, 699–712. [CrossRef]
- Wang, M.; Ali, F.; Wang, M.; Dinh, Q.T.; Zhou, F.; Bañuelos, G.S.; Liang, D. Understanding boosting selenium accumulation in Wheat (*Triticum aestivum* L.) following foliar selenium application at different stages, forms, and doses. *Environ. Sci. Pollut. Res.* 2020, 27, 717–728. [CrossRef] [PubMed]
- Lidon, F.C.; Oliveira, K.; Ribeiro, M.M.; Pelica, J.; Pataco, I.; Ramalho, J.C.; Leitão, A.E.; Almeida, A.S.; Campos, P.S.; Ribeiro-Barros, A.I.; et al. Selenium biofortification of rice grains and implications on macronutrients quality. *J. Cereal Sci.* 2018, *81*, 22–29. [CrossRef]

- Lidon, F.C.; Oliveira, K.; Galhano, C.; Guerra, M.; Ribeiro, M.M.; Pelica, J.; Pataco, I.; Ramalho, J.C.; Leitão, A.E.; Almeida, A.S.; et al. Selenium biofortification of rice through foliar application with selenite and selenate. *Exp. Agric.* 2019, 55, 528–542. [CrossRef]
- 69. Pannico, A.; El-Nakhel, C.; Kyriacou, M.C.; Giordano, M.; Stazi, S.R.; De Pascale, S.; Rouphael, Y. Combating Micronutrient Deficiency and Enhancing Food Functional Quality Through Selenium Fortification of Select Lettuce Genotypes Grown in a Closed Soilless System. *Front. Plant Sci.* **2019**, *10*, 1495. [CrossRef] [PubMed]
- 70. Alfthan, G.; Eurola, M.; Ekholm, P.; Venäläinen, E.-R.; Root, T.; Korkalainen, K.; Hartikainen, H.; Salminen, P.; Hietaniemi, V.; Aspila, P.; et al. Effects of nationwide addition of selenium to fertilizers on foods, and animal and human health in Finland: From deficiency to optimal selenium status of the population. *J. Trace Elem. Med. Biol.* 2015, *31*, 142–147. [CrossRef]
- 71. Ebrahimi, N.; Stoddard, F.L.; Hartikainen, H.; Seppänen, M.M. Plant species and growing season weather influence the efficiency of selenium biofortification. *Nutr. Cycl. Agroecosystems* **2019**, *114*, 111–124. [CrossRef]
- 72. Gupta, M.; Gupta, S. An Overview of Selenium Uptake, Metabolism, and Toxicity in Plants. *Front. Plant Sci.* 2017, 7, 2074. [CrossRef]
- 73. Dongli, L.; Liang, D.; Qin, S.; Feng, P.; Wu, X. Effects of selenite and selenate application on growth and shoot selenium accumulation of pak choi (*Brassica chinensis* L.) during successive planting conditions. *Environ. Sci. Pollut. Res.* **2015**, *22*, 11076–11086. [CrossRef]
- 74. White, P.J. Selenium accumulation by plants. Ann. Bot. 2015, 117, 217–235. [CrossRef]
- 75. Wu, Z.; Banuelos, G.S.; Lin, Z.Q.; Liu, Y.; Yuan, L.; Yin, X.; Li, M. Biofortification and phytoremediation of selenium in China. *Front. Plant Sci.* **2015**, *6*, 136. [CrossRef]
- Wan, Y.; Yu, Y.; Wang, Q.; Qiao, Y.; Li, H. Cadmium uptake dynamics and translocation in rice seedling: Influence of different forms of selenium. *Ecotoxicol. Environ. Saf.* 2016, 133, 127–134. [CrossRef]
- 77. Winkel, L.H.E.; Vriens, B.; Jones, G.D.; Schneider, L.S.; Pilon-Smits, E.; Bañuelos, G.S. Selenium Cycling Across Soil-Plant-Atmosphere Interfaces: A Critical Review. *Nutrients* **2015**, *7*, 4199–4239. [CrossRef] [PubMed]
- 78. Zhang, L.; Hu, B.; Li, W.; Che, R.; Deng, K.; Li, H.; Yu, F.; Ling, H.; Li, Y.; Chu, C. OsPT2, a phosphate transporter, is involved in the active uptake of selenite in rice. *New Phytol.* **2014**, *201*, 1183–1191. [CrossRef] [PubMed]
- 79. da Silva, D.F.; Cipriano, P.E.; de Souza, R.R.; Júnior, M.S.; Faquin, V.; de Souza Silva, M.L.; Guilherme, L.R.G. Biofortification with selenium and implications in the absorption of macronutrients in Raphanus sativus L. *J. Food Compos. Anal.* 2020, *86*, 103382. [CrossRef]
- Zhou, F.; Peng, Q.; Wang, M.; Liu, N.; Dinh, Q.T.; Zhai, H.; Xue, M.; Liang, D. Influence of processing methods and exogenous selenium species on the content and in vitro bioaccessibility of selenium in Pleurotus eryngii. *Food Chem.* 2020, 338, 127661. [CrossRef] [PubMed]
- 81. de Oliveira, A.P.; Naozuka, J. Preliminary results on the feasibility of producing selenium-enriched pink (*Pleurotus djamor*) and white (*Pleurotus ostreatus*) oyster mushrooms: Bioaccumulation, bioaccessibility, and Se-proteins distribution. *Microchem. J.* **2019**, 145, 1143–1150. [CrossRef]
- 82. Barret, M.; Morrissey, J.P.; O'Gara, F. Functional genomics analysis of plant growth-promoting rhizobacterial traits involved in rhizosphere competence. *Biol. Fertil. Soils* 2011, 47, 729–743. [CrossRef]
- Sarwar, N.; Imran, M.; Shaheen, M.R.; Ishaque, W.; Kamran, M.A.; Matloob, A.; Rehim, A.; Hussain, S. Phytoremediation strategies for soils contaminated with heavy metals: Modifications and future perspectives. *Chemosphere* 2017, 171, 710–721. [CrossRef]
- 84. Fordyce, F. Selenium Geochemistry and Health. AMBIO 2007, 36, 94–97. [CrossRef]
- 85. Acuña, J.J.; Jorquera, M.A.; Barra, P.J.; Crowley, D.E.; Mora, M.D.L.L. Selenobacteria selected from the rhizosphere as a potential tool for Se biofortification of wheat crops. *Biol. Fertil. Soils* **2012**, *49*, 175–185. [CrossRef]
- Yasin, M.; El Mehdawi, A.F.; Jahn, C.E.; Anwar, A.; Turner, M.F.S.; Faisal, M.; Pilon-Smits, E.A.H. Seleniferous soils as a source for production of selenium-enriched foods and potential of bacteria to enhance plant selenium uptake. *Plant Soil* 2014, 386, 385–394. [CrossRef]
- Golubkina, N.; Zamana, S.; Seredin, T.; Poluboyarinov, P.; Sokolov, S.; Baranova, H.; Krivenkov, L.; Pietrantonio, L.; Caruso, G. Effect of Selenium Biofortification and Beneficial Microorganism Inoculation on Yield, Quality and Antioxidant Properties of Shallot Bulbs. *Plants* 2019, *8*, 102. [CrossRef]
- 88. Yuan, Y.; Liu, D.; Huang, X.; Wang, S.; Qiu, R.; Zhang, Z.; Ming, J. Effect of Enterobacter sp. EG16 on Selenium biofortification and speciation in pak choi (*Brassica rapa* ssp. *chinensis*). *Sci. Hortic.* **2023**, *310*, 111723. [CrossRef]
- Husen, A.; Siddiqi, K.S. Phytosynthesis of nanoparticles: Concept, controversy and application. *Nanoscale Res. Lett.* 2014, *9*, 229. [CrossRef] [PubMed]
- Oremland, R.S.; Herbel, M.J.; Blum, J.S.; Langley, S.; Beveridge, T.J.; Ajayan, P.M.; Sutto, T.; Ellis, A.V.; Curran, S. Structural and Spectral Features of Selenium Nanospheres Produced by Se-Respiring Bacteria. *Appl. Environ. Microbiol.* 2004, 70, 52–60. [CrossRef]
- Kieliszek, M.; Błażejak, S.; Gientka, I.; Bzducha-Wróbel, A. Accumulation and metabolism of selenium by yeast cells. *Appl. Microbiol. Biotechnol.* 2015, 99, 5373–5382. [CrossRef]
- Gharieb, M.; Gadd, G. Role of glutathione in detoxification of metal(loid)s by Saccharomyces cerevisiae. *Biometals* 2004, 17, 183–188. [CrossRef]
- 93. Rayman, M.P. Selenium and human health. Lancet 2012, 379, 1256–1268. [CrossRef]

- 94. White, P.J.; Broadley, M.R. Biofortification of crops with seven mineral elements often lacking in human diets–iron, zinc, copper, calcium, magnesium, selenium and iodine. *New Phytol.* **2009**, *182*, 49–84. [CrossRef]
- 95. Zhu, Y.-G.; Pilon-Smits, E.A.H.; Zhao, F.-J.; Williams, P.N.; Meharg, A.A. Selenium in higher plants: Understanding mechanisms for biofortification and phytoremediation. *Trends Plant Sci.* 2009, *14*, 436–442. [CrossRef]
- Ates, D.; Sever, T.; Aldemir, S.; Yagmur, B.; Temel, H.Y.; Kaya, H.B.; Tanyolac, B. Identification QTLs controlling genes for Se uptake in lentil seeds. *PLoS ONE* 2016, 11, e0149210.
