

Review

The *Hedyotis diffusa* Willd. (Rubiaceae): A Review on Phytochemistry, Pharmacology, Quality Control and Pharmacokinetics

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Abstract: *Hedyotis diffusa* Willd (*H. diffusa*) is a well-known Chinese medicine with a variety of activities, especially its anti-cancer effect in the clinic. Up to now, 171 compounds have been reported from *H. diffusa*, including 32 iridoids, 26 flavonoids, 24 anthraquinones, 26 phenolics and their derivatives, 50 volatile oils and 13 miscellaneous compounds. *In vitro* and *in vivo* studies show these phytochemicals and plant extracts to exhibit a range of pharmacological activities of anti-cancer, antioxidant, anti-inflammatory, anti-fibroblast, immunomodulatory and neuroprotective effects. Although a series of methods have been established for the quality control of *H. diffusa*, a feasible and reliable approach is still needed in consideration of its botanical origin, collecting time and bioactive effects. Meanwhile, more pharmacokinetics researches are needed to illustrate the characteristics of *H. diffusa in vivo*. The present review aims to provide up-to-date and comprehensive information on the phytochemistry, pharmacology, quality control and pharmacokinetic characteristics of *H. diffusa* for its clinical use and further development.

Keywords: *H. diffusa*; phytochemistry; pharmacology; quality control; pharmacokinetics

1. Introduction

Hedyotis diffusa Willd (*H. diffusa*, Family Rubiaceae), known as *Oldenlandia diffusa* (Willd) Roxb, is a well-known Chinese medicine used for the treatment of inflammation-linked diseases, such as hepatitis, appendicitis and urethritis, for thousands of years in China [1]. In our previous studies, the water extract of *H. diffusa* has been proved to have an obvious protective effect in lipopolysaccharide-induced renal inflammation in mice. Recently, *H. diffusa* has gained increasing attention for its properties of anti-proliferative activity in cancer cells and anti-tumor activity in tumor-bearing animals [2–5]. It has been proved as the most commonly prescribed single Chinese herb used for colon cancer and breast cancer patients [6,7], according to the statistics from the National Health Insurance Research Database of Taiwan.

H. diffusa is an annual herb, widely distributed in the orient and tropical Asia, such as China, Japan and Indonesia [1,8]. Generally, the plant grows in humid fields and ridges of farmlands, ascending to procumbent, to 50 cm tall; the stem is slightly flattened to terete, glabrescent to glabrous and the papilla was observed in the transverse section of the stem; the leaves are opposite, sessile or subsessile and blade drying membranous, linear, narrowly elliptic, 1–4 × 0.1–0.4 cm; the flowers with pedicels are

pairs in axillary racemes and the corolla is white [1,9]. Together with these phenotypic characteristics of *H. diffusa*, methods of thin-layer chromatography (TLC) [10], gas chromatography-mass spectrometer (GC-MS) [11], high performance liquid chromatography (HPLC) [9] and DNA sequencing [8,12] have been developed to differentiate *H. diffusa* from related species (e.g., *Hedyotis corymbosa* (L.) Lam) to give the right prescription for illnesses.

Although there are numbers of published scientific literature on the chemical constituents, pharmacological activities and quantitative analysis of *H. diffusa*, a systematic and updated review is unavailable. Therefore, the aim of this review is to extensively summarize the phytochemistry, pharmacology, quality control and pharmacokinetic characteristics of *H. diffusa*, as well as being an evidence for clinical uses and further researches of this herb.

2. Phytochemistry

With the advancement of analysis technologies like mass spectrometer (MS), liquid chromatograph–mass spectrometer (LC-MS), nuclear magnetic resonance–mass spectrometer (NMR-MS) etc., many studies on *H. diffusa* revealed numbers of important phytochemicals, including iridoids, triterpenes, flavonoids, anthraquinones, phenolic acids and their derivatives, sterols, alkaloids, volatile oils, polysaccharides, cyclotides, coumarins and alkaloids. The detailed information for these compounds is summarized in Table 1.

Table 1. Compounds of the *H. diffusa*.

NO.	Compound Name	Molecular Formula	Reference
Iridoids			
1	Asperuloside	C ₁₈ H ₂₂ O ₁₁	[13,14]
2	Deacetyl asperuloside	C ₁₆ H ₂₀ O ₁₀	[15]
3	Asperuloside acid	C ₁₈ H ₂₄ O ₁₂	[16]
4	Deacetyl asperulosidic acid	C ₁₆ H ₂₂ O ₁₁	[15]
5	Deacetyl asperulosidic acid methyl ester	C ₁₇ H ₂₄ O ₁₁	[15,17]
6	Geniposidic acid	C ₁₆ H ₂₂ O ₁₀	[18]
7	10-O-Acetyl geniposidic acid	C ₁₈ H ₂₄ O ₁₁	[15]
8	10-Dehydro geniposide	C ₁₇ H ₂₂ O ₁₀	[17]
9	10-Dehydro geniposidic acid	C ₁₆ H ₂₀ O ₁₀	[19]
10	Diffusoside A	C ₁₉ H ₂₈ O ₁₁	[20]
11	Diffusoside B	C ₁₉ H ₂₈ O ₁₁	[20]
12	Lupenylacetate	C ₃₂ H ₅₂ O ₂	[21]
13	Alpigenoside	C ₁₈ H ₂₈ O ₁₂	[15]
14	Oldenlandoside III	C ₃₄ H ₄₄ O ₂₀	[22]
15	5-O-Feruloyl scandoside methyl ester	C ₂₇ H ₃₂ O ₁₄	[23]
16	Hehycoryside C	C ₂₃ H ₂₆ O ₁₁	[22]
17	6- α -Hydro scandoside	C ₁₆ H ₂₂ O ₁₁	[24]
18	6- β -Hydro scandoside	C ₁₆ H ₂₂ O ₁₁	[24]
19	6-Dehydro scandoside	C ₁₆ H ₂₂ O ₁₀	[19]
20	6- α -Hydro scandoside methyl ester	C ₁₇ H ₂₄ O ₁₁	[24]
21	6- β -Hydro scandoside methyl ester	C ₁₇ H ₂₄ O ₁₁	[24]
22	6- α -Hydro-10-acetyl asperuloside acid	C ₁₈ H ₂₄ O ₁₂	[24]
23	6- β -Hydro-10-acetyl asperuloside acid	C ₁₈ H ₂₄ O ₁₂	[24]
24	6-O-Methoxyl cinnamoyl scandoside	C ₂₇ H ₃₂ O ₁₃	[23]
25	6-O- <i>p</i> -Hydro cinnamoyl scandoside	C ₂₆ H ₃₀ O ₁₃	[23]
26	(<i>E</i>)-6-O- <i>p</i> -Coumaroyl-10-O-formoxyl scandoside methyl ester	C ₂₇ H ₃₂ O ₁₃	[14]
27	(<i>E</i>)-6-O- <i>p</i> -Coumaroyl scandoside methyl ester	C ₂₆ H ₃₀ O ₁₃	[14,15,25]
28	(<i>Z</i>)-6-O- <i>p</i> -Coumaroyl scandoside methyl ester	C ₂₆ H ₃₀ O ₁₃	[18]
29	(<i>E</i>)-6-O- <i>p</i> -Methoxy cinnamoyl scandoside methyl ester	C ₂₇ H ₃₂ O ₁₃	[15,25,26]
30	(<i>Z</i>)-6-O- <i>p</i> -Methoxy cinnamoyl scandoside methyl ester	C ₂₇ H ₃₂ O ₁₃	[26]
31	(<i>E</i>)-6-O-Feruloyl scandoside methyl ester	C ₂₇ H ₃₂ O ₁₄	[15,25,26]
32	(<i>Z</i>)-6-O-Feruloyl scandoside methyl ester	C ₂₇ H ₃₂ O ₁₄	[27]

Table 1. Cont.

