

Supplementary Materials: Effects of Dihydrophaseic Acid 3'-O-β-D-Glucopyranoside Isolated from *Lycii radicis* Cortex on Osteoblast Differentiation

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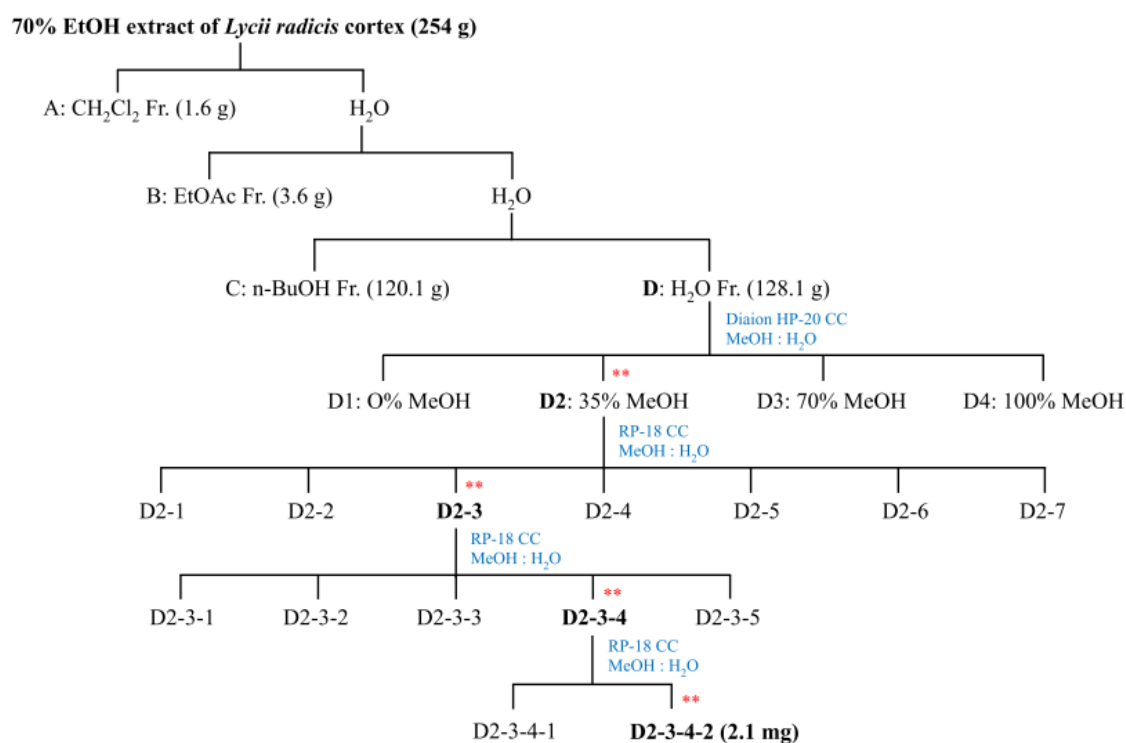


Figure S1. Fractionation and isolation of the bioactive component enhancing osteoblast differentiation from 70% ethanol extract of *Lycii radicis* cortex. Abbreviations: Fr., fraction; CC, column chromatography; EtOH, ethanol; CH₂Cl₂, dichloromethane; EtOAc, ethyl acetate; *n*-BuOH, *n*-butanol; MeOH, methanol. **: Bioactivity-guided fractionation.