- 97. Wang, P.; Wang, H.; Liu, Q.; Tian, X.; Shi, Y.; Zhang, X. QTL mapping of selenium content using a RIL population in wheat. *PLoS* ONE 2017, 12, e0184351. [CrossRef] [PubMed]
- Schiavon, M.; Nardi, S.; dalla Vecchia, F.; Ertani, A. Selenium biofortification in the 21st century: Status and challenges for healthy human nutrition. *Plant Soil* 2020, 453, 245–270. [CrossRef] [PubMed]
- Garg, M.; Sharma, N.; Sharma, S.; Kapoor, P.; Kumar, A.; Chunduri, V.; Arora, P. Biofortified Crops Generated by Breeding, Agronomy, and Transgenic Approaches Are Improving Lives of Millions of People around the World. *Front. Nutr.* 2018, *5*, 12. [CrossRef] [PubMed]
- LeDuc, D.L.; AbdelSamie, M.; Móntes-Bayon, M.; Wu, C.P.; Reisinger, S.J.; Terry, N. Overexpressing both ATP sulfurylase and selenocysteine methyltransferase enhances selenium phytoremediation traits in Indian mustard. *Environ. Pollut.* 2006, 144, 70–76. [CrossRef] [PubMed]
- 101. Liu, C.; Ding, S.; Zhang, A.; Hong, K.; Jiang, H.; Yang, S.; Ruan, B.; Zhang, B.; Dong, G.; Guo, L.; et al. Development of nutritious rice with high zinc/selenium and low cadmium in grains through QTL pyramiding. *J. Integr. Plant Biol.* 2020, 62, 349–359. [CrossRef]
- 102. Liang, Y.; Farooq, M.U.; Zeng, R.; Tang, Z.; Zhang, Y.; Zheng, T.; Zhu, J. Breeding of selenium rich red glutinous rice, protein extraction and analysis of the distribution of selenium in grain. *Int. J. Agric. Biol.* **2018**, *20*, 1005–1011.
- 103. Mao, H.; Wang, J.; Wang, Z.; Zan, Y.; Lyons, G.; Zou, C. Using agronomic biofortification to boost zinc, selenium, and iodine concentrations of food crops grown on the loess plateau in China. *J. Soil Sci. Plant Nutr.* **2014**, *14*, 459–470. [CrossRef]
- 104. Yuan, L.; Yin, X.; Zhu, Y.; Li, F.; Huang, Y.; Liu, Y.; Lin, Z. Selenium in plants and soils, and selenosis in Enshi, China: Implications for selenium biofortification. *Phytoremediation Biofortification Two Sides One Coin* **2012**, 7–31. [CrossRef]
- Rayman, M.P. The use of high-selenium yeast to raise selenium status: How does it measure up? Br. J. Nutr. 2004, 92, 557–573.
 [CrossRef]
- Jeong, D.; Lee, K.H. Bimodal actions of selenium essential for antioxidant and toxic pro-oxidant activities: The selenium paradox (Review). Mol. Med. Rep. 2011, 5, 299–304. [CrossRef] [PubMed]
- 107. Navarro-Alarcon, M.; Cabrera-Vique, C. Selenium in food and the human body: A review. *Sci. Total. Environ.* **2008**, 400, 115–141. [CrossRef]
- 108. Alfthan, G.; Aspila, P.; Ekholm, P.; Eurola, M.; Hartikainen, H.; Hero, H.; Hietaniemi, V.; Root, T.; Salminen, P.; Venäläinen, E.R.; et al. Nationwide supplementation of sodium selenate to commercial fertilizers: History and 25-year results from the Finnish selenium monitoring programme. *Combat. Micronutr. Defic. Food-Based Approaches* 2011, 312–337. [CrossRef]
- Hu, T.; Hui, G.; Li, H.; Guo, Y. Selenium biofortification in Hericium erinaceus (Lion's Mane mushroom) and its in vitro bioaccessibility. *Food Chem.* 2020, 331, 127287. [CrossRef] [PubMed]
- 110. Yläranta, T. Increasing the Selenium Content of Cereal and Grass Crops in Finland; Yliopistopaino: Cygnaeuksenkatu, Finland, 1985.
- 111. Ligowe, I.; Young, S.; Ander, E.; Kabambe, V.; Chilimba, A.; Bailey, E.; Lark, R.; Nalivata, P. Selenium biofortification of crops on a Malawi Alfisol under conservation agriculture. *Geoderma* **2020**, *369*, 114315. [CrossRef]
- 112. Alfthan, G. Longitudinal study on the selenium status of healthy adults in Finland during 1975–1984. *Nutr. Res.* **1988**, *8*, 467–476. [CrossRef]
- 113. Bañuelos, G.S.; Lin, Z.Q.; Broadley, M. Selenium biofortification. Selenium Plants Mol. Physiol. Ecol. Evol. Asp. 2017, 11, 231–255.
- 114. He, Y.; Xiang, Y.; Zhou, Y.; Yang, Y.; Zhang, J.; Huang, H.; Shang, C.; Luo, L.; Gao, J.; Tang, L. Selenium contamination, consequences and remediation techniques in water and soils: A review. *Environ. Res.* **2018**, *164*, 288–301. [CrossRef]
- 115. Hurst, R.; Siyame, E.W.P.; Young, S.D.; Chilimba, A.D.C.; Joy, E.J.M.; Black, C.R.; Ander, E.L.; Watts, M.J.; Chilima, B.; Gondwe, J.; et al. Soil-type influences human selenium status and underlies widespread selenium deficiency risks in Malawi. *Sci. Rep.* 2013, *3*, srep01425. [CrossRef]
- Mehdi, Y.; Hornick, J.-L.; Istasse, L.; Dufrasne, I. Selenium in the Environment, Metabolism and Involvement in Body Functions. *Molecules* 2013, 18, 3292–3311. [CrossRef]
- Bitterli, C.; Bañuelos, G.S.; Schulin, R. Use of transfer factors to characterize uptake of selenium by plants. J. Geochem. Explor. 2010, 107, 206–216. [CrossRef]
- 118. Ei, H.H.; Zheng, T.; Farooq, M.U.; Zeng, R.; Su, Y.; Zhang, Y.; Zhu, J. Impact of selenium, zinc and their interaction on key enzymes, grain yield, selenium, zinc concentrations, and seedling vigor of biofortified rice. *Environ. Sci. Pollut. Res.* 2020, 27, 16940–16949. [CrossRef]
- 119. Hasanuzzaman, M.; Bhuyan, M.B.; Raza, A.; Hawrylak-Nowak, B.; Matraszek-Gawron, R.; Al Mahmud, J.; Fujita, M. Selenium in plants: Boon or bane? *Environ. Exp. Bot.* 2020, 178, 104170. [CrossRef]
- 120. Feng, R.; Wei, C.; Tu, S. The roles of selenium in protecting plants against abiotic stresses. *Environ. Exp. Bot.* **2013**, *87*, 58–68. [CrossRef]
- 121. Mroczek-Zdyrska, M.; Wójcik, M. The Influence of Selenium on Root Growth and Oxidative Stress Induced by Lead in Vicia faba L. minor Plants. *Biol. Trace Element Res.* 2011, 147, 320–328. [CrossRef]

- 122. Ashraf, M.A.; Akbar, A.; Parveen, A.; Rasheed, R.; Hussain, I.; Iqbal, M. Phenological application of selenium differentially improves growth, oxidative defense and ion homeostasis in maize under salinity stress. *Plant Physiol. Biochem.* 2018, 123, 268–280. [CrossRef]
- 123. Bybordi, A.; Saadat, S.; Zargaripour, P. The effect of zeolite, selenium and silicon on qualitative and quantitative traits of onion grown under salinity conditions. *Arch. Agron. Soil Sci.* **2017**, *64*, 520–530. [CrossRef]
- 124. Habibi, G. Physiological, photochemical and ionic responses of sunflower seedlings to exogenous selenium supply under salt stress. *Acta Physiol. Plant.* 2017, *39*, 213. [CrossRef]
- 125. Alyemeni, M.N.; Ahanger, M.A.; Wijaya, L.; Alam, P.; Bhardwaj, R.; Ahmad, P. Selenium mitigates cadmium-induced oxidative stress in tomato (*Solanum lycopersicum* L.) plants by modulating chlorophyll fluorescence, osmolyte accumulation, and antioxidant system. *Protoplasma* 2017, 255, 459–469. [CrossRef]
- 126. Iqbal, M.; Hussain, I.; Liaqat, H.; Ashraf, M.A.; Rasheed, R.; Rehman, A.U. Exogenously applied selenium reduces oxidative stress and induces heat tolerance in spring wheat. *Plant Physiol. Biochem.* **2015**, *94*, 95–103. [CrossRef]
- 127. Pukacka, S.; Ratajczak, E.; Kalemba, E. The protective role of selenium in recalcitrant Acer saccharium L. seeds subjected to desiccation. *J. Plant Physiol.* 2011, 168, 220–225. [CrossRef]
- 128. Karimi, R.; Ghabooli, M.; Rahimi, J.; Amerian, M. Effects of foliar selenium application on some physiological and phytochemical parameters of Vitis vinifera L. cv. Sultana under salt stress. *J. Plant Nutr.* **2020**, *43*, 2226–2242. [CrossRef]
- Khan, S.; Waqas, M.; Ding, F.; Shamshad, I.; Arp, H.P.H.; Li, G. The influence of various biochars on the bioaccessibility and bioaccumulation of PAHs and potentially toxic elements to turnips (*Brassica rapa* L.). *J. Hazard. Mater.* 2015, 300, 243–253. [CrossRef] [PubMed]
- Moynihan, M.; Peterson, K.E.; Cantoral, A.; Song, P.X.K.; Jones, A.; Solano-González, M.; Meeker, J.D.; Basu, N.; Tellez-Rojo, M.M. Dietary predictors of urinary cadmium among pregnant women and children. *Sci. Total Environ.* 2017, 575, 1255–1262. [CrossRef] [PubMed]