NO.	Compound Name	Molecular Formula	Reference
Triterpenes			
33	Arborinone	C ₃₀ H ₄₈ O	[28]
34	Isoarborinol	C ₃₀ H ₅₀ O	[28]
35	Oleanolic acid	C ₃₀ H ₄₈ O ₃	[19]
36	Ursolic acid	C ₃₀ H ₄₈ O ₃	[19]
Flavonoids			
37	Amentoflavone	C ₃₀ H ₁₈ O ₁₀	[26,29]
38	Chrysin-6-C-glucosyl-8-C-arabinosyl	C ₂₆ H ₂₈ O ₁₃	[22]
39	Chrysin-6-C-arabinosyl-8-C-glucosyl	C ₂₆ H ₂₈ O ₁₃	[22]
40	Oroxylin-A-O-glucuronic acid	C ₂₂ H ₂₀ O ₁₁	[22]
41	Wogonin-O-glucuronic acid	C ₂₂ H ₂₀ O ₁₁	[22]
42	5,7-Dihydroxy-3-methoxy flavonol	C ₁₆ H ₁₂ O ₅	[13]
43	5,7,4'-Trihydroxy flavonol	C ₁₅ H ₁₀ O ₆	[13]
44	5-Hydroxy-6,7,3',4'-tetramethoxy flavone	C ₁₉ H ₁₈ O ₇	[21]
45	Quercetin	C ₁₅ H ₁₀ O ₇	[17,19,30]
46	Rutin	C ₂₇ H ₃₀ O ₁₆	[15,25,31]
47	Quercetin-3-O-β-D-glucopyranside	C ₂₁ H ₂₀ O ₁₂	[25,32,33]
48	Quercetin-3-O-β-D-galactopyranoside	C ₂₁ H ₂₀ O ₁₂	[32]
49	Quercetin-3-O-(2-O-glucopyranosyl)-β-D-glucopyranside	C ₂₇ H ₃₀ O ₁₇	[15,25,32,33]
50	Quercetin-3-O-(2-O-glucopyranosyl)-β-D-galactopyranoside	C ₂₇ H ₃₀ O ₁₇	[11,34]
51	Quercetin-3-O-sambubioside	C ₂₆ H ₂₈ O ₁₆	[15,25]
52	Quercetin-3-O-[2-O-(6-O-E-ferloyl)-β-D-glucopyranosyl]-β-D-galactopyranoside	C ₃₇ H ₃₈ O ₂₀	[11,34]
53	Quercetin-3-O-[2-O-(6-O-E-ferloyl)-β-D-glucopyranosyl]-β-D-glucopyranoside	C ₃₇ H ₃₈ O ₂₀	[11,15,25]
54	Quercetin-3-O-[2-O-(6-O-E-sinapoyl)-β-D-glucopyranosyl]-β-D-glucopyranoside	C ₃₈ H ₄₀ O ₂₁	[15]
55	Quercetin-3-O-[2-O-(6-O-E-sinapoyl)-β-D-glucopyranosyl]-β-D-galactopyranoside	C ₃₈ H ₄₀ O ₂₁	[25]
56	Kaempferol	C ₁₅ H ₁₀ O ₆	[17,35]
57	Kaempferol-3-O-β-D-glucopyranside	C ₂₁ H ₂₀ O ₁₁	[32]
58	Kaempferol-3-O-β-D-galactopyranoside	C ₂₁ H ₂₀ O ₁₁	[32]
59	Kaempferol-3-O-(2-O-β-D-glucopyranosyl)-β-D-galactopyranoside	C ₂₇ H ₃₀ O ₁₆	[11,25,34]
60	Kaempferol-3-O-(6-O-α-L-rhamnosyl)-β-D-glucopyranside	C ₂₇ H ₃₀ O ₁₆	[32]
61	Kaempferol-3-O-[2-O-(E-6-O-feruloyl)-β-D-glucopyranosyl]-β-D-glucopyranosyl	C ₃₇ H ₃₈ O ₁₉	[11,25,33]
62	Kaempferol-3-O-[2-O-(6-O-E-feruloyl)-β-D-glucopyranosyl]-β-D-galactopyranoside	C ₃₇ H ₃₈ O ₁₉	[21,34]
Athraquinones			
63	2-Methyl-3-methoxy anthraquinone	C ₁₆ H ₁₂ O ₃	[19]
64	2-Hydroxy-1,3-dimethoxy anthraquinone	C ₁₆ H ₁₂ O ₅	[29]
65	2-Hydroxy-3-methyl-1-methoxy anthraquinone	C ₁₆ H ₁₂ O ₄	[36]
66	2-Hydroxy-3-methyl-4-methoxy anthraquinone	C ₁₆ H ₁₂ O ₄	[37]
67	2-Hydroxy-7-methyl-3-methoxy anthraquinone	C ₁₆ H ₁₂ O ₄	[36]
68	2-Hydroxy-1-methoxy-3-methyl anthraquinone	C ₁₆ H ₁₄ O ₄	[31]
69	2-Hydroxy-3-methyl anthraquinone	C ₁₅ H ₁₀ O ₃	[17,27]
70	2-Hydroxy-1-methoxy anthraquinone	C ₁₅ H ₁₀ O ₄	[27,29]
71	2-Hydroxy-4-methoxy anthraquinone	C ₁₅ H ₁₀ O ₄	[38]
72	2-Hydroxy-3-methoxy-7-methyl anthraquinone	C ₁₆ H ₁₂ O ₄	[36]
73	2-Hydroxy-6-methyl anthraquinone	C ₁₅ H ₁₀ O ₃	[18]
74	2-Hydroxy-3-methoxy-6-methyl anthraquinone	C ₁₆ H ₁₂ O ₄	[18]
75	2,7-Dihydroxy-3-methyl anthraquinone	C ₁₅ H ₁₀ O ₄	[39]
76	3-Hydroxy-2-methyl anthraquinone	C ₁₅ H ₁₀ O ₃	[19]
77	3-Hydroxy-2-methyl-4-methoxy anthraquinone	C ₁₆ H ₁₂ O ₄	[40]
78	2,3-Dimethoxy-6-methyl anthraquinone	C ₁₇ H ₁₄ O ₄	[18]

Table 1. Cont.

