

Supplementary Materials

# Plasmacytoid Dendritic Cell, Slan<sup>+</sup>-Monocyte and Natural Killer Cell Counts Function as Blood Cell-Based Biomarkers for Predicting Responses to Immune Checkpoint Inhibitor Monotherapy in Non-Small Cell Lung Cancer Patients

Francesca Pettinella <sup>1</sup>, Chiara Lattanzi <sup>1</sup>, Marta Donini <sup>1</sup>, Elena Cavegion <sup>1</sup>, Olivia Marini <sup>1</sup>, Giulia Iannoto <sup>1</sup>, Sara Costa <sup>1</sup>, Elena Zenaro <sup>1</sup>, Tiago Moderno Fortunato <sup>1</sup>, Sara Gasperini <sup>1</sup>, Matteo Giani <sup>1</sup>, Lorenzo Belluomini <sup>2</sup>, Marco Sposito <sup>2</sup>, Jessica Insolda <sup>2</sup>, Iliaria Mariangela Scaglione <sup>2</sup>, Michele Milella <sup>2</sup>, Annalisa Adamo <sup>3</sup>, Ornella Poffe <sup>3</sup>, Vincenzo Bronte <sup>4</sup>, Stefano Dusi <sup>1</sup>, Marco A. Cassatella <sup>1</sup>, Stefano Ugel <sup>3</sup>, Sara Pilotto <sup>2</sup> and Patrizia Scapini <sup>1,\*</sup>

<sup>1</sup> General Pathology Section, Department of Medicine, University of Verona, 37134 Verona, Italy

<sup>2</sup> Section of Innovation Biomedicine—Oncology Area, Department of Engineering for Innovation Medicine (DIMI), University of Verona and University and Hospital Trust (AOUI) of Verona, 37134 Verona, Italy

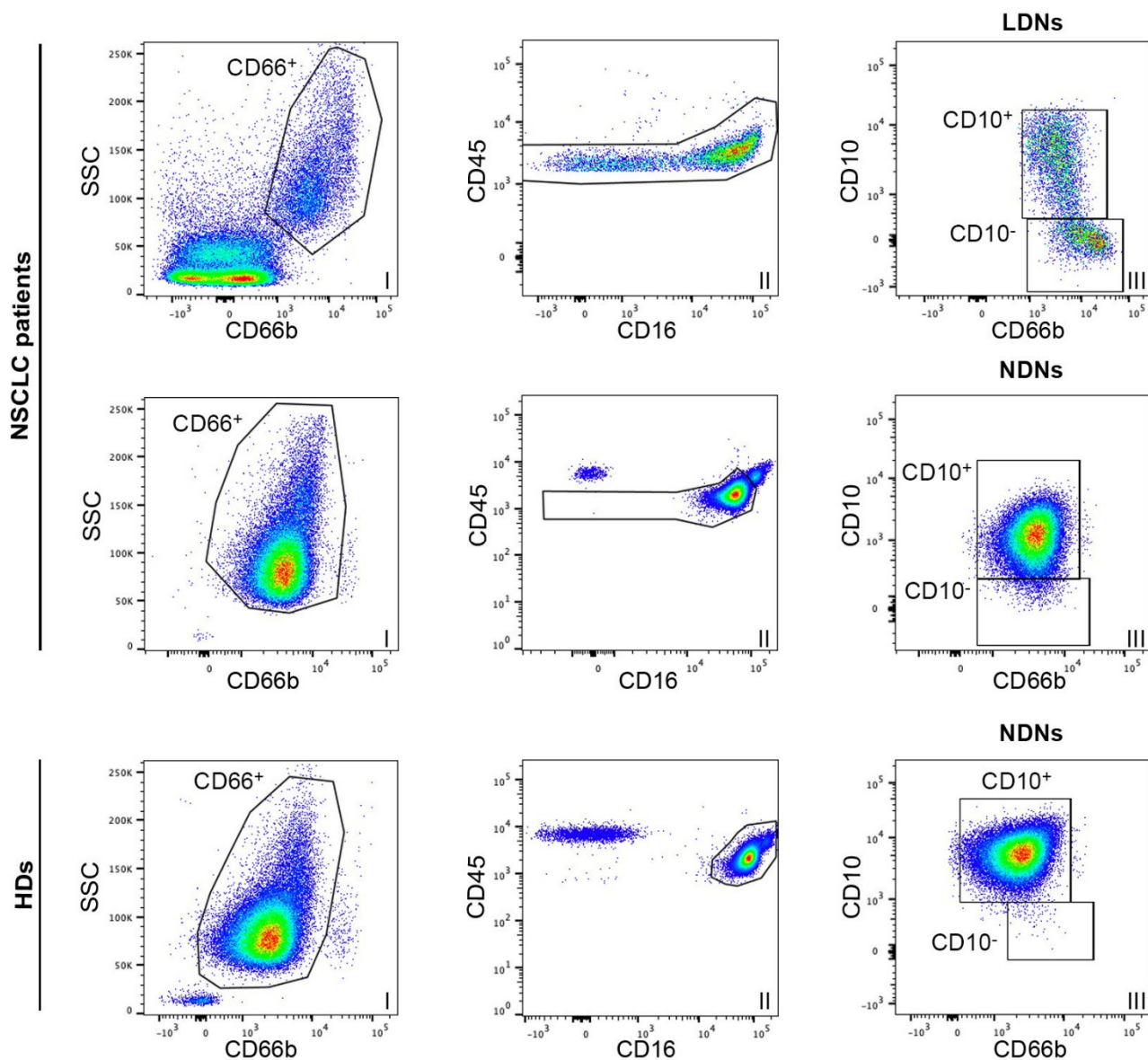
<sup>3</sup> Immunology Section, Department of Medicine, University and Hospital Trust (AOUI) of Verona, 37134 Verona, Italy