- 131. Suleiman, N.; Ibitoye, E.; Jimoh, A.; Sani, Z. Assessment of heavy metals in chicken feeds available in Sokoto, Nigeria. *Sokoto J. Vet.- Sci.* **1970**, *13*, 17–21. [CrossRef]
- Deng, S.; Li, P.; Li, Y.; Ran, Z.; Peng, Y.; Yang, S.; Yu, J. Alleviating Cd translocation and accumulation in soil-rice systems: Combination of foliar spraying of nano-Si or nano-Se and soil application of nano-humus. *Soil Use Manag.* 2021, 37, 319–329. [CrossRef]
- Li, H.; Liu, X.; Wassie, M.; Chen, L. Selenium supplementation alleviates cadmium-induced damages in tall fescue through modulating antioxidant system, photosynthesis efficiency, and gene expression. *Environ. Sci. Pollut. Res.* 2020, 27, 9490–9502. [CrossRef]
- 134. Huang, B.; Xin, J.; Dai, H.; Zhou, W. Effects of Interaction between Cadmium (Cd) and Selenium (Se) on Grain Yield and Cd and Se Accumulation in a Hybrid Rice (*Oryza sativa*) System. *J. Agric. Food Chem.* **2017**, *65*, 9537–9546. [CrossRef]
- 135. Hu, L.; Wang, X.; Wu, D.; Zhang, B.; Fan, H.; Shen, F.; Liao, Y.; Huang, X.; Gao, G. Effects of organic selenium on absorption and bioaccessibility of arsenic in radish under arsenic stress. *Food Chem.* **2020**, *344*, 128614. [CrossRef]
- Feng, R.; Zhao, P.; Zhu, Y.; Yang, J.; Wei, X.; Yang, L.; Liu, H.; Rensing, C.; Ding, Y. Application of inorganic selenium to reduce accumulation and toxicity of heavy metals (metalloids) in plants: The main mechanisms, concerns, and risks. *Sci. Total. Environ.* 2021, 771, 144776. [CrossRef]
- 137. Guo, Y.; Mao, K.; Cao, H.; Ali, W.; Lei, D.; Teng, D.; Chang, C.; Yang, X.; Yang, Q.; Niazi, N.K.; et al. Exogenous selenium (cadmium) inhibits the absorption and transportation of cadmium (selenium) in rice. *Environ. Pollut.* **2021**, *268*, 115829. [CrossRef]
- Huang, C.; Qin, N.; Sun, L.; Yu, M.; Hu, W.; Qi, Z. Selenium Improves Physiological Parameters and Alleviates Oxidative Stress in Strawberry Seedlings under Low-Temperature Stress. Int. J. Mol. Sci. 2018, 19, 1913. [CrossRef] [PubMed]
- Hasanuzzaman, M.; Nahar, K.; Alam, M.; Fujita, M. Modulation of Antioxidant Machinery and the Methylglyoxal Detoxification System in Selenium-Supplemented Brassica napus Seedlings Confers Tolerance to High Temperature Stress. *Biol. Trace Element Res.* 2014, 161, 297–307. [CrossRef] [PubMed]
- Hawrylak-Nowak, B.; Dresler, S.; Rubinowska, K.; Matraszek-Gawron, R.; Woch, W.; Hasanuzzaman, M. Selenium biofortification enhances the growth and alters the physiological response of lamb's lettuce grown under high temperature stress. *Plant Physiol. Biochem.* 2018, 127, 446–456. [CrossRef] [PubMed]
- 141. Alves, L.R.; Rossatto, D.R.; Rossi, M.L.; Martinelli, A.P.; Gratão, P.L. Selenium improves photosynthesis and induces ultrastructural changes but does not alleviate cadmium-stress damages in tomato plants. *Protoplasma* **2019**, 257, 597–605. [CrossRef]
- 142. Taha, R.; Seleiman, M.; Shami, A.; Alhammad, B.; Mahdi, A. Integrated Application of Selenium and Silicon Enhances Growth and Anatomical Structure, Antioxidant Defense System and Yield of Wheat Grown in Salt-Stressed Soil. *Plants* 2021, 10, 1040. [CrossRef] [PubMed]
- 143. Djanaguiraman, M.; Prasad, P.; Seppanen, M. Selenium protects sorghum leaves from oxidative damage under high temperature stress by enhancing antioxidant defense system. *Plant Physiol. Biochem.* **2010**, *48*, 999–1007. [CrossRef]
- 144. Nawaz, F.; Naeem, M.; Ashraf, M.Y.; Tahir, M.N.; Zulfiqar, B.; Salahuddin, M.; Shabbir, R.N.; Aslam, M. Selenium Supplementation Affects Physiological and Biochemical Processes to Improve Fodder Yield and Quality of Maize (*Zea mays* L.) under Water Deficit Conditions. Front. Plant Sci. 2016, 7, 1438. [CrossRef]
- 145. Huang, A.; Huang, K.; Peng, J.; Huang, S.; Bi, X.; Zhai, R.; Tan, H. Effects of foliar spraying of selenium fertilizer on seleniumenriched content, heavy metal content and yield of sweet corn grain. *J. South. Agric.* **2019**, *50*, 40–44.

- 146. Jiang, C.; Zu, C.; Lu, D.; Zheng, Q.; Shen, J.; Wang, H.; Li, D. Effect of exogenous selenium supply on photosynthesis, Na+ accumulation and antioxidative capacity of maize (*Zea mays* L.) under salinity stress. *Sci. Rep.* **2017**, *7*, srep42039. [CrossRef]
- Yildiztugay, E.; Ozfidan-Konakci, C.; Kucukoduk, M.; Tekis, S.A. The impact of selenium application on enzymatic and non-enzymatic antioxidant systems in Zea mays roots treated with combined osmotic and heat stress. *Arch. Agron. Soil Sci.* 2017, 63, 261–275. [CrossRef]
- 148. Elkelish, A.A.; Soliman, M.H.; Alhaithloul, H.A.; El-Esawi, M.A. Selenium protects wheat seedlings against salt stress-mediated oxidative damage by up-regulating antioxidants and osmolytes metabolism. *Plant Physiol. Biochem.* 2019, 137, 144–153. [CrossRef] [PubMed]
- Sattar, A.; Cheema, M.A.; Sher, A.; Ijaz, M.; Ul-Allah, S.; Nawaz, A.; Abbas, T.; Ali, Q. Physiological and biochemical attributes of bread wheat (*Triticum aestivum* L.) seedlings are influenced by foliar application of silicon and selenium under water deficit. *Acta Physiol. Plant.* 2019, 41, 146. [CrossRef]
- Chauhan, R.; Awasthi, S.; Tripathi, P.; Mishra, S.; Dwivedi, S.; Niranjan, A.; Mallick, S.; Tripathi, P.; Pande, V.; Tripathi, R.D. Selenite modulates the level of phenolics and nutrient element to alleviate the toxicity of arsenite in rice (*Oryza sativa* L.). *Ecotoxicol. Environ. Saf.* 2017, 138, 47–55. [CrossRef]
- 151. Singh, R.; Upadhyay, A.; Singh, D. Regulation of oxidative stress and mineral nutrient status by selenium in arsenic treated crop plant Oryza sativa. *Ecotoxicol. Environ. Saf.* **2017**, *148*, 105–113. [CrossRef]
- 152. Moulick, D.; Ghosh, D.; Santra, S.C. Evaluation of effectiveness of seed priming with selenium in rice during germination under arsenic stress. *Plant Physiol. Biochem.* **2016**, *109*, 571–578. [CrossRef]
- 153. Hernández-Hernández, H.; Quiterio-Gutiérrez, T.; Cadenas-Pliego, G.; Ortega-Ortiz, H.; Hernández-Fuentes, A.D.; Cabrera de la Fuente, M.; Juárez-Maldonado, A. Impact of selenium and copper nanoparticles on yield, antioxidant system, and fruit quality of tomato plants. *Plants* 2019, *8*, 355. [CrossRef] [PubMed]
- 154. Shekari, L.; Kamelmanesh, M.M.; Mozafariyan, M.; Hasanuzzaman, M.; Sadeghi, F. Role of selenium in mitigation of cadmium toxicity in pepper grown in hydroponic condition. *J. Plant Nutr.* **2016**, *40*, 761–772. [CrossRef]
- 155. Mozafariyan, M.; Shekari, L.; Hawrylak-Nowak, B.; Kamelmanesh, M.M. Protective Role of Selenium on Pepper Exposed to Cadmium Stress During Reproductive Stage. *Biol. Trace Element Res.* **2014**, *160*, 97–107. [CrossRef]
- Astaneh, R.K.; Bolandnazar, S.; Nahandi, F.Z.; Oustan, S. Effects of selenium on enzymatic changes and productivity of garlic under salinity stress. *South Afr. J. Bot.* 2019, 121, 447–455. [CrossRef]
- 157. Haghighi, M.; Sheibanirad, A.; Pessarakli, M. Effects of selenium as a beneficial element on growth and photosynthetic attributes of greenhouse cucumber. *J. Plant Nutr.* **2015**, *39*, 1493–1498. [CrossRef]
- 158. Handa, N.; Kohli, S.K.; Thukral, A.K.; Bhardwaj, R.; Alyemeni, M.N.; Wijaya, L.; Ahmad, P. Protective role of selenium against chromium stress involving metabolites and essential elements in Brassica juncea L. seedlings. 3 Biotech 2018, 8, 66. [CrossRef] [PubMed]
- Mroczek-Zdyrska, M.; Strubińska, J.; Hanaka, A. Selenium Improves Physiological Parameters and Alleviates Oxidative Stress in Shoots of Lead-Exposed Vicia faba L. minor Plants Grown Under Phosphorus-Deficient Conditions. J. Plant Growth Regul. 2016, 36, 186–199. [CrossRef]
- Tavakoli, S.; Enteshari, S.; Yousefifard, M. Investigation of the effect of selenium on growth, antioxidant capacity and secondary metabolites in Melissa officinalis. *Iran. J. Plant Physiol.* 2020, 10, 3125–3134.