NO.	Compound Name	Molecular Formula	Reference
Athraquinones			
79	1,3-Dihydroxy-2-methyl anthraquinone	C ₁₅ H ₁₀ O ₄	[41]
80	1,7-Dihydroxy-6-methoxy-2-methyl anthraquinone	C ₁₆ H ₁₂ O ₅	[41]
81	3-Hydroxy-2-methyl-4-methoxy anthraquinone	C ₁₆ H ₁₀ O ₄	[18]
82	2,6-Dihydroxy-3-methyl-4-methoxy anthraquinone	C ₁₆ H ₁₂ O ₅	[42]
83	2,6-Dihydroxy-1-methoxy-3-methyl anthraquinone	C ₁₆ H ₁₂ O ₅	[31]
84	1-Hydroxy-4-methoxy anthraquinone	C ₁₅ H ₁₀ O ₄	[43]
85	2-Hydroxymethyl-1-hydroxy anthraquinone	C ₁₅ H ₁₀ O ₄	[5]
86	2-Hydroxymethyl anthraquinone	C ₁₅ H ₁₀ O ₃	[5]
Phenolic acids and their derivatives			
87	3,4-Dihydroxy benzoic acid	C ₇ H ₆ O ₄	[21]
88	4-Hydroxy-3-methoxy benzoic acid	C ₈ H ₈ O ₄	[30]
89	<i>trans</i> -Hydroxybenzoic acid	C ₇ H ₆ O ₃	[30]
90	4-Hydroxy-3,5-dimethoxy benzoic acid	C ₉ H ₁₀ O ₅	[30]
91	<i>p</i> -Coumaric acid	C ₉ H ₈ O ₃	[19,29]
92	<i>p</i> -Coumaric acid- <i>O</i> -glucopyranside	C ₁₅ H ₁₈ O ₈	[22]
93	Caffeic acid	C ₉ H ₈ O ₄	[21]
94	Caffeoyl hexoside	C ₁₅ H ₁₈ O ₉	[22]
95	Ferulic acid	C ₁₀ H ₁₀ O ₄	[41]
96	Ferulic acid hexoside	C ₁₆ H ₂₀ O ₉	[22]
97	<i>p</i> -Methoxy cinnamic acid	C ₁₀ H ₁₀ O ₃	[44]
98	4,4'-Dihydroxy- α -truxillic acid	C ₁₈ H ₁₆ O ₆	[44]
99	4,4'-Dimethoxyl- α -truxillic acid	C ₁₉ H ₁₈ O ₆	[45]
100	Octadecyl (<i>E</i>)- <i>p</i> -coumarate	C ₂₇ H ₄₄ O ₃	[46]
101	3-Caffeoyl quinic acid	C ₁₆ H ₁₈ O ₉	[22]
102	4-Caffeoyl quinic acid	C ₁₆ H ₁₈ O ₉	[22]
103	5-Caffeoyl quinic acid	C ₁₆ H ₁₈ O ₉	[22]
104	3- <i>p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	[22]
105	4- <i>p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	[22]
106	5- <i>p</i> -Coumaroyl quinic acid	C ₁₆ H ₁₈ O ₈	[22]
107	3-Feruloyl quinic acid	C ₁₇ H ₂₀ O ₉	[22]
108	4-Feruloyl quinic acid	C ₁₇ H ₂₀ O ₉	[22]
109	5-Feruloyl quinic acid	C ₁₇ H ₂₀ O ₉	[22]
Sterols			
110	Daucosterol	C ₃₅ H ₆₀ O ₆	[19]
111	β -Sitosterol	C ₂₉ H ₅₀ O	[19]
112	Stigmasterol	C ₂₉ H ₄₈ O	[17,19]
113	Stigmasterol-5,2-diene-3 β , 7 α -glycol	C ₂₉ H ₄₈ O ₂	[47]
Volatile oils			
114	6,10,14-Trimethyl-2-pentadecanone	C ₁₈ H ₃₆ O	[48]
115	Phytol	C ₂₀ H ₄₀ O	[48]
116	α -Cedrol	C ₁₅ H ₂₆ O	[48]
117	Tetradecanoic acid	C ₁₄ H ₂₈ O ₂	[48]
118	Hexadecanoic acid, methyl ester	C ₁₇ H ₃₄ O ₂	[48]
119	Hexadecanoic acid,	C ₁₆ H ₃₂ O ₂	[48]
121	1,2-Benzenedicarboxylic acid isobutyl ester	C ₁₆ H ₂₂ O ₄	[48]
122	1,2-Benzenedicarboxylic acid, bis(2-methylpropyl)ester	C ₁₆ H ₂₂ O ₄	[48]
123	9,12,15-Octadecatrienoic acid, methyl ester	C ₁₉ H ₃₂ O ₂	[48]
124	9-Octadecenoic acid	C ₁₈ H ₃₄ O ₂	[48]
125	9,12-Octadecenoic acid	C ₁₈ H ₃₂ O ₂	[48]
126	Ethyl linoleate	C ₂₀ H ₃₆ O ₂	[48]
127	Triethyl phosphate	C ₆ H ₁₅ O ₄ P	[48]
128	4-Vinyl phenol	C ₈ H ₈ O	[48]
129	2-Methoxy-4-vinylphenol	C ₉ H ₁₀ O ₂	[48]
130	<i>n</i> -Pentadecanoic acid	C ₁₅ H ₃₀ O ₂	[48]

Table 1. Cont.

NO.	Compound Name	Molecular Formula	Reference
Volatile oils			
131	4,8,12,16-Tetramethyl heptadecan-4-olide	C ₂₁ H ₄₀ O ₂	[48]
132	2,6,10,14,18,22-Tetracosahexaene	C ₃₀ H ₅₀	[48]
133	α-Terpineol	C ₁₀ H ₁₈ O	[11]
134	Geranyl acetate	C ₁₂ H ₂₀ O ₂	[11]
135	β-Ionone	C ₁₃ H ₂₀ O	[11]
136	Lauric acid	C ₁₂ H ₂₄ O ₂	[11]
137	Myristic acid	C ₁₄ H ₂₈ O ₂	[11]
138	Palmitic acid	C ₁₆ H ₃₂ O ₂	[11]
139	Linoleic acid	C ₁₈ H ₃₂ O ₂	[11]
140	β-Linalool	C ₁₀ H ₁₈ O	[11]
141	Isoborneol	C ₁₀ H ₁₈ O	[49]
142	3-(2-Propenyl)-cyclohexene	C ₉ H ₁₄	[49]
143	2-Pentyl-furam	C ₉ H ₁₄ O	[49]
144	Cis-2-(2-pentenyl)-furan	C ₉ H ₁₂ O	[49]
145	Limonene	C ₁₀ H ₁₈	[49]
146	3,7-Dimethyl-1,6-octadiem-3-ol	C ₁₀ H ₁₈ O	[49]
147	trans-5-Methyl-2-(1-methylethyl)-cyclohexanope	C ₁₀ H ₁₈ O	[49]
148	(1S-endo)-1,7,7-Trimethyl-bicyclo[2,2,1]heptan-2-ol	C ₁₀ H ₁₈ O	[49]
149	p-Menth-1-en-8-ol	C ₁₀ H ₁₈ O	[49]
150	Pulegone	C ₁₀ H ₁₆ O	[49]
151	4-(2,6,6-Trimethyl-1-cyclohexen-1-yl)-3-buten-2-one	C ₁₃ H ₂₀ O	[49]
152	Hexadecanal	C ₁₆ H ₃₂ O	[49]
153	2,6,10,14-Tetramethyl-hexadecane	C ₂₀ H ₄₂	[49]
154	(Z,Z)-9,12-octadecadienoic acid	C ₁₈ H ₃₂ O ₂	[49]
155	(Z)-9,17-octadecadienal	C ₁₈ H ₃₂ O	[49]
156	Cis,cis,cis-7,10,13-hexadecatrienal	C ₁₆ H ₂₆ O	[49]
157	Oleic acid	C ₁₈ H ₃₄ O ₂	[49]
158	Hexaldehyde	C ₆ H ₁₂ O	[49]
159	Borneol	C ₁₀ H ₁₈ O	[49]
160	Docosane	C ₂₂ H ₄₆	[49]
161	Tetracosane	C ₂₄ H ₅₀	[49]
162	Hexacosane	C ₂₆ H ₅₄	[49]
163	Heptacosane	C ₂₇ H ₅₆	[49]
Polysaccharides			
164	ODP-1		[50]
Cyclotides			
165	CD1		[51]
166	CD2		[51]
167	CD3		[51]
Coumarins			
168	7-Hydroxy-6-methoxy-Coumarin	C ₁₀ H ₈ O ₄	[17]
169	Esculetin	C ₉ H ₆ O ₄	[46]
Alkaloids			
170	10(S)-hydroxy pheophytin a	C ₅₅ H ₇₄ N ₄ O ₆	[52]
171	Aurantiamide acetate	C ₂₇ H ₂₈ N ₂ O ₄	[46]

2.1. Iridoids and Triterpenes

Iridoids are one of the most important components in *H. diffusa* with various bioactivities, such as anti-oxidant, neuroprotective and anti-inflammatory effects [33,53]. Accompanied with the analysis of the NMR spectra of the pure compounds, the methods of tandem mass spectrometry (MS^n) and time-of-flight mass spectrometry (TOF/MS) have become more popular for the identification of these compounds [11,14,15,25,52]. To date, thirty-two iridoids and their iridoid glucosides (1–32) have been isolated and identified from *H. diffusa* (Figure 1).