<sup>4</sup> Veneto Institute of Oncology—Istituto di Ricovero e Cura a Carattere Scientifico (IOV-IRCCS), 35128 Padova, Italy

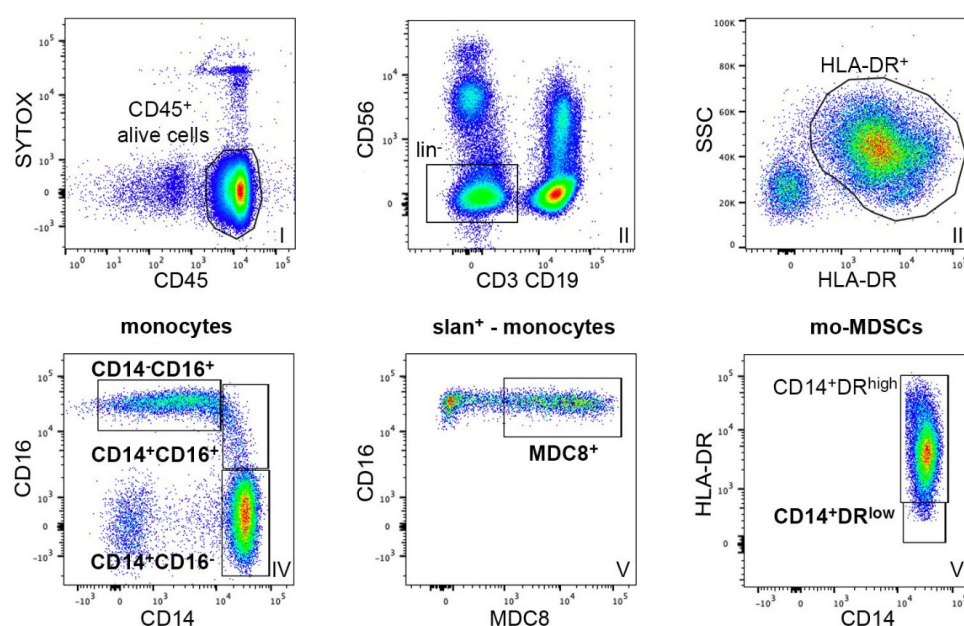
\* Correspondence: patrizia.scapini@univr.it; Tel.: +39-045-8027-556; Fax: +39-045-8027-127