- Zahedi, S.M.; Abdelrahman, M.; Hosseini, M.S.; Hoveizeh, N.F.; Tran, L.-S.P. Alleviation of the effect of salinity on growth and yield of strawberry by foliar spray of selenium-nanoparticles. *Environ. Pollut.* 2019, 253, 246–258. [CrossRef] [PubMed]
- 162. Zahedi, S.M.; Hosseini, M.S.; Meybodi, N.D.H.; da Silva, J.A.T. Foliar application of selenium and nano-selenium affects pomegranate (Punica granatum cv. Malase Saveh) fruit yield and quality. *South Afr. J. Bot.* **2019**, *124*, 350–358. [CrossRef]
- 163. Manaf, H.H. Beneficial effects of exogenous selenium, glycine betaine and seaweed extract on salt stressed cowpea plant. *Ann. Agric. Sci.* **2016**, *61*, 41–48. [CrossRef]
- Han, D.; Xiong, S.; Tu, S.; Liu, J.; Chen, C. Interactive effects of selenium and arsenic on growth, antioxidant system, arsenic and selenium species of Nicotiana tabacum L. *Environ. Exp. Bot.* 2015, 117, 12–19. [CrossRef]
- 165. Underwood, E.; Suttle, N. The Mineral Nutrition of Livestock, 3rd ed.; CABI: Wallingford, UK, 1999.
- 166. Mehdi, Y.; Dufrasne, I. Selenium in Cattle: A Review. Molecules 2016, 21, 545. [CrossRef]
- 167. Moir, D.C.; Masters, H.G. Selenium deficiency and hepatosis dietica in pigs. Aust. Vet. J. 1970, 55, 360–366. [CrossRef]
- 168. Helmer, C.; Hannemann, R.; Humann-Ziehank, E.; Kleinschmidt, S.; Koelln, M.; Kamphues, J.; Ganter, M. A Case of Concurrent Molybdenosis, Secondary Copper, Cobalt and Selenium Deficiency in a Small Sheep Herd in Northern Germany. *Animals* 2021, 11, 1864. [CrossRef]
- Hosnedlova, B.; Kepinska, M.; Skalickova, S.; Fernandez, C.; Ruttkay-Nedecky, B.; Malevu, T.D.; Sochor, J.; Baron, M.; Melcova, M.; Zidkova, J.; et al. A Summary of New Findings on the Biological Effects of Selenium in Selected Animal Species—A Critical Review. Int. J. Mol. Sci. 2017, 18, 2209. [CrossRef]
- Sordillo, L.M. Selenium-Dependent Regulation of Oxidative Stress and Immunity in Periparturient Dairy Cattle. *Vet.- Med. Int.* 2013, 2013, 154045. [CrossRef]
- 171. Koller, L.D.; Exon, J.H. The two faces of selenium-deficiency and toxicity–are similar in animals and man. *Can. J. Vet.-Res.* = *Rev. Can. Rech. Vet.* **1986**, *50*, 297–306.
- 172. Kumssa, D.; Joy, E.; Broadley, M. Global Trends (1961–2017) in Human Dietary Potassium Supplies. *Nutrients* 2021, 13, 1369. [CrossRef] [PubMed]

- 173. McDowell, L.R. Vitamins in Animal and Human Nutrition, 2nd ed.; Wiley-Blackwell: Oxford, UK, 2000. [CrossRef]
- 174. Gissel-Nielsen, G. Comparison of selenium treatments of crops in the field. *Biol. Trace Element Res.* **1986**, *10*, 209–213. [CrossRef] [PubMed]
- Eliopoulos, G.D.; Eliopoulos, I.-P.D.; Tsioubri, M.; Economou-Eliopoulos, M. Distribution of Selenium in the Soil–Plant– Groundwater System: Factors Controlling Its Bio-Accumulation. *Minerals* 2020, 10, 795. [CrossRef]
- 176. Li, H.; Zhang, J.; Xia, Y.; Pan, W.; Zhou, D. Antagonistic effect of nano-selenium on hepatocyte apoptosis induced by DEHP via PI3K/AKT pathway in chicken liver. *Ecotoxicol. Environ. Saf.* 2021, 218, 112282. [CrossRef] [PubMed]
- 177. Shen, Q.; Zhang, B.; Xu, R.; Wang, Y.; Ding, X.; Li, P. Antioxidant activity in vitro of the selenium-contained protein from the Se-enriched Bifidobacterium animalis 01. *Anaerobe* **2010**, *16*, 380–386. [CrossRef]
- 178. Zhao, M.; Wen, K.; Xue, Y.; Liu, L.; Geng, T.; Gong, D.; Yu, L. Probing the effects of dietary selenised glucose on the selenium concentration, quality, and antioxidant activity of eggs and production performances of laying hens. *Animal* 2021, 15, 100374. [CrossRef]
- Li, J.-L.; Jiang, C.-Y.; Li, S.; Xu, S.-W. Cadmium induced hepatotoxicity in chickens (*Gallus domesticus*) and ameliorative effect by selenium. *Ecotoxicol. Environ. Saf.* 2013, 96, 103–109. [CrossRef] [PubMed]
- Hao, P.; Zhu, Y.; Wang, S.; Wan, H.; Chen, P.; Wang, Y.; Cheng, Z.; Liu, Y.; Liu, J. Selenium Administration Alleviates Toxicity of Chromium(VI) in the Chicken Brain. *Biol. Trace Element Res.* 2016, *178*, 127–135. [CrossRef]
- Han, X.; Qin, P.; Li, W.; Ma, Q.; Ji, C.; Zhang, J.; Zhao, L. Effect of sodium selenite and selenium yeast on performance, egg quality, antioxidant capacity, and selenium deposition of laying hens. *Poult. Sci.* 2017, *96*, 3973–3980. [CrossRef]
- 182. Shojadoost, B.; Kulkarni, R.R.; Yitbarek, A.; Laursen, A.; Taha-Abdelaziz, K.; Alkie, T.N.; Barjesteh, N.; Quinteiro-Filho, W.M.; Smith, T.K.; Sharif, S. Dietary selenium supplementation enhances antiviral immunity in chickens challenged with low pathogenic avian influenza virus subtype H9N2. *Vet.-Immunol. Immunopathol.* 2018, 207, 62–68. [CrossRef]
- 183. Wang, Y.; Jiang, L.; Li, Y.; Luo, X.; He, J. Effect of Different Selenium Supplementation Levels on Oxidative Stress, Cytokines, and Immunotoxicity in Chicken Thymus. *Biol. Trace Element Res.* 2016, 172, 488–495. [CrossRef] [PubMed]
- Chen, X.; Ren, F.; Hesketh, J.; Shi, X.; Li, J.; Gan, F.; Huang, K. Selenium blocks porcine circovirus type 2 replication promotion induced by oxidative stress by improving GPx1 expression. *Free. Radic. Biol. Med.* 2012, 53, 395–405. [CrossRef] [PubMed]
- 185. Beck, M.A.; Levander, O.A.; Handy, J. Selenium Deficiency and Viral Infection. J. Nutr. 2003, 133, 1463S–1467S. [CrossRef]
- Reffett, J.K.; Spears, J.W.; Brown, T.T. Effect of Dietary Selenium and Vitamin E on the Primary and Secondary Immune Response in Lambs Challenged with Parainfluenza Virus. J. Anim. Sci. 1988, 66, 1520–1528. [CrossRef]
- 187. Khalili, M.; Chamani, M.; Amanlou, H.; Nikkhah, A.; Sadeghi, A.; Dehkordi, F.K.; Rafiei, M.; Shirani, V. The effect of feeding inorganic and organic selenium sources on the hematological blood parameters, reproduction and health of dairy cows in the transition period. *Acta Sci. Anim. Sci.* 2019, 42, e45371. [CrossRef]
- Sun, L.; Liu, G.; Xu, D.; Wu, Z.; Ma, L.; Victoria, S.-F.M.; Baumgard, L.H.; Bu, D. Milk selenium content and speciation in response to supranutritional selenium yeast supplementation in cows. *Anim. Nutr.* 2021, 7, 1087–1094. [CrossRef]
- 189. Hall, J.A.; Bobe, G.; Vorachek, W.R.; Kasper, K.; Traber, M.G.; Mosher, W.D.; Pirelli, G.J.; Gamroth, M. Effect of Supranutritional Organic Selenium Supplementation on Postpartum Blood Micronutrients, Antioxidants, Metabolites, and Inflammation Biomarkers in Selenium-Replete Dairy Cows. *Biol. Trace Element Res.* 2014, 161, 272–287. [CrossRef]