Four triterpenes, named arborinone (33), isoarborinol (34), oleanolic acid (35) and ursolic acid (36), were isolated from *H. diffusa* and their structures were established by 1D-, 2D-NMR spectroscopic analysis and high-resolution electrospray ionization mass spectroscopy (HRESIMS) [53].

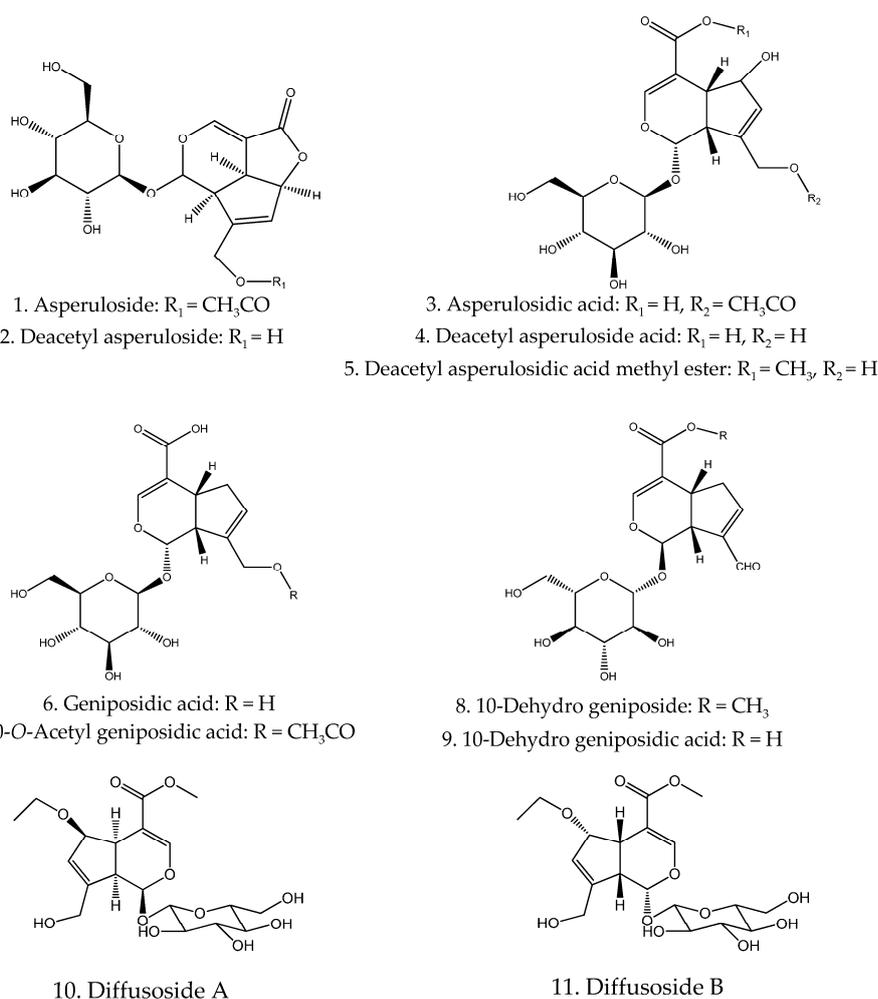
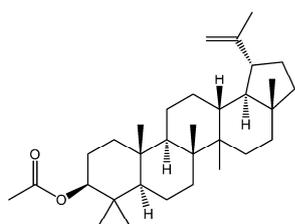
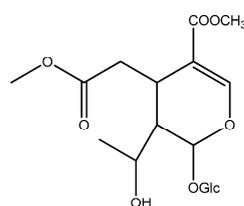


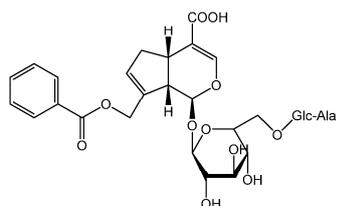
Figure 1. Cont.



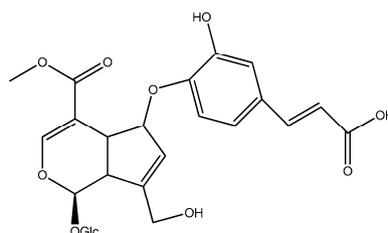
12. Lupenylacetate



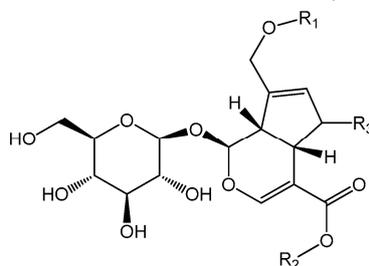
13. Alpigenoside



14. Oldenlandoside III



15. 5-O-Feruloyl scandoside methyl ester



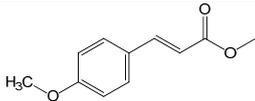
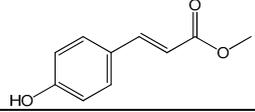
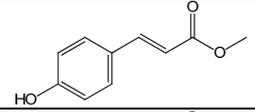
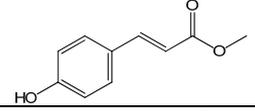
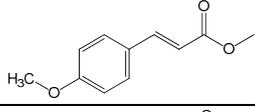
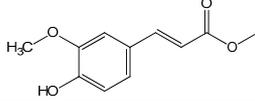
Compounds	R ₁	R ₂	R ₃
16. Hehycoryside C	benzoyl	H	H
17/18. 6- α/β -Hydro scandoside	H	H	OH
19. 6-Dehydro scandoside	H	H	H
20/21. 6- α/β -Hydro Scandoside methyl ester	H	CH ₃	OH
22/23. 6- α/β -Hydro-10-acetyl asperuloside acid	CH ₃ CO	H	OH
24. 6-O- <i>p</i> -Methoxy cinnamoyl scandoside	H	H	
25. 6-O- <i>p</i> -Hydro cinnamoyl scandoside	H	H	
26. (<i>E</i>)-6-O- <i>p</i> -Coumaroyl-10-O-formoxyl scandoside methyl ester	HCO	CH ₃	
27/28. (<i>E/Z</i>)-6-O- <i>p</i> -Coumaroyl scandoside methyl ester	H	CH ₃	
29/30. (<i>E/Z</i>)-6-O- <i>p</i> -Methoxy cinnamoyl scandoside methyl ester	H	CH ₃	
31/32. (<i>E/Z</i>)-6-O-Feruloyl scandoside methyl ester	H	CH ₃	

Figure 1. Cont.