**Table S1.** Fluorochrome-conjugated antibodies for flow cytometry.

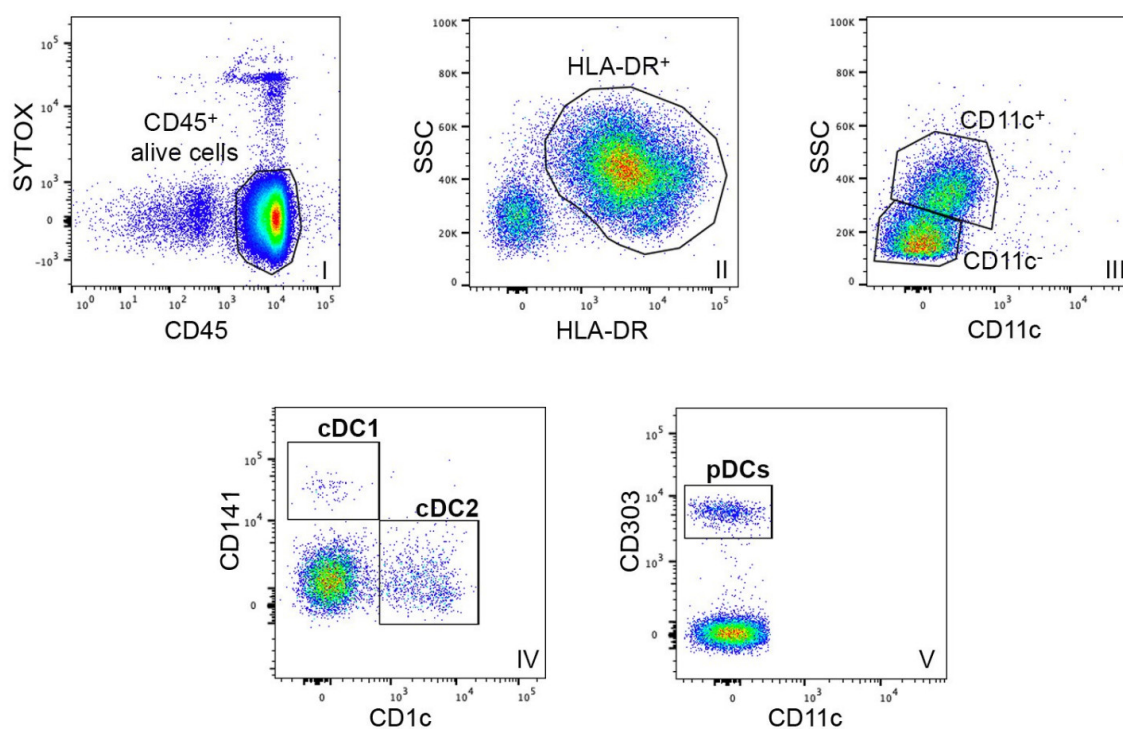
| Target        | Clone  | Label           | Source          | Identifier                         |
|---------------|--------|-----------------|-----------------|------------------------------------|
| CD56          | REA196 | PE              | Miltenyi Biotec | Cat# 130-112-312, RRID: AB_2726090 |
| CD45          | HI30   | BV510           | BioLegend       | Cat# 304036, RRID: AB_2561940      |
|               | 5B1    | VioGreen        | Miltenyi Biotec | Cat# 130-113-124, RRID: AB_2725952 |
| CD66b         | G10F5  | FITC            | BioLegend       | Cat# 305104, RRID: AB_314496       |
|               | G10F5  | APC             | BioLegend       | Cat# 305118, RRID: AB_2566607      |
|               | G10F5  | PerCP-Cy5.5     | BioLegend       | Cat# 305108, RRID: AB_2077855      |
|               | 6/40c  | BV421           | BioLegend       | Cat# 392916, RRID: AB_2888722      |
| CD11b         | ICRF44 | PE-Cy7          | BioLegend       | Cat# 301322, RRID: AB_830644       |
|               | ICRF44 | APC-Cy7         | BD Bioscience   | Cat# 557754, RRID: AB_396860       |
| CD16          | 3G8    | APC-Cy7         | BioLegend       | Cat# 301018, RRID: AB_314218       |
| CD16          | 3G8    | PE-Cy7          | BD Bioscience   | Cat# 557744, RRID: AB_396850       |
| CD10          | HI10a  | PE              | BioLegend       | Cat# 312204, RRID: AB_314915       |
|               | HI10a  | APC             | BioLegend       | Cat# 312210, RRID: AB_314921       |
| CD62L         | 145/15 | APC             | Miltenyi Biotec | Cat# 130-113-617, RRID: AB_2733392 |
| CD14          | TÜK4   | VioBlue         | Miltenyi Biotec | Cat# 130-113-152, RRID: AB_2725980 |
| CD11c         | 3.9    | PE              | eBioscience     | Cat# 12-0116-42, RRID: AB_10597432 |
| CD1c          | REA694 | Vio Bright FITC | Miltenyi Biotec | Cat# 130-110-598, RRID: AB_2656035 |
| CD303         | AC144  | FITC            | Miltenyi Biotec | Cat# 130-113-192, RRID: AB_2726017 |
| CD141         | REA674 | PE-Vio770       | Miltenyi Biotec | Cat# 130-113-663, RRID: AB_2751169 |
| CD3           | REA613 | PE-Vio770       | Miltenyi Biotec | Cat# 130-113-140, RRID: AB_2725968 |
| CD19          | HIB19  | PE-Cy7          | BioLegend       | Cat# 302216, RRID: AB_314246       |
| M-DC8         | DD-1   | FITC            | Miltenyi Biotec | Cat# 130-117-371, RRID: AB_2733608 |
| CD274 / PD-L1 | MIH1   | BV421           | BD Bioscience   | Cat# 563738, RRID: AB_2738396      |
| HLA-DR        | L243   | APC             | BioLegend       | Cat# 307610, RRID: AB_314688       |
|               | L243   | APC-Cy7         | BioLegend       | Cat# 307618, RRID: AB_493586       |