- 190. Pan, Q.; Huang, K.; He, K.; Lu, F. Effect of different selenium sources and levels on porcine circovirus type 2 replication in vitro. J. Trace Elements Med. Biol. 2008, 22, 143–148. [CrossRef] [PubMed]
- Lv, L.; Zhang, H.; Liu, Z.; Lei, L.; Feng, Z.; Zhang, D.; Zhao, S. Comparative study of yeast selenium vs. sodium selenite on growth performance, nutrient digestibility, anti-inflammatory and anti-oxidative activity in weaned piglets challenged by Salmonella typhimurium. *Innate Immun.* 2020, 26, 248–258. [CrossRef] [PubMed]
- 192. Shi, X.; Wang, W.; Zheng, S.; Zhang, Q.; Xu, S. Selenomethionine relieves inflammation in the chicken trachea caused by LPS though inhibiting the NF-κB pathway. *Biol. Trace Elem. Res.* **2020**, *194*, 525–535. [CrossRef] [PubMed]
- Muhammad, A.I.; Mohamed, D.A.; Chwen, L.T.; Akit, H.; Samsudin, A.A. Effect of Selenium Sources on Laying Performance, Egg Quality Characteristics, Intestinal Morphology, Microbial Population and Digesta Volatile Fatty Acids in Laying Hens. *Animals* 2021, 11, 1681. [CrossRef]
- 194. Lee, S.H.; Lillehoj, H.S.; Jang, S.I.; Jeong, M.S.; Xu, S.Z.; Kim, J.B.; Park, H.J.; Kim, H.R.; Lillehoj, E.P.; Bravo, D.M. Effects of in ovo injection with selenium on immune and antioxidant responses during experimental necrotic enteritis in broiler chickens. *Poult. Sci.* 2014, 93, 1113–1121. [CrossRef]
- 195. Mengistu, B.M.; Bitsue, H.K.; Huang, K. The Effects of Selenium-Enriched Probiotics on Growth Performance, Oocysts Shedding, Intestinal Cecal Lesion Scores, Antioxidant Capacity, and mRNA Gene Expression in Chickens Infected with Eimeria tenella. *Biol. Trace Element Res.* 2020, 199, 278–291. [CrossRef]
- 196. Hugejiletu, H.; Bobe, G.; Vorachek, W.R.; Gorman, M.E.; Mosher, W.D.; Pirelli, G.J.; Hall, J.A. Selenium Supplementation Alters Gene Expression Profiles Associated with Innate Immunity in Whole-Blood Neutrophils of Sheep. *Biol. Trace Element Res.* 2013, 154, 28–44. [CrossRef] [PubMed]
- 197. Jia, X.; Li, J.; Li, S.; Zhao, Q.; Zhang, K.; Tang, C.; Yang, Y.; Ma, Q.; Wang, J.; Zhao, Z.; et al. Effects of dietary supplementation with different levels of selenium yeast on growth performance, carcass characteristics, antioxidant capacity, and meat quality of Tan sheep. *Livest. Sci.* 2021, 255, 104783. [CrossRef]

- 198. Ebeid, T.; Zeweil, H.; Basyony, M.; Dosoky, W.; Badry, H. Fortification of rabbit diets with vitamin E or selenium affects growth performance, lipid peroxidation, oxidative status and immune response in growing rabbits. *Livest. Sci.* **2013**, *155*, 323–331. [CrossRef]
- Mattioli, S.; Dal Bosco, A.; Duarte, J.M.M.; D'Amato, R.; Castellini, C.; Beone, G.M.; Proietti, P. Use of Selenium-enriched olive leaves in the feed of growing rabbits: Effect on oxidative status, mineral profile and Selenium speciation of Longissimus dorsi meat. J. Trace Elem. Med. Biol. 2019, 51, 98–105. [CrossRef] [PubMed]
- 200. Thomson, C.D. Assessment of requirements for selenium and adequacy of selenium status: A review. *Eur. J. Clin. Nutr.* 2004, *58*, 391–402. [CrossRef] [PubMed]
- Thomson, C.D. Selenium and iodine intakes and status in New Zealand and Australia. Br. J. Nutr. 2004, 91, 661–672. [CrossRef]
 [PubMed]
- 202. Fernández-Quintela, A.; Milton-Laskibar, I.; Trepiana, J.; Gómez-Zorita, S.; Kajarabille, N.; Léniz, A.; González, M.; Portillo, M.P. Key Aspects in Nutritional Management of COVID-19 Patients. J. Clin. Med. 2020, 9, 2589. [CrossRef]
- Varo, P.E.R.T.I.I.; Alfthan, G.; Huttunen, J.K.; Aro, A. Nationwide selenium supplementation in Finland-effects on diet, blood and tissue levels, and health. In *Selenium in Biology and Human Health*; Springer: New York, NY, USA, 1994; Volume 98, pp. 198–218.
- Williams, P.; Islam, S.; Islam, R.; Jahiruddin, M.; Adomako, E.; Soliaman, A.R.M.; Rahman, G.K.M.M.; Lu, Y.; Deacon, C.; Zhu, Y.-G.; et al. Arsenic Limits Trace Mineral Nutrition (Selenium, Zinc, and Nickel) in Bangladesh Rice Grain. *Environ. Sci. Technol.* 2009, 43, 8430–8436. [CrossRef]
- 205. Williams, P.; Lombi, E.; Sun, G.-X.; Scheckel, K.; Zhu, Y.-G.; Feng, X.; Zhu, J.; Carey, A.-M.; Adomako, E.; Lawgali, Y.; et al. Selenium Characterization in the Global Rice Supply Chain. *Environ. Sci. Technol.* **2009**, *43*, 6024–6030. [CrossRef]
- Waegeneers, N.; Thiry, C.; De Temmerman, L.; Ruttens, A. Predicted dietary intake of selenium by the general adult population in Belgium. *Food Addit. Contam. Part A* 2013, 30, 278–285. [CrossRef]
- 207. Dreher, I.; Schütze, N.; Baur, A.; Hesse, K.; Schneider, D.; Köhrle, J.; Jakob, F. Selenoproteins Are Expressed in Fetal Human Osteoblast-like Cells. *Biochem. Biophys. Res. Commun.* **1998**, 245, 101–107. [CrossRef]
- Food, N.B.; Board, N. Institute of Medicine. Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium and Carotenoids; National Academy Press: Washington, DC, USA, 2000.
- Rayman, M.P.; Infante, H.G.; Sargent, M. Food-chain selenium and human health: Spotlight on speciation. Br. J. Nutr. 2008, 100, 238–253.
 [CrossRef]
- Bossola, M.; Di Stasio, E.; Viola, A.; Leo, A.; Carlomagno, G.; Monteburini, T.; Cenerelli, S.; Santarelli, S.; Boggi, R.; Miggiano, G.; et al. Dietary intake of trace elements, minerals, and vitamins of patients on chronic hemodialysis. *Int. Urol. Nephrol.* 2014, 46, 809–815. [CrossRef]
- Pograjc, L.; Stibilj, V.; Falnoga, I. Impact of Intensive Physical Activity on Selenium Status. *Biol. Trace Element Res.* 2011, 145, 291–299.
 [CrossRef]
- Valent, F.; Horvat, M.; Mazej, D.; Stibilj, V.; Barbone, F. Maternal Diet and Selenium Concentration in Human Milk From an Italian Population. J. Epidemiology 2011, 21, 285–292. [CrossRef]
- Al-Saleh, I.; El-Doush, I.; Billedo, G.; Mohamed, G.E.-D.; Yosef, G. Status of Selenium, Vitamin E, and Vitamin A among Saudi Adults: Potential Links with Common Endemic Diseases. J. Environ. Pathol. Toxicol. Oncol. 2007, 26, 221–243. [CrossRef]
- 214. Al-Awadi, F.M.; Srikumar, T.S. Determination of selenium concentration and its chemical forms in the milk of Kuwaiti and non-Kuwaiti lactating mothers. J. Trace Elem. Exp. Med. Off. Publ. Int. Soc. Trace Elem. Res. Hum. 2001, 14, 57–67. [CrossRef]
- 215. Özdemir, H.S.; Karadas, F.; Pappas, A.; Cassey, P.; Oto, G.; Tunçer. The Selenium Levels of Mothers and Their Neonates Using Hair, Breast Milk, Meconium, and Maternal and Umbilical Cord Blood in Van Basin. *Biol. Trace Element Res.* 2008, 122, 206–215. [CrossRef] [PubMed]
- 216. Joint, F. Vitamin and Mineral Requirements in Human Nutrition; Diamond Pocket Books (P) Ltd.: New Delhi, India, 2004.