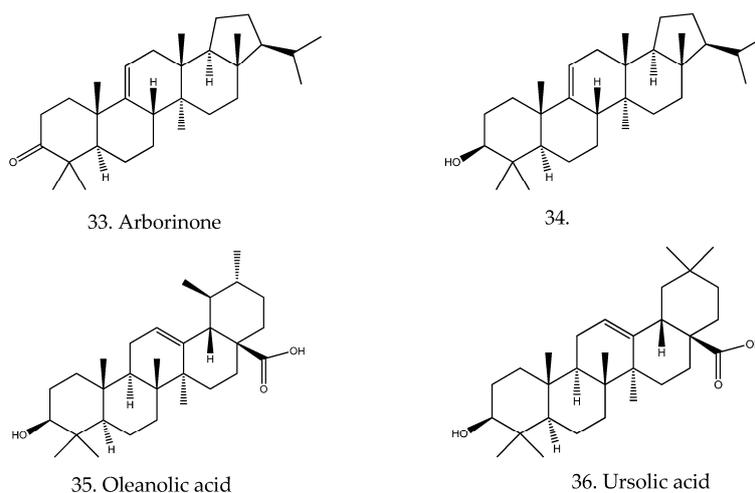
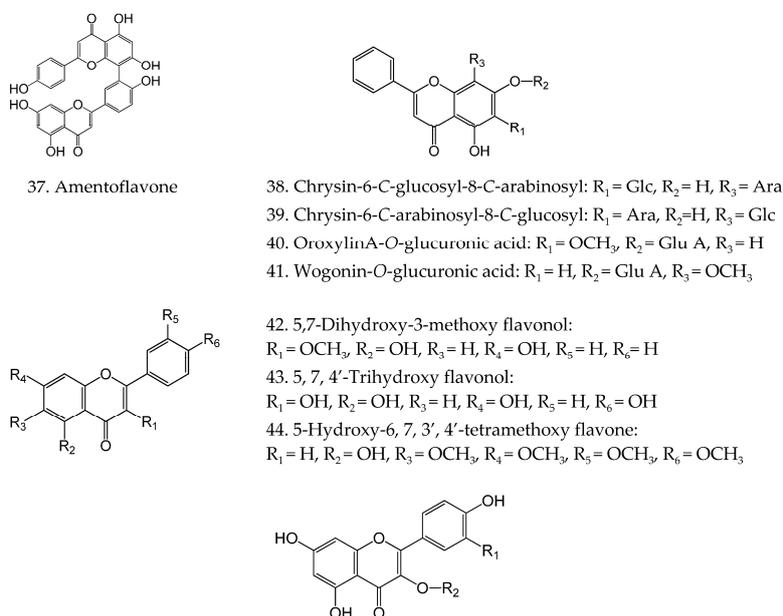


Figure 1. Chemical structures of iridoids and triterpenes in *H. diffusa*.

2.2. Flavonoids

Flavonoids are a major group presented in *H. Diffusa*, and most of them are derivatives of the flavonol aglycones of kaempferol and quercetin. Recently, other aglycones, such as chrysin, oroxylin and wogonin, have been characterized by ultra-performance liquid chromatography–diode array detector/quadrupole time-of-flight mass spectrometry (UPLC-DAD/Q-TOF-MS). To date, twenty-six flavonoids (37–62) with various substitutions have been identified and their chemical structures are prescribed in Figure 2.



Compounds	R_1	R_2
45. Quercetin	OH	H
46. Rutin	OH	rutinose
47. Quercetin-3-O- β -D-glucopyranside	OH	β -D-Glc
48. Quercetin-3-O- β -D-galactopyranside	OH	β -D-Gal
49. Quercetin-3-O-(2-O-glucopyransyl)- β -D-glucopyranside	OH	β -D-Glc-(1 \rightarrow 2)-D-Glc

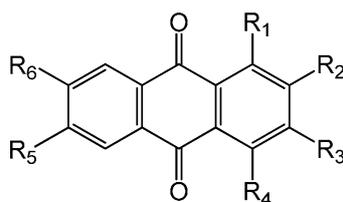
Figure 2. Cont.

Compounds	R ₁	R ₂
50. Quercetin-3-O-(2-O-glucopyranosyl)-β-D-galactopyranoside	OH	β-D-Glc-(1→2)-D-Gal
51. Quercetin-3-O-sambubioside	OH	β-D-Xyl-(1→2)-D-Glc
52. Quercetin-3-O-[2-O-(6-O-E-feruloyl)-β-D-glucopyranosyl]-β-D-galactopyranoside	OH	6'-O-E-feruloyl-β-D-Glc-(1→2)-D-Gal
53. Quercetin-3-O-[2-O-(6-O-E-feruloyl)-β-D-glucopyranosyl]-β-D-glucopyranoside	OH	6'-O-E-feruloyl-β-D-Glc-(1→2)-D-Glc
54. Quercetin-3-O-[2-O-(6-O-E-sinapoyl)-β-D-glucopyranosyl]-β-D-glucopyranoside	OH	6'-O-E-sinapoyl-β-D-Glc-(1→2)-D-Glc
55. Quercetin-3-O-[2-O-(6-O-E-sinapoyl)-β-D-glucopyranosyl]-β-D-galactopyranoside	OH	6'-O-E-sinapoyl-β-D-Glc-(1→2)-D-Gal
56. Kaempferol	H	H
57. Kaempferol-3-O-β-D-glucopyranoside	H	β-D-Glc
58. Kaempferol-3-O-β-D-galactopyranoside	H	β-D-Gal
59. Kaempferol-3-O-(2-O-β-D-glucopyranosyl)-β-D-galactopyranoside	H	β-D-Glc-(1→2)-D-Gal
60. Kaempferol-3-O-(6-O-α-L-rhamnosyl)-β-D-glucopyranoside	H	α-L-Rha-(1→6)-β-D-Glc
61. Kaempferol-3-O-[2-O-(6-O-E-feruloyl)-β-D-glucopyranosyl]-β-D-glucopyranoside	H	6'-O-E-feruloyl-β-D-Glc-(1→2)-D-Glc
62. Kaempferol-3-O-[2-O-(6-O-E-feruloyl)-β-D-glucopyranosyl]-β-D-galactopyranoside	H	6'-O-E-feruloyl-β-D-Glc-(1→2)-D-Gal

Figure 2. Chemical structures of flavonoids in *H. diffusa*.

2.3. Anthraquinones

Anthraquinones are also a major group of bioactive components in *H. diffusa*. Up to now, twenty-four anthraquinones with various substitutions (63–86) have been obtained and identified from this herb. These compounds have a typical characteristic of the 9, 10-anthraquinone skeleton with the presence of hydroxy, methyl and/or methoxy groups, for example, 2-hydroxy-3-methoxy-6-methyl anthraquinone (77). Their chemical structures are shown in Figure 3.