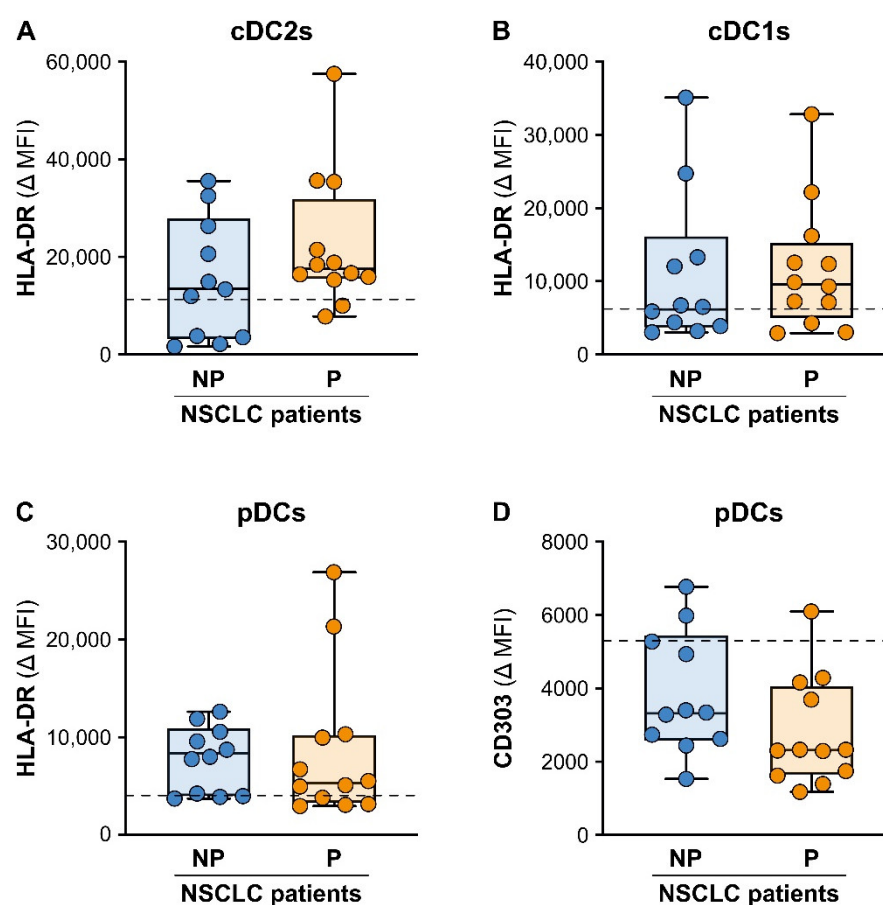
**Figure S1. Identification of CD66b<sup>+</sup>LDNs and CD66b<sup>+</sup>NDNs from advanced NSCLC patients and HDs by flow cytometry.** After gating on singlets, excluding debris and gating on CD45<sup>+</sup>Sytox<sup>-</sup> cells (not shown), CD66b<sup>+</sup>CD10<sup>+/−</sup>CD16<sup>high</sup> LDNs/NDNs from NSCLC patients (top and middle panels) and CD66b<sup>+</sup>CD10<sup>+/−</sup>CD16<sup>high</sup> NDNs from HDs (bottom panels) were identified by gating on total CD66b<sup>+</sup> granulocytes (I), excluding CD45<sup>high</sup>CD16<sup>-</sup> eosinophils (II) and finally gating on CD10<sup>+/−</sup> LDNs/NDNs (III).