- Sunde, R.A.; Paterson, E.; Evenson, J.K.; Barnes, K.M.; Lovegrove, J.A.; Gordon, M.H. Longitudinal selenium status in healthy British adults: Assessment using biochemical and molecular biomarkers. *Br. J. Nutr.* 2008, 99, S37–S47. [CrossRef]
- González, S.; Huerta, J.M.; Fernández, S.; Patterson, D.M.; Lasheras, C. Food intake and serum selenium concentration in elderly people. Ann. Nutr. Metab. 2006, 50, 126–131. [CrossRef] [PubMed]
- Jiménez-Redondo, S.; de Miguel, B.B.; Gómez-Pavón, J.; Vives, C.C. Non-institutionalized nonagenarians health-related quality of life and nutritional status: Is there a link between them? *Nutr. Hosp.* 2014, 30, 602–608. [PubMed]
- 220. Navia, B.; Ortega, R.M.; Perea, J.M.; Aparicio, A.; López-Sobaler, A.M.; Rodríguez-Rodríguez, E.; Research Group: UCM 920030 (VALORNUT). Selenium status in a group of schoolchildren from the region of M adrid, S pain. J. Hum. Nutr. Diet. 2014, 27, 239–246. [CrossRef] [PubMed]
- Rivas, A.; Romero, A.; Mariscal-Arcas, M.; Monteagudo, C.; López, G.; Ocaña-Peinado, F.M.; Olea-Serrano, F. Association between dietary antioxidant quality score (DAQs) and bone mineral density in Spanish women. *Nutr. Hosp.* 2012, 27, 1886–1893. [PubMed]
- 222. Golubkina, N.A. Selenium accumulation by cereals in Russia. Russ. Agric. Sci. 2007, 33, 288–291. [CrossRef]
- 223. Retondario, A.; Souza, A.D.M.; Fernandes, R.; Bricarello, L.P.; Alves, M.D.A.; Zeni, L.A.R.; Trindade, E.B.D.M.; Vasconcelos, F.D.A.G.D. Usual intake and dietary sources of Selenium in adolescents: A cross-sectional school-based study. *Clin. Nutr. ESPEN* 2019, 33, 91–97. [CrossRef] [PubMed]
- 224. Dumont, E.; Vanhaecke, F.; Cornelis, R. Selenium speciation from food source to metabolites: A critical review. *Anal. Bioanal. Chem.* **2006**, *385*, 1304–1323. [CrossRef] [PubMed]

- 225. Rayman, M.P. The importance of selenium to human health. Lancet 2000, 356, 233–241. [CrossRef]
- 226. Laclaustra, M.; Navas-Acien, A.; Stranges, S.; Ordovas, J.; Guallar, E. Serum Selenium Concentrations and Diabetes in U.S. Adults: National Health and Nutrition Examination Survey (NHANES) 2003–2004. *Environ. Health Perspect.* 2009, 117, 1409–1413. [CrossRef]
- 227. Laclaustra, M.; Stranges, S.; Navas-Acien, A.; Ordovas, J.; Guallar, E. Serum selenium and serum lipids in US adults: National Health and Nutrition Examination Survey (NHANES) 2003–2004. *Atherosclerosis* **2010**, *210*, 643–648. [CrossRef]
- 228. Tinggi, U. Essentiality and toxicity of selenium and its status in Australia: A review. Toxicol. Lett. 2002, 137, 103–110. [CrossRef]
- 229. Foster, L.H.; Sumar, S. Selenium in health and disease: A review. Crit. Rev. Food Sci. Nutr. 1997, 37, 211–228. [CrossRef]
- Arikan, D.C.; Coskun, A.; Ozer, A.; Kilinc, M.; Atalay, F.; Arikan, T. Plasma Selenium, Zinc, Copper and Lipid Levels in Postmenopausal Turkish Women and Their Relation with Osteoporosis. *Biol. Trace Element Res.* 2011, 144, 407–417. [CrossRef]
- 231. Aydemir, B.; Akdemir, R.; Vatan, M.B.; Cinemre, F.B.; Cinemre, H.; Kiziler, A.R.; Bahtiyar, N.; Buyukokuroglu, M.E.; Gurol, G.; Ogut, S. The Circulating Levels of Selenium, Zinc, Midkine, Some Inflammatory Cytokines, and Angiogenic Factors in Mitral Chordae Tendineae Rupture. *Biol. Trace Element Res.* 2015, 167, 179–186. [CrossRef] [PubMed]
- 232. Bay, A.; Dogan, M.; Bulan, K.; Kaba, S.; Demir, N.; Öner, A.F. A study on the effects of pica and iron-deficiency anemia on oxidative stress, antioxidant capacity and trace elements. *Hum. Exp. Toxicol.* **2013**, *32*, 895–903. [CrossRef] [PubMed]
- Coskun, A.; Arikan, T.; Kilinc, M.; Arikan, D.C.; Ekerbiçer, H. Plasma selenium levels in Turkish women with polycystic ovary syndrome. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 2013, 168, 183–186. [CrossRef]
- Erkekoğlu, P.; Aşçı, A.; Ceyhan, M.; Kızılgün, M.; Schweizer, U.; Ataş, C.; Koçer-Giray, B. Selenium levels, selenoenzyme activities and oxidant/antioxidant parameters in H1N1-infected children. *Turk. J. Pediatr.* 2013, 55, 271–282. [PubMed]
- 235. Eroglu, M.; Sahin, S.; Durukan, B.; Ozakpinar, O.B.; Erdinc, N.; Turkgeldi, L.; Sofuoglu, K.; Karateke, A. Blood Serum and Seminal Plasma Selenium, Total Antioxidant Capacity and Coenzyme Q10 Levels in Relation to Semen Parameters in Men with Idiopathic Infertility. *Biol. Trace Element Res.* 2014, 159, 46–51. [CrossRef]
- 236. Hıncal, F. Trace elements in growth: Iodine and selenium status of Turkish children. J. Trace Elements Med. Biol. 2007, 21, 40–43. [CrossRef]
- Kilinc, M.; Guven, M.A.; Ezer, M.; Ertas, I.E.; Coskun, A. Evaluation of Serum Selenium Levels in Turkish Women with Gestational Diabetes Mellitus, Glucose Intolerants, and Normal Controls. *Biol. Trace Element Res.* 2008, 123, 35–40. [CrossRef]
- Sakız, D.; Kaya, A.; Kulaksizoglu, M. Serum selenium levels in euthyroid nodular thyroid diseases. *Biol. Trace Elem. Res.* 2016, 174, 21–26. [CrossRef]
- Seven, M.; Basaran, S.Y.; Cengiz, M.; Unal, S.; Yuksel, A. Deficiency of selenium and zinc as a causative factor for idiopathic intractable epilepsy. *Epilepsy Res.* 2013, 104, 35–39. [CrossRef]
- Kadrabová, J.; Mad'arič, A.; Ginter, E. Determination of the daily selenium intake in Slovakia. *Biol. Trace Elem. Res.* 1998, 61, 277–286.
 [CrossRef]
- 241. Al-Ahmary, K.M. Selenium content in selected foods from the Saudi Arabia market and estimation of the daily intake. *Arab. J. Chem.* 2009, *2*, 95–99. [CrossRef]
- Mansour, A.; Ahadi, Z.; Qorbani, M.; Hosseini, S. Association between dietary intake and seasonal variations in postmenopausal women. J. Diabetes Metab. Disord. 2014, 13, 52. [CrossRef]
- Kieliszek, M.; Błażejak, S. Current Knowledge on the Importance of Selenium in Food for Living Organisms: A Review. *Molecules* 2016, 21, 609. [CrossRef] [PubMed]
- 244. Wasowicz, W.; Gromadzinska, J.; Rydzynski, K.; Tomczak, J. Selenium status of low-selenium area residents: Polish experience. *Toxicol. Lett.* **2002**, *137*, 95–101. [CrossRef]
- 245. Surai, P.F. Selenium in Nutrition and Health; Nottingham University Press: Nottingham, UK, 2006; Volume 974.
- Smrkolj, P.; Pograjc, L.; Hlastan-Ribič, C.; Stibilj, V. Selenium content in selected Slovenian foodstuffs and estimated daily intakes of selenium. *Food Chem.* 2005, 90, 691–697. [CrossRef]
- 247. Arafa, A.M.; Waly, M.; Jriesat, S.; Al Khafajei, A.; Sallam, S. Dietary and lifestyle characteristics of colorectal cancer in Jordan: A case-control study. *Asian Pac. J. Cancer Prev.* 2011, 12, 1931–1936. [PubMed]
- 248. Hansen, J.C.; Pedersen, H.S. Environmental exposure to heavy metals in North Greenland. Arct. Med. Res. 1986, 41, 21–34.
- Li, S.; Xiao, T.; Zheng, B. Medical geology of arsenic, selenium and thallium in China. *Sci. Total Environ.* 2012, 421, 31–40. [CrossRef]
- Zachara, B.A. Selenium and Selenium-Dependent Antioxidants in Chronic Kidney Disease. Adv. Clin. Chem. 2015, 68, 131–151. [CrossRef] [PubMed]
- Wang, N.; Tan, H.-Y.; Li, S.; Xu, Y.; Guo, W.; Feng, Y. Supplementation of Micronutrient Selenium in Metabolic Diseases: Its Role as an Antioxidant. Oxidative Med. Cell. Longev. 2017, 2017, 7478523. [CrossRef]
- Keshan, D.R.G.O.T.C.A.O.M.S.B.; Antiepidemic, S.O.S.P.C.; Antiepidemic, S.O.X.D.S.; Antiepidemic, S.O.M.C.S. Observations on effect of sodium selenite in prevention of Keshan disease. *Chin. Med. J.* 1979, 92, 471–476.
- Papp, L.V.; Holmgren, A.; Khanna, K.K. Selenium and Selenoproteins in Health and Disease. *Antioxid. Redox Signal.* 2010, 12, 793–795.