Compounds	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆
63. 2-Methyl-3-methoxy anthraquinone	H	CH ₃	OCH ₃	H	H	H
64. 2-Hydroxy-1,3-dimethoxy anthraquinone	OCH ₃	OH	OCH ₃	H	H	H
65. 2-Hydroxy-3-methyl-1-methoxy anthraquinone	OCH ₃	OH	CH ₃	H	H	H
66. 2-Hydroxy-3-methyl-4-methoxy anthraquinone	H	OH	CH ₃	OCH ₃	H	H
67. 2-Hydroxy-7-methyl-3-methoxy anthraquinone	H	OH	OCH ₃	H	H	CH ₃
68. 2-Hydroxy-1-methoxy-3-methyl anthraquinone	OCH ₃	OH	CH	H	H	H
69. 2-Hydroxy-3-methyl anthraquinone	H	OH	CH ₃	H	H	H
70. 2-Hydroxy-1-methoxy anthraquinone	OCH ₃	OH	H	H	H	H
71. 2-Hydroxy-4-methoxy anthraquinone	H	OH	H	OCH ₃	H	H
72. 2-Hydroxy-3-methoxy-7-methyl anthraquinone	H	OH	OCH ₃	H	H	CH ₃

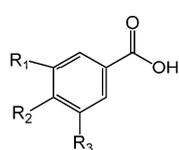
Figure 3. Cont.

Compounds	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆
73. 2-Hydroxy-6-methyl anthraquinone	H	OH	H	H	CH ₃	H
74. 2-Hydroxy-3-methoxy-6-methyl anthraquinone	H	OH	OCH ₃	H	CH ₃	H
75. 2,7-Dihydroxy-3-methyl anthraquinone	H	OH	CH ₃	H	H	OH
76. 3-Hydroxy-2-methyl anthraquinone	H	CH ₃	OH	H	H	H
77. 3-Hydroxy-2-methyl-4-methoxy anthraquinone	H	CH ₃	OH	OCH ₃	H	H
78. 2,3-Dimethoxy-6-methyl anthraquinone	H	OCH ₃	OCH ₃	H	CH ₃	H
79. 1,3-Dihydroxy-2-methyl anthraquinone	OH	CH ₃	OH	H	H	H
80. 1,7-Dihydroxy-6-methoxy-2-methyl anthraquinone	OH	CH ₃	H	H	OCH ₃	OH
81. 3-Hydroxy-2-methyl-4-methoxy anthraquinone	H	CH ₃	OH	OCH ₃	H	H
82. 2,6-Dihydroxy-3-methyl-4-methoxy anthraquinone	H	OH	CH ₃	OCH ₃	OH	H
83. 2,6-Dihydroxy-1-methoxy-3-methyl anthraquinone	OCH ₃	OH	CH ₃	H	OH	H
84. 1-Hydroxy-4-methoxy anthraquinone	OH	H	H	OCH ₃	H	H
85. 2-hydroxymethyl-1-hydroxy anthraquinone	OH	CH ₂ OH	H	H	H	H
86. 2-hydroxymethyl anthraquinone	H	CH ₂ OH	H	H	H	H

Figure 3. Chemical structures of anthraquinones in *H. diffusa*.

2.4. Phenolic Acids and Their Derivatives

Phenolic acids are very common and important secondary metabolites in nature. To date, twenty-three phenolic acids (87–109) have been identified from the herb of *H. Diffusa*, including four benzoic acid derivatives (87–90), coumaric acid (91) and its derivative (92), caffeic acid (93) and its derivative (94), ferulic acid (95) and its derivative (96), *p*-methoxyl cinnamic acid (97), two truxillic acid derivatives (98–99), octadecyl (*E*)-*p*-coumarate (100) and nine quinic acid derivatives (101–109). Their chemical structures are prescribed in Figure 4.

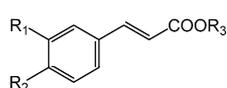


87. 3,4-Dihydroxy benzoic acid: R₁ = OH, R₂ = OH, R₃ = H

88. 4-Hydroxy-3-methoxy benzoic acid: R₁ = OCH₃, R₂ = OH, R₃ = H

89. 4-Hydroxyl benzoic acid: R₁ = OH, R₂ = OH, R₃ = H

90. 4-Hydroxy-3,5-dimethoxy benzoic acid: R₁ = OCH₃, R₂ = OH, R₃ = OCH₃



91. *p*-Coumaric acid: R₁ = H, R₂ = OH, R₃ = H

92. *p*-Coumaric acid-*O*-glucoside: R₁ = H, R₂ = *O*-Glc, R₃ = H

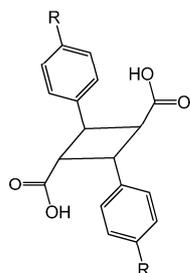
93. Caffeic acid: R₁ = R₂ = OH, R₃ = H

94. Caffeoyl hexoside : R₁ = R₂ = OH, R₃ = Hex

95. Ferulic acid: R₁ = OCH₃, R₂ = OH, R₃ = H

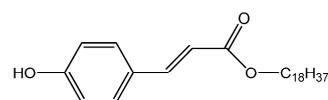
96. Ferulic acid hexoside: R₁ = OCH₃, R₂ = *O*-Hex, R₃ = H

97. *p*-Methoxyl cinnamic acid: R₁ = OCH₃, R₂ = H, R₃ = H



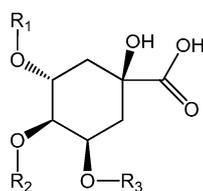
98. 4,4'-Dihydroxy- α -truxillic acid: R = OH

99. 4,4'-Dimethoxyl- α -truxillic acid: R = OCH₃



100. Octadecyl (*E*)-*p*-coumarate

Figure 4. Cont.



101. 3-Caffeoyl quinic acid: $R_1 = \text{caffeoyl}$, $R_2 = \text{H}$, $R_3 = \text{H}$
 102. 4-Caffeoyl quinic acid: $R_1 = \text{H}$, $R_2 = \text{caffeoyl}$, $R_3 = \text{H}$
 103. 5-Caffeoyl quinic acid: $R_1 = \text{H}$, $R_2 = \text{H}$, $R_3 = \text{caffeoyl}$
 104. 3-*p*-Coumaroyl quinic acid: $R_1 = p\text{-coumaroyl}$, $R_2 = \text{H}$, $R_3 = \text{H}$
 105. 4-*p*-Coumaroyl quinic acid: $R_1 = \text{H}$, $R_2 = p\text{-coumaroyl}$, $R_3 = \text{H}$
 106. 5-*p*-Coumaroyl quinic acid: $R_1 = \text{H}$, $R_2 = \text{H}$, $R_3 = p\text{-coumaroyl}$
 107. 3-Feruloyl quinic acid: $R_1 = \text{feruloyl}$, $R_2 = \text{H}$, $R_3 = \text{H}$
 108. 4-Feruloyl quinic acid: $R_1 = \text{H}$, $R_2 = \text{feruloyl}$, $R_3 = \text{H}$
 109. 5-Feruloyl quinic acid: $R_1 = \text{H}$, $R_2 = \text{H}$, $R_3 = \text{feruloyl}$

Figure 4. Chemical structures of phenolic acids and their derivatives in *H. diffusa*.

2.5. Polysaccharides

The polysaccharides in *H. diffusa* have been researched for their immuno-enhancing activity. They are mainly composed of glucose, galactose and mannose, with the content of 15.10% determined by the spectrophotometry method at 490 nm [54]. Up to now, only one homogeneous polysaccharide, ODP-1, has been separated from *H. diffusa*, with the relative molecular weight of 20.88 kDa. It consists of mannose, rhamnose, galacturonic acid, glucose, galactose and arabinose, with the molar ratio of 0.005:0.033:0.575:1.000:0.144:0.143 [50].