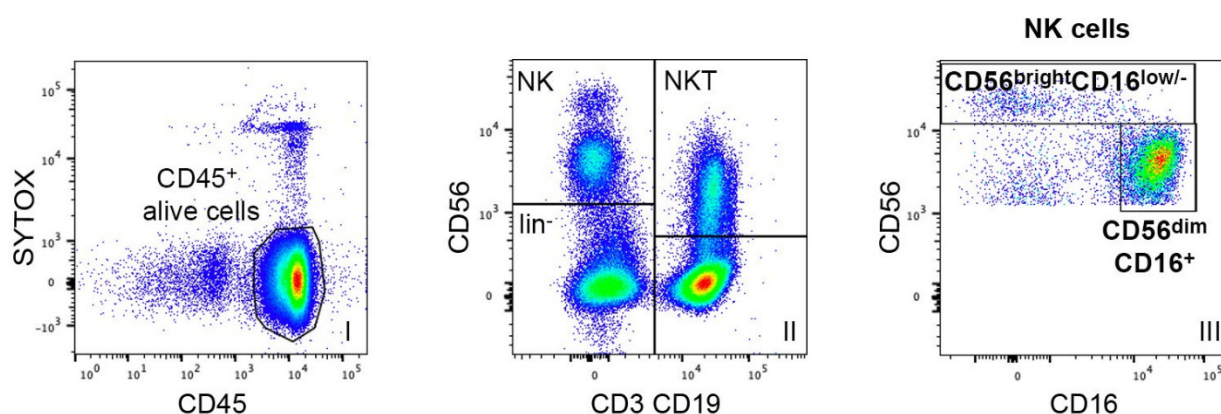
**Figure S2.** Characterization of CD14<sup>+</sup>CD16<sup>-</sup> classical, CD14<sup>+</sup>CD16<sup>-</sup> intermediate, and CD14<sup>dim</sup>/CD16<sup>+</sup> non-classical monocytes, slan (MDC8)<sup>-</sup> monocytes and CD14<sup>+</sup>HLA-DR<sup>low</sup> mo-MDSCs by flow cytometry. After gating on singlets (not shown), excluding debris (not shown), gating on CD45<sup>+</sup>Sytox<sup>-</sup> cells (I), gating on lin<sup>-</sup> (CD13-CD19-CD56<sup>-</sup>) cells (II), gating on HLA-DR<sup>+</sup> cells (III), CD14<sup>+</sup>CD16<sup>-</sup> classical, CD14<sup>+</sup>CD16<sup>-</sup> intermediate, and CD14<sup>dim</sup>/CD16<sup>+</sup> non-classical monocytes (IV), slan (MDC8)<sup>-</sup> monocytes (V) and CD14<sup>+</sup>HLA-DR<sup>low</sup> mo-MDSCs (VI) were identified as reported in the representative plots.