 [CrossRef] [PubMed]
- Albert Christophersen, O.; Haug, A. Possible roles of oxidative stress, local circulatory failure and nutrition factors in the pathogenesis of hypervirulent influenza: Implications for therapy and global emergency preparedness. *Microb. Ecol. Health Dis.* 2005, 17, 189–199. [CrossRef]

- 255. Cengiz, B.; Söylemez, F.; Öztürk, E.; Çavdar, A.O. Serum zinc, selenium, copper, and lead levels in women with second-trimester induced abortion resulting from neural tube defects: A preliminary study. *Biol. Trace Elem. Res.* 2004, 97, 225–235. [CrossRef] [PubMed]
- Ruder, E.H.; Hartman, T.J.; Blumberg, J.; Goldman, M.B. Oxidative stress and antioxidants: Exposure and impact on female fertility. *Hum. Reprod. Updat.* 2008, 14, 345–357. [CrossRef]
- Ruder, E.H.; Hartman, T.J.; Goldman, M.B. Impact of oxidative stress on female fertility. *Curr. Opin. Obstet. Gynecol.* 2009, 21, 219–222. [CrossRef]
- 258. Frost, D.V.; Ingvoldstad, D. Ecological aspects of selenium and tellurium in human and animal health. Chem. Scr. 1975, 8, 96–107.
- 259. Schrauzer, G.; White, D.; Schneider, C. Cancer mortality correlation studies-III: Statistical associations with dietary selenium intakes. *Bioinorg. Chem.* **1977**, *7*, 23–34. [CrossRef]
- 260. Hofmarcher, T.; Lindgren, P.; Wilking, N.; Jönsson, B. The cost of cancer in Europe 2018. Eur. J. Cancer 2020, 129, 41–49. [CrossRef]
- 261. Ip, C. Lessons from Basic Research in Selenium and Cancer Prevention. J. Nutr. **1998**, 128, 1845–1854. [CrossRef]
- 262. Hossain, A.; Skalicky, M.; Brestic, M.; Maitra, S.; Sarkar, S.; Ahmad, Z.; Vemuri, H.; Garai, S.; Mondal, M.; Bhatt, R.; et al. Selenium Biofortification: Roles, Mechanisms, Responses and Prospects. *Molecules* 2021, 26, 881. [CrossRef] [PubMed]
- Bentley-Hewitt, K.L.; Chen, R.K.Y.; Lill, R.E.; Hedderley, D.I.; Herath, T.D.; Matich, A.J.; McKenzie, M.J. Consumption of selenium-enriched broccoli increases cytokine production in human peripheral blood mononuclear cells stimulated ex vivo, a preliminary human intervention study. *Mol. Nutr. Food Res.* 2014, 58, 2350–2357. [CrossRef] [PubMed]
- Kuehnelt, D.; Juresa, D.; Francesconi, K.A.; Fakih, M.; Reid, M.E. Selenium metabolites in urine of cancer patients receiving l-selenomethionine at high doses. *Toxicol. Appl. Pharmacol.* 2007, 220, 211–215. [CrossRef]
- Zeng, H.; Combs, G.F. Selenium as an anticancer nutrient: Roles in cell proliferation and tumor cell invasion. *J. Nutr. Biochem.* 2008, 19, 1–7. [CrossRef]
- 266. Stranges, S.; Galletti, F.; Farinaro, E.; D'Elia, L.; Russo, O.; Iacone, R.; Capasso, C.; Carginale, V.; De Luca, V.; Della Valle, E.; et al. Associations of selenium status with cardiometabolic risk factors: An 8-year follow-up analysis of the Olivetti Heart Study. *Atherosclerosis* 2011, 217, 274–278. [CrossRef] [PubMed]
- Navarro-Alarcon, M.; López-Martínez, M. Essentiality of selenium in the human body: Relationship with different diseases. *Sci. Total. Environ.* 2000, 249, 347–371. [CrossRef]
- Cao, S.; Durrani, F.A.; Rustum, Y.M. Selective Modulation of the Therapeutic Efficacy of Anticancer Drugs by Selenium Containing Compounds against Human Tumor Xenografts. *Clin. Cancer Res.* 2004, 10, 2561–2569. [CrossRef]
- Cao, S.; Durrani, A.F.; Tóth, K.; Rustum, Y.M. Se-methylselenocysteine offers selective protection against toxicity and potentiates the antitumour activity of anticancer drugs in preclinical animal models. *Br. J. Cancer* 2014, *110*, 1733–1743. [CrossRef]
- 270. Burbano, X.; Miguez-Burbano, M.J.; McCollister, K.; Zhang, G.; Rodriguez, A.; Ruiz, P.; Shor-Posner, G. Impact of a selenium chemoprevention clinical trial on hospital admissions of HIV-infected participants. *HIV Clin. Trials* **2002**, *3*, 483–491.
- Hurwitz, B.E.; Klaus, J.R.; Llabre, M.M.; Gonzalez, A.; Lawrence, P.J.; Maher, K.J.; Schneiderman, N. Suppression of human immunodeficiency virus type 1 viral load with selenium supplementation: A randomized controlled trial. *Arch. Intern. Med.* 2007, 167, 148–154. [CrossRef]
- 272. Kupka, R.; Mugusi, F.; Aboud, S.; Msamanga, I.G.; Finkelstein, J.L.; Spiegelman, D.; Fawzi, W.W. Randomized, double-blind, placebo-controlled trial of selenium supplements among HIV-infected pregnant women in Tanzania: Effects on maternal and child outcomes. Am. J. Clin. Nutr. 2008, 87, 1802–1808. [CrossRef] [PubMed]
- 273. Broome, C.S.; McArdle, F.; Kyle, J.A.; Andrews, F.; Lowe, N.M.; Hart, C.A.; Arthur, J.R.; Jackson, M.J. An increase in selenium intake improves immune function and poliovirus handling in adults with marginal selenium status. *Am. J. Clin. Nutr.* 2004, *80*, 154–162. [CrossRef]
- 274. Hu, M.; Fang, J.; Zhang, Y.; Wang, X.; Zhong, W.; Zhou, Z. Design and evaluation a kind of functional biomaterial for bone tissue engineering: Selenium/mesoporous bioactive glass nanospheres. J. Colloid Interface Sci. 2020, 579, 654–666. [CrossRef] [PubMed]
- 275. Tan, X.; Liao, L.; Wan, Y.-P.; Li, M.-X.; Chen, S.-H.; Mo, W.-J.; Zhao, Q.-L.; Huang, L.-F.; Zeng, G.-Q. Downregulation of seleniumbinding protein 1 is associated with poor prognosis in lung squamous cell carcinoma. *World J. Surg. Oncol.* 2016, 14, 1–8. [CrossRef]
- 276. Klein, E.A.; Thompson, I.; Tangen, C.M.; Lucia, M.S.; Goodman, P.; Minasian, L.M.; Ford, L.G.; Parnes, H.L.; Gaziano, J.M.; Karp, D.D.; et al. Vitamin E and the risk of prostate cancer: Updated results of the Selenium and Vitamin E Cancer Prevention Trial (SELECT). J. Clin. Oncol. 2012, 30, 7. [CrossRef]
- 277. Kora, A.J.; Rastogi, L. Bacteriogenic synthesis of selenium nanoparticles by *Escherichia coli* ATCC 35218 and its structural characterisation. *IET Nanobiotechnol.* 2016, *11*, 179–184. [CrossRef] [PubMed]
- 278. Deng, X.F.; Zhao, Z.Q.; Han, Z.Y.; Huang, L.Q.; Lv, C.H.; Zhang, Z.H.; Liu, X.W. Selenium uptake and fruit quality of pear (*Pyrus communis* L.) treated with foliar Se application. *J. Plant Nutr. Soil Sci.* **2019**, *182*, 637–646. [CrossRef]
- Ros, G.H.; van Rotterdam, A.M.D.; Bussink, D.W.; Bindraban, P.S. Selenium fertilization strategies for bio-fortification of food: An agro-ecosystem approach. *Plant Soil* 2016, 404, 99–112. [CrossRef]
- Chen, Q.; Shi, W.; Wang, X. Selenium Speciation and Distribution Characteristics in the Rhizosphere Soil of Rice (*Oryza sativa* L.) Seedlings. *Commun. Soil Sci. Plant Anal.* 2010, 41, 1411–1425. [CrossRef]
- Zhou, X.-B.; Shi, W.-M.; Zhang, L.-H. Iron plaque outside roots affects selenite uptake by rice seedlings (*Oryza sativa* L.) grown in solution culture. *Plant Soil* 2006, 290, 17–28. [CrossRef]

- Niazi, N.K.; Bibi, I.; Shahid, M.; Ok, Y.S.; Burton, E.D.; Wang, H.; Shaheen, S.M.; Rinklebe, J.; Lüttge, A. Arsenic removal by perilla leaf biochar in aqueous solutions and groundwater: An integrated spectroscopic and microscopic examination. *Environ. Pollut.* 2018, 232, 31–41. [CrossRef] [PubMed]
- Golob, A.; Gadžo, D.; Stibilj, V.; Djikić, M.; Gavrić, T.; Kreft, I.; Germ, M. Sulphur interferes with selenium accumulation in Tartary buckwheat plants. *Plant Physiol. Biochem.* 2016, 108, 32–36. [CrossRef] [PubMed]
- Naz, F.S.; Yusuf, M.; Khan, T.A.; Fariduddin, Q.; Ahmad, A. Low level of selenium increases the efficacy of 24-epibrassinolide through altered physiological and biochemical traits of Brassica juncea plants. *Food Chem.* 2015, 185, 441–448. [CrossRef]
- Mackowiak, C.L.; Amacher, M.C. Soil Sulfur Amendments Suppress Selenium Uptake by Alfalfa and Western Wheatgrass. J. Environ. Qual. 2008, 37, 772–779. [CrossRef]
- 286. Lyons, G.H.; Lewis, J.; Lorimer, M.F.; Holloway, R.E.; Brace, D.M.; Stangoulis, J.C.; Graham, R.D. High-selenium wheat: Agronomic biofortification strategies to improve human nutrition. *Food Agric. Environ.* **2004**, *2*, 171–178.