2.6. Essential Oils

The reports of essential oils in this plant were mainly on isolating many fatty acids, fatty acid esters, etc. [11]. Yang *et al.* [49] identified 29 compounds representing 81.45% of the total oil content by GC/MS combined with the Kovats Retention index. *n*-Hexadecanoic acid (**119**) (31.22%), oleic acid (**157**) (6.74%), tetracosane (**161**) (4.94%) and 9,12-octadecadienoic acid (**125**) (4.87%) were found to be the main constituents. Liu *et al.* [48] compared the constituents and their content in *H. diffusa* collected from the provinces of Guangdong, Jiangxi and Guangxi in China and also revealed that the oil of *H. diffusa* was mainly composed of fatty acids with an oil extraction rate from 0.25% to 0.30%.

2.7. Cyclotides

Three novel cyclotides from *H. diffusa*, named CD1 (**165**), CD2 (**166**) and CD3 (**167**), with an anti-cancer effect on prostate cancer cells, were reported by Hu *et al.* [51]. The primary sequences were GAFLKCGESCIVYLPCLTTVVGCSQNSVCYRD, GAVPCGETCVYLPCLTPDIGCSCQNKVCYRD and G-TSCGETCVLLPCLSSVLGCTCQNKRCYKD for DC1, DC2 and DC3, respectively.

2.8. Miscellaneous

Only four sterols of daucosterol (**110**), β -sitosterol (**111**), stigmasterol (**112**) and stigmasterol-5,2-diene-3 β , 7 α -glycol (**113**), two coumarins of 7-hydroxy-6-methoxy-coumarin (**168**) and esculetin (**169**) and two alkaloids of 10-hydroxyphenophytin a (**170**) and aurantiamide acetate (**171**) have been purified and characterized from *H. diffusa* and their structures are shown in Figure 5.

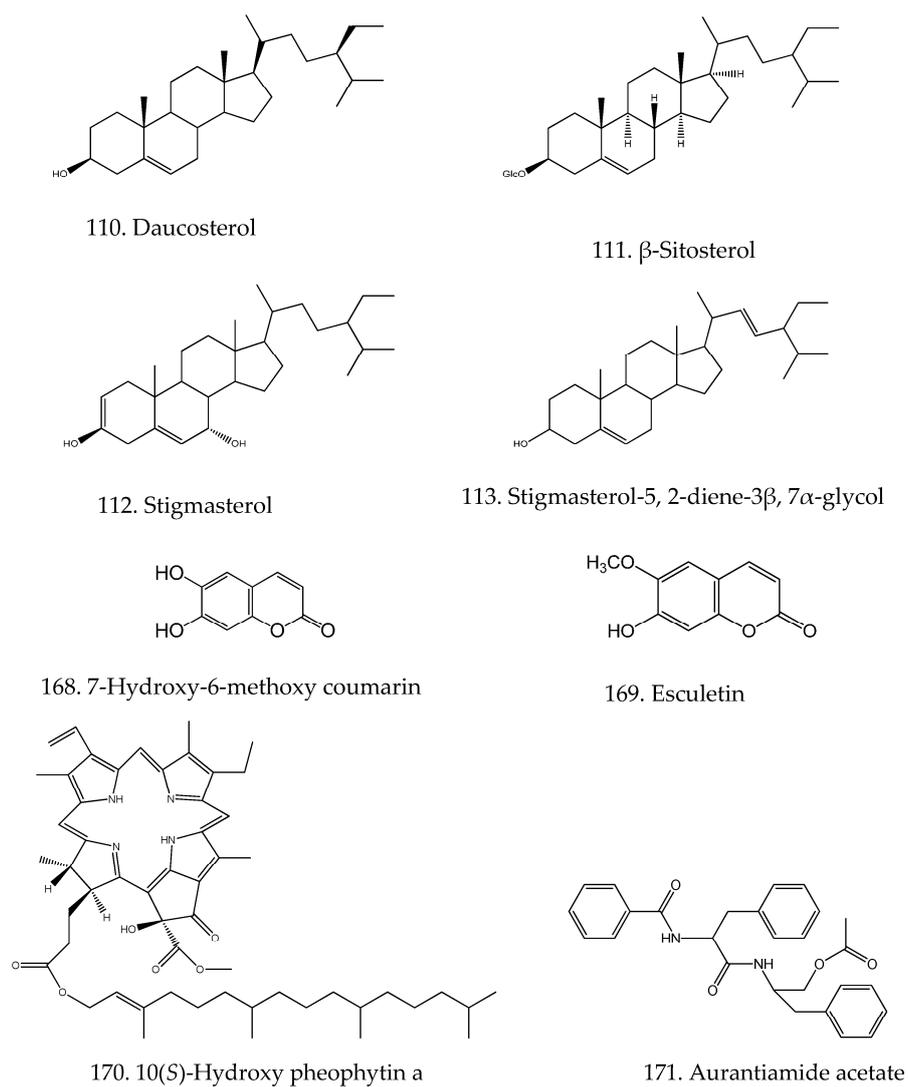


Figure 5. Chemical structures of miscellaneous components in *H. diffusa*.

3. Pharmacology

H. diffusa has long been used therapeutically in China, due to its broad spectrum of biological and pharmacological activities. Now we have enlisted an overview of the modern pharmacological studies in the following sections (Table 2).

Table 2. Pharmacological effects of *H. diffusa*.

Activities	Model	Formulation/Dosage/Extract	Reference
Anti-tumor activity			
Colorectal cancer	HT-29 cells	Ethanol extract	The extract suppressed HT-29 cell growth and induced apoptosis via inactivation of the IL-6/STAT3-signaling pathway. [2]
	HT-29 cells	Ethanol extract	The extract reduced HT-29 cell viability and survival. It could suppress cancer cell proliferation by blocking the cell cycle, preventing G ₁ to S progression, and reducing mRNA expression of pro-proliferative PCNA, Cyclin D1 and CDK4, but increasing that of anti-proliferative p21. [55]
	HT-29 cells	Ethanol extract	The extract induced the HT-29 cell morphological changes and reduced cell viability. In addition, the extract treatment resulted in DNA fragmentation, loss of plasma membrane asymmetry, collapse of mitochondrial membrane potential, activation of caspase-9 and caspase-3 and increase of the ratio of pro-apoptotic Bax to anti-apoptotic Bcl-2. [56]
	HT-29 cells	Ethanol extract	The extract treatment downregulated the mRNA and protein expression levels of VEGF-A in HT-29 human colon carcinoma cells. [57]
	HT-29 cells	Ethanol extract	The extract inhibits colorectal cancer growth <i>in vivo</i> via inhibition of SHH-mediated tumor angiogenesis. [58]
	CRC mouse xenograft model	Ethanol extract	The extract inhibited the expression of the gene VEGF-A and VEGFR2, thus, suppressed the activation of Sonic hedgehog (SHH)-signaling in CRC xenograft tumors; it inhibits colorectal cancer growth. [58]
	CRC mouse xenograft model	Ethanol extract	The extract suppressed the STAT3 pathway by suppressing STAT3 phosphorylation in tumor tissues, altering the expression pattern of target genes of Cyclin D1, CDK4 and Bcl-2, as well as upregulating p21 and Bax. [59]
	CT-26 cells	Ethanol extract	The extract can inhibit the proliferation of CT-26 colon cancer cells from BALB/c mice in a time- and dose- dependent manner. [60]
	HCT-8/5-FU cells	Ethanol extracts	The extract treatment significantly reduced the cell viability of HCT-8/5-FU cells by downregulating the expression of P-gp and ABCG2. [61]
	Caco-2 cells	Aqueous extracts	The decoction of <i>H. diffusa</i> and its fraction 9 contained sufficient ursolic acid and oleanolic acid to possibly induce apoptosis of Caco-2 cells. [62]
	Caco-2 cells	Nine pure compounds isolated from <i>H. diffusa</i>	2-Hydroxymethy-1-hydroxy anthraquinone (IC ₅₀ 45 mM) and ursolic acid (IC ₅₀ 71 mM) exhibited the highest inhibition of Caco-2 cell proliferation. [5]
Leukemia	CEM cells	Aqueous extract	The extract inhibited Leukemia CEM cells growth in time- and concentration-dependent manners. And the inhibition mechanism has greater correlation with the upregulation of P53 expression. [63]

Table 2. Cont.