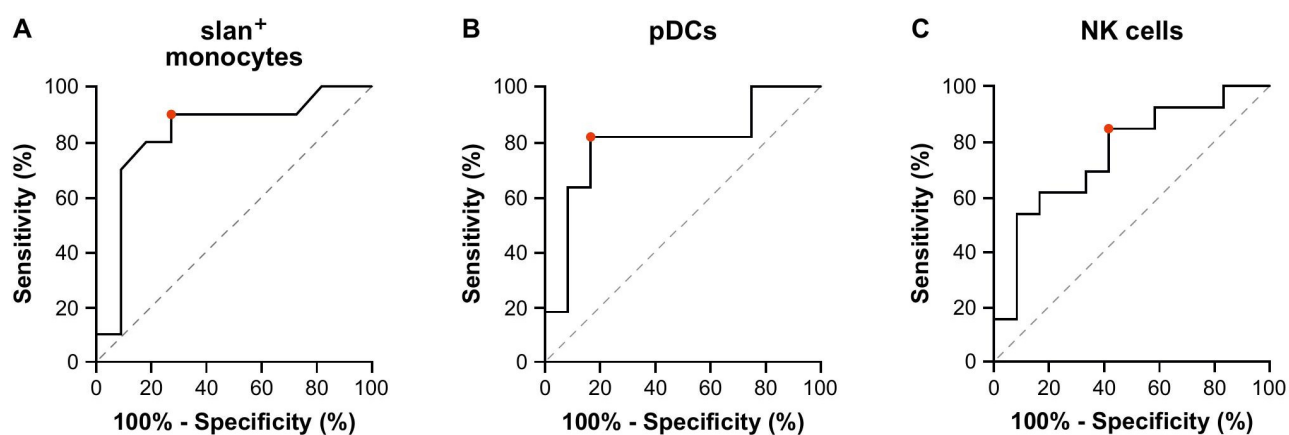
**Figure S3.** Characterization of cDC1s, cDC2s and pDCs by flow cytometry. After gating on singlets (not shown), excluding debris (not shown), gating on CD45<sup>+</sup>Sytox<sup>-</sup> cells (I), gating on HLA-DR<sup>+</sup> cells (II), cDC1s and cDC2s were identified as CD141<sup>+</sup> cells (cDC1s, IV) or CD1c<sup>+</sup> cells (cDC2, IV), respectively, within total CD11c<sup>high</sup> cells (III), while pDCs were identified as CD303<sup>+</sup> cells within total CD11c<sup>low</sup> cells (III, V).



**Figure S4.** Baseline HLA-DR and CD303 expression on circulating DC subpopulations from NP and P NSCLC patients undergoing ICI monotherapy. HLA-DR (A - C) and CD303 (D) expression levels were evaluated by flow cytometry on circulating CD141<sup>+</sup> cDC2s (A), CD1c<sup>+</sup> cDC1s (B) and CD303<sup>+</sup> pDCs (C, D) in NP (n = 11) or P (n = 12) NSCLC patients. The median HLA-DR (A - C) or CD303 (D)  $\Delta$  MFI value (25<sup>th</sup> - 75<sup>th</sup> percentile), calculated as described in Methods, for each DC population is reported. Each symbol stands for a single NSCLC patient sample. The median HLA-DR (A - C) or CD303 (D)  $\Delta$  MFI value for each DC population for reference HDs is reported as dashed line.



**Figure S5.** Characterization of total CD56<sup>+</sup>, CD56<sup>dim</sup>CD16<sup>+</sup> or CD56<sup>bright</sup>CD16<sup>-</sup> NK cells by flow cytometry. After gating on singlets (not shown), excluding debris (not shown), gating on CD45<sup>+</sup>Sytox<sup>-</sup> cells (I), total CD56<sup>+</sup> (II), CD56<sup>dim</sup>CD16<sup>+</sup> (III) or CD56<sup>bright</sup>CD16<sup>-</sup> (III) NK cells were identified as reported in the representative plots.



**Figure S6.** Receiver operating characteristic (ROC) curves to assess therapy responses based on baseline values of predictive biomarkers. (A - C) ROC curves for prediction of the therapy response based on baseline cell counts of slan<sup>+</sup>-monocytes (A), pDCs (B) and total NK cells (C) are shown. The optimal cut-off for each parameter (red dot) is based on trade-off between specificity and sensitivity.