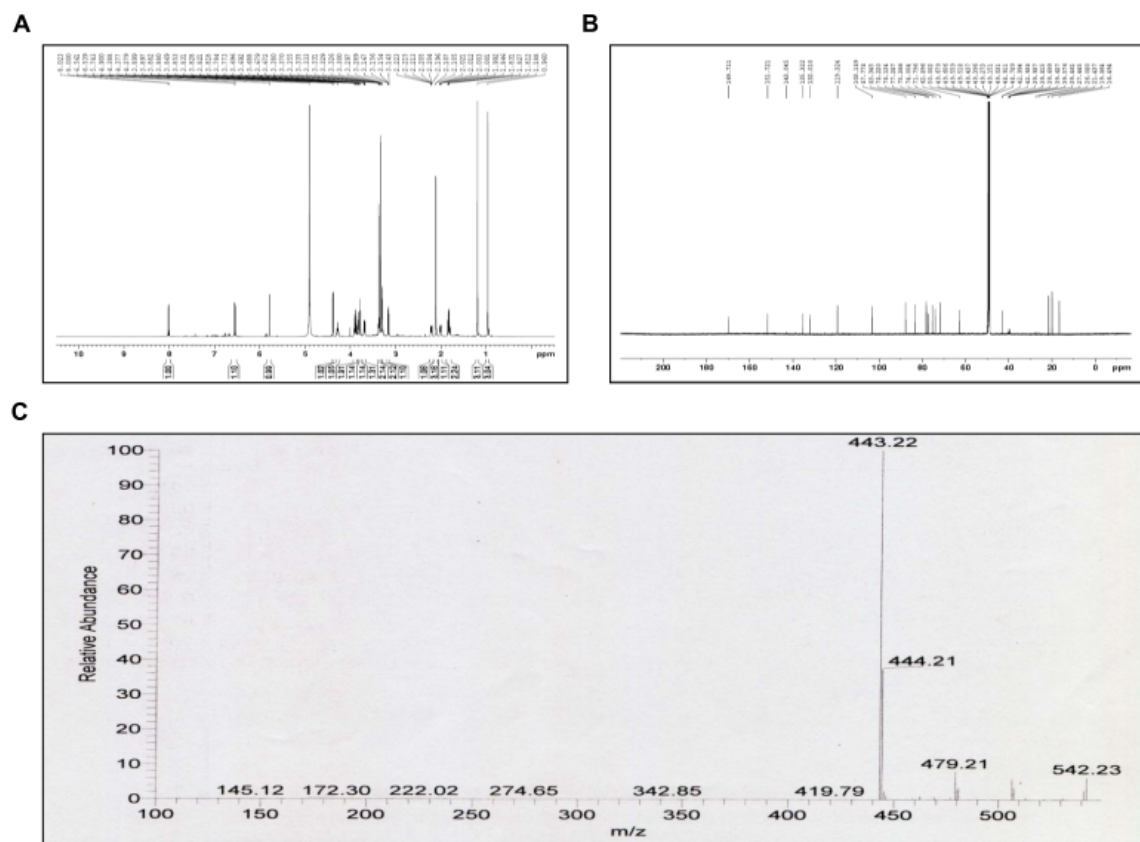


Figure S2. Proton nuclear magnetic resonance (^1H -NMR) (A), carbon-13 nuclear magnetic resonance (^{13}C -NMR) (B), and mass spectral (C) analyses of the D2-3-4-2 fraction of Supplementary Figure S1.

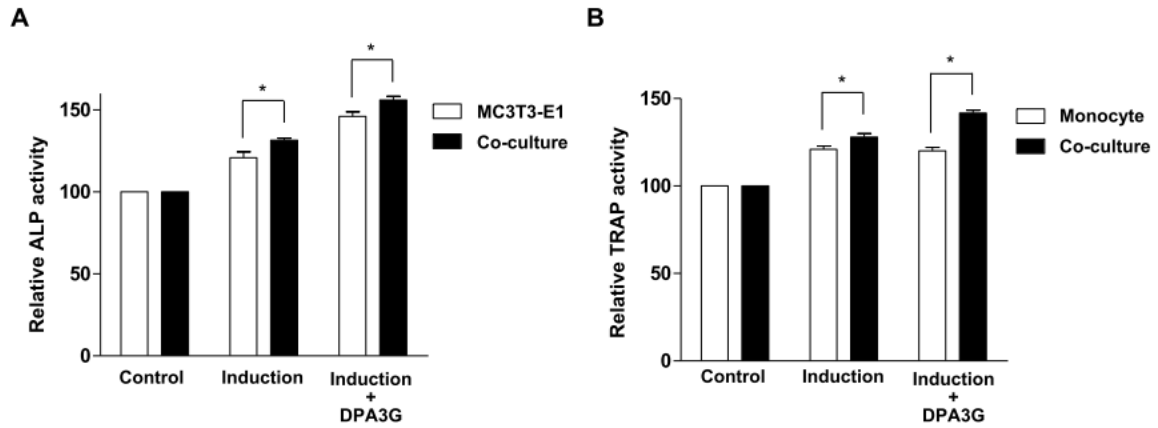


Figure S3. Effect of (1'R,2'S,5'R,8'S,2'Z,4'E)-dihydrophaseic acid 3'-O- β -D-glucopyranoside (DPA3G) on osteoblast and osteoclast differentiation in the co-culture of preosteoblasts and primary monocytes. MC3T3-E1 preosteoblasts and primary monocytes were cultured separately or co-cultured. In the separate culture, MC3T3-E1 and primary monocyte cells were treated with osteoblast differentiation reagents (50 μ g/mL ascorbic acid and 10 mM β -glycerophosphate) and osteoclast differentiation reagents (30 ng/mL of M-CSF and 50 ng/mL of RANKL), respectively, with DPA3G (Induction+DPA3G) or without (Induction). In co-culture, cells were treated with osteoblast differentiation reagents (50 μ g/mL ascorbic acid and 10 mM β -glycerophosphate) with DPA3G (Induction+DPA3G) or without (Induction). After 5 day culture, alkaline phosphatase (ALP) activity (A) and tartrate-resistant acid phosphatase (TRAP) activity (B) were assessed in each cell group. Control: non-induction of osteoblast or osteoclast differentiation. DPA3G: (1'R,2'S,5'R,8'S,2'Z,4'E)-dihydrophaseic acid 3'-O- β -D-glucopyranoside. *: $p < 0.05$.

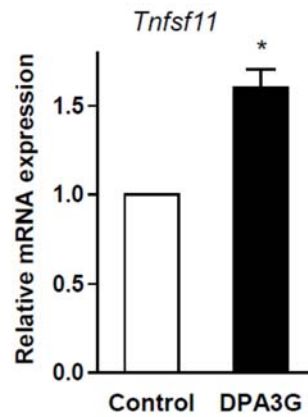


Figure S4. Effects of (1'R,2'S,5'R,8'S,2'Z,4'E)-dihydrophaseic acid 3'-O- β -D-glucopyranoside (DPA3G) on *Tnfsf11* (LANKL) mRNA expression in the co-culture of preosteoblasts and primary monocytes. MC3T3-E1 preosteoblast and primary monocyte cells were co-cultured for 1 day and then added with 50 μ g/ml of ascorbic acid and 10 mM of β -glycerophosphate for induction of osteoblast differentiation. Cells were treated with 5 μ g/ml of DPA3G fraction for 5 days and then total RNA of the cells was extracted. The mRNA expression level of *Tnfsf11* gene was assessed by quantitative reverse-transcription PCR and then normalized to *Gapdh* mRNA expression. Control: non-DPA3G-treated cells. *: $p < 0.05$ vs. Control.