- Sors, T.G.; Ellis, D.R.; Salt, D.E. Selenium uptake, translocation, assimilation and metabolic fate in plants. *Photosynth. Res.* 2005, 86, 373–389. [CrossRef]
- Pierart, A.; Shahid, M.; Séjalon-Delmas, N.; Dumat, C. Antimony bioavailability: Knowledge and research perspectives for sustainable agricultures. J. Hazard. Mater. 2015, 289, 219–234. [CrossRef]
- Xing, K.; Zhou, S.; Wu, X.; Zhu, Y.; Kong, J.; Shao, T.; Tao, X. Concentrations and characteristics of selenium in soil samples from Dashan Region, a selenium-enriched area in China. *Soil Sci. Plant Nutr.* 2015, *61*, 889–897. [CrossRef]
- Munier-Lamy, C.; Deneux-Mustin, S.; Mustin, C.; Merlet, D.; Berthelin, J.; Leyval, C. Selenium bioavailability and uptake as affected by four different plants in a loamy clay soil with particular attention to mycorrhizae inoculated ryegrass. *J. Environ. Radioact.* 2007, 97, 148–158. [CrossRef]
- 291. Johnsson, L. Selenium uptake by plants as a function of soil type, organic matter content and pH. *Plant Soil* **1991**, *133*, 57–64. [CrossRef]
- 292. White, P.J. Selenium metabolism in plants. Biochim. Et Biophys. Acta BBA-Gen. Subj. 2018, 1862, 2333–2342. [CrossRef] [PubMed]
- Jacobs, L.W. Selenium in Agriculture and the Environment; Soil Science Society of America Special Publication 23; SSSA: Madison, WI, USA, 1989.
- 294. Elrashidi, M.A.; Adriano, D.C.; Workman, S.M.; Lindsay, W.L. Chemical equilibria of selenium in soils: A theoretical development: 1. *Soil Sci.* **1987**, *144*, 141–152. [CrossRef]
- 295. Barker, A.V.; Pilbeam, D.J. (Eds.) Handbook of Plant Nutrition; CRC Press: Boca Raton, FL, USA, 2015.
- 296. Wang, C.; Ji, J.; Zhu, F. Characterizing Se transfer in the soil-crop systems under field condition. *Plant Soil* **2017**, *415*, 535–548. [CrossRef]
- 297. Borawska, M.H.; Witkowska, A.M.; Hukałowicz, K.; Markiewicz, R. Influence of dietary habits on serum selenium concentration. *Ann. Nutr. Metab.* **2004**, *48*, 134–140. [CrossRef] [PubMed]
- Dong, Z.; Liu, Y.; Dong, G.; Wu, H. Effect of boiling and frying on the selenium content, speciation, and in vitro bioaccessibility of selenium-biofortified potato (*Solanum tuberosum* L.). *Food Chem.* 2021, 348, 129150. [CrossRef]
- Pérez, M.B.; Maniero, M.; Londonio, A.; Smichowski, P.; Wuilloud, R.G. Effects of common cooking heat treatments on selenium content and speciation in garlic. J. Food Compos. Anal. 2018, 70, 54–62. [CrossRef]
- Khanam, A.; Platel, K. Influence of domestic processing on the bioaccessibility of selenium from selected food grains and composite meals. J. Food Sci. Technol. 2015, 53, 1634–1639. [CrossRef]
- Lu, X.; He, Z.; Lin, Z.; Zhu, Y.; Yuan, L.; Liu, Y.; Yin, X. Effects of Chinese Cooking Methods on the Content and Speciation of Selenium in Selenium Bio-Fortified Cereals and Soybeans. *Nutrients* 2018, 10, 317. [CrossRef]
- Moreda-Piñeiro, J.; Moreda-Piñeiro, A.; Bermejo-Barrera, P. In vivo and in vitro testing for selenium and selenium compounds bioavailability assessment in foodstuff. *Crit. Rev. Food Sci. Nutr.* 2015, 57, 805–833. [CrossRef]
- 303. Wang, C.; Duan, H.-Y.; Teng, J.-W. Assessment of Microwave Cooking on the Bioaccessibility of Cadmium from Various Food Matrices Using an In Vitro Digestion Model. *Biol. Trace Element Res.* 2014, 160, 276–284. [CrossRef]
- 304. Matos, J.; Lourenço, H.M.; Brito, P.; Maulvault, A.L.; Martins, L.L.; Afonso, C. Influence of bioaccessibility of total mercury, methyl-mercury and selenium on the risk/benefit associated to the consumption of raw and cooked blue shark (*Prionace glauca*). *Environ. Res.* 2015, 143, 123–129. [CrossRef] [PubMed]
- 305. Raisbeck, M.F. Selenosis in ruminants. Vet. Clin. Food Anim. Pract. 2020, 36, 775–789. [CrossRef] [PubMed]
- 306. Molnár, Á.; Kolbert, Z.; Kéri, K.; Feigl, G.; Ördög, A.; Szőllősi, R.; Erdei, L. Selenite-induced nitro-oxidative stress processes in Arabidopsis thaliana and Brassica juncea. *Ecotoxicol. Environ. Saf.* 2018, 148, 664–674. [CrossRef]
- 307. Moreno, O.D.G.; Aguilar, F.J.A.; Barrientos, E.Y. Selenium Uptake and Biotransformation and Effect of Selenium Exposure on the Essential and Trace Elements Status: Comparative Evaluation of Four Edible Plants. J. Mex. Chem. Soc. 2018, 62, 247–258. [CrossRef]
- Lehotai, N.; Lyubenova, L.; Schröder, P.; Feigl, G.; Ördög, A.; Szilágyi, K.; Kolbert, Z. Nitro-oxidative stress contributes to selenite toxicity in pea (Pisum sativum L). *Plant Soil* 2016, 400, 107–122. [CrossRef]
- Hawrylak-Nowak, B. Comparative effects of selenite and selenate on growth and selenium accumulation in lettuce plants under hydroponic conditions. *Plant Growth Regul.* 2013, 70, 149–157. [CrossRef]

- 310. Łabanowska, M.; Filek, M.; Kościelniak, J.; Kurdziel, M.; Kuliś, E.; Hartikainen, H. The effects of short-term selenium stress on Polish and Finnish wheat seedlings—EPR, enzymatic and fluorescence studies. J. Plant Physiol. 2012, 169, 275–284. [CrossRef] [PubMed]
- 311. Lehotai, N.; Kolbert, Z.; Pető, A.; Feigl, G.; Ördög, A.; Kumar, D.; Erdei, L. Selenite-induced hormonal and signalling mechanisms during root growth of Arabidopsis thaliana L. J. Exp. Bot. 2012, 63, 5677–5687. [CrossRef] [PubMed]
- Saffaryazdi, A.; Lahouti, M.; Ganjeali, A.; Bayat, H. Impact of Selenium Supplementation on Growth and Selenium Accumulation on Spinach (*Spinacia oleracea* L.) Plants. Not. Sci. Biol. 2012, 4, 95–100. [CrossRef]
- 313. Freeman, J.L.; Tamaoki, M.; Stushnoff, C.; Quinn, C.F.; Cappa, J.J.; Devonshire, J.; Fakra, S.C.; Marcus, M.A.; McGrath, S.P.; Van Hoewyk, D.; et al. Molecular Mechanisms of Selenium Tolerance and Hyperaccumulation in *Stanleya pinnata*. *Plant Physiol.* 2010, 153, 1630–1652. [CrossRef]
- 314. Feng, R.; Wei, C.; Tu, S.; Wu, F. Effects of Se on the uptake of essential elements in Pteris vittata L. *Plant Soil* **2009**, 325, 123–132. [CrossRef]
- 315. Ramos, S.J.; Faquin, V.; Almeida, H.J.D.; Ávila, F.W.; Guilherme, L.R.G.; Bastos, C.E.A.; Ávila, P.A. Selenato e selenito na produção, nutrição mineral e biofortificação com selênio em cultivares de alface¹. *Rev. Bras. Ciência Solo* **2011**, *35*, 1347–1355. [CrossRef]
- 316. Zhang, M.; Tang, S.; Huang, X.; Zhang, F.; Pang, Y.; Huang, Q.; Yi, Q. Selenium uptake, dynamic changes in selenium content and its influence on photosynthesis and chlorophyll fluorescence in rice (*Oryza sativa* L.). *Environ. Exp. Bot.* 2014, 107, 39–45. [CrossRef]
- Matraszek, R.; Hawrylak-Nowak, B. Macronutrients accumulation in useable parts of lettuce as affected by nickel and selenium concentrations in nutrient solution. *Fresenius Environ. Bull.* 2009, 18, 1059–1065.
- 318. Sarwar, N.; Akhtar, M.; Kamran, M.A.; Imran, M.; Riaz, M.A.; Kamran, K.; Hussain, S. Selenium biofortification in food crops: Key mechanisms and future perspectives. *J. Food Compos. Anal.* **2020**, *93*, 103615. [CrossRef]
- 319. Bouis, H.E.; Chassy, B.M.; Ochanda, J.O. 2. Genetically modified food crops and their contribution to human nutrition and food quality. *Trends Food Sci. Technol.* 2003, 14, 191–209. [CrossRef]

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