Activities	Model	Formulation/Dosage/Extract		Reference
Anti-tumor activity				
	BALB/c mice	Aqueous extract	The extract had anti-leukemia effects on WEHI-3 cell-induced leukemia <i>in vivo</i> .	[64]
	HL-60 cells	<i>H. diffusa</i> injection	The extract could induce HL-60 cells differentiation, and suppress the expression of the anti-apoptosis-related gene to inhibit the growth of HL-60 cells.	[65]
	HL-60 cells, WEHI-3 cells	Ethanol extract	The extract inhibited the cell proliferation of HL-60 cells. It triggered an arrest of HL-60 cells at the G ₀ /G ₁ phase and sub-G ₁ population, provoked DNA condensation and DNA damage, but the activities of caspase-3, caspase-8, and caspase-9 were elevated in <i>H. diffusa</i> -treated HL-60 cells.	[66]
	U937 cells	2-Hydroxy-3-methyl anthraquinone	2-Hydroxy-3-methyl anthraquinone enhanced apoptosis of U937 cells through the activation of p-p38MAPK and downregulation of p-ERK1/2.	[67]
	THP-1 Cells	2-Hydroxy-3-methyl anthraquinone	2-Hydroxy-3-methyl anthraquinone induced THP-1 cell apoptosis, which was associated with a more prominent induction expression of Fas/FasL, DR4 and TRAIL. Moreover, 2-Hydroxy-3-methylanthraquinone treatment resulted in activation of caspase-8.	[68]
Liver cancer	H22 mice	Aqueous extract	The extract had an inhibitory effect on the metastasis of hepatocarcinoma in blood.	[69]
	HepG2 cells	Aqueous extract	The extract remarkably inhibited HepG2 cell proliferation via arrest of HepG2 cells at the G ₀ /G ₁ phase and induction of S phase delay. In addition, the extract potentiated the anticancer effect of low-dose 5-FU in the absence of overt toxicity by downregulating the mRNA and protein levels of CDK2, cyclin E and E2F1.	[70]
	MHCC97-H cells	Total flavones extract	The extract treatment reduced the level of E-cadherin protein and increased the expression of vimentin protein in TGF-β1-induced MHCC97-H.	[71]
	HepG2 cells	1,3-Dihydroxy-2-Methylanthraquinone Ethyl acetate extract	Both 1,3-Dihydroxy-2-Methylanthraquinone and ethyl acetate extract exhibited an inhibitory effect on HepG2 cells, resulting in upregulation of Bax, p53, Fas, FasL, p21 and cytoplasmic cytochrome C levels and caspase-3, -8, -9 proteases activities, while downregulating Bcl-2, mitochondrial cytochrome C, cyclin E and CDK 2 in a dose-dependent manner.	[72]
	HepG2 cells	Nine pure compounds isolated from <i>H. diffusa</i>	Ursolic acid exhibited a strong inhibition of cell survival with C ₅₀ 37 mM.	[5]
	HepG2 cells	2-Hydroxy-3-methyl anthraquinone 1-Methoxy-2-hydroxy anthraquinone	Both compounds showed inhibitory activity against protein tyrosine kinases v-src and pp60src and arrested the growth of HepG2 cancer cells.	[38]

Table 2. Cont.

Activities	Model	Formulation/Dosage/Extract	Reference
Anti-tumor activity			
Lung cancer	A549 cells, H1355 cells, LLC cells	Ethanol extract	The extract suppressed the cell proliferation of A549 and H1355 cells as well as reduced cell viability in a concentration-dependent manner. [66]
	SPC-1-A cells	2-Hydroxy-3-methyl anthraquinone 1-Methoxy-2-hydroxy anthraquinone	Both compounds showed inhibitory activity against protein tyrosine kinases v-src and pp60src and arrested the growth of SPC-1-A. [38]
Breast cancer	MCF-7 cells	Compounds of anthraquinones, iridoid glucosides, stigmasterols and alkaloids/flavonoids	Alkaloids/flavonoids possessed antitumor activity against the human breast cancer cell line MCF7 [73]
	MCF-7 cells	Methyl anthraquinone	Methyl anthraquinone-induced MCF-7 cells apoptosis via Ca ²⁺ /calpain/caspase-4 pathway. [74]
	Bcap37 cells	2-Hydroxy-3-methyl anthraquinone, 1-Methoxy-2-hydroxy anthraquinone	Both compounds showed inhibitory activity against protein tyrosine kinases v-src and pp60src and arrested the growth of Bcap37 cells. [38]
Cervical tumor	Nude mouse model	Aqueous extract	The extract had an inhibitory effect on cervical cancer cells with the expression of Ki-67 protein significantly decreased, and the mean survival time of the mice was significantly extended. [3]
	HeLa cells	Nine pure compounds isolated from <i>H. diffusa</i>	2-Hydroxymethyl-1-hydroxy anthraquinone exhibited the strongest inhibitory effect on cell viability. [5]
Prostate Cancer	DU145 cells, PC-3 cells LNCaP cells	Nine pure compounds isolated from <i>H. diffusa</i>	2-Methyl-3-methoxy anthraquinone, 2-hydroxy-3-methyl anthraquinone and ursolic acid exhibited inhibitory effects on prostate cancer cell survival. [5]
	PC3 cells LNCaP cells	6- <i>O</i> -(<i>E</i>)- <i>p</i> -Coumaroyl scandoside methyl ester 10(<i>S</i>)-Hydroxy pheophytin	Two compounds showed a moderate anti-proliferation effect on PC3 human androgen-independent prostate cancer cells, while 10(<i>S</i>)-hydroxy pheophytin also showed a strong anti-proliferation effect on LNCaP human androgen-sensitive prostate cancer cells. [52]
Multiple myeloma	RPMI 8226 cells	Nine pure compounds isolated from <i>H. diffusa</i>	2-Hydroxymethyl-1-hydroxy anthraquinone exhibited the strongest inhibition of RPMI 8226 cells growth. [5]
	RPMI 8226 cells	Polysaccharides extracts	Polysaccharides extracts suppressed the growth of RPMI 8226 cells in a dose- and time-dependent manner. [75]
	RPMI 8226 cells	<i>H. diffusa</i> injection	<i>H. diffusa</i> injection could inhibit the proliferation of RPMI 8226 cells. [76]
Others	B16F10 cells	Ethanol extract	The extract suppressed the cell proliferation of B16F10 cells as well as reducing cell viability in a concentration-dependent manner. [66]
	S180 cells	Decoction, lipophilic extract, crude polysaccharide	Lipophilic extract and crude polysaccharide showed anti-tumor activities and a protective effect on chemotherapeutic damage. However, the aqueous extract had no marked anti-tumor effect on S-180 cells. [77]

Table 2. Cont.

Activities	Model	Formulation/Dosage/Extract		Reference
Anti-tumor activity				
	MG-63 cells	<i>H. diffusa</i> injection	<i>H. diffusa</i> injection could inhibit the proliferation of MG-63 cells, and Bax gene expression was significantly increased.	[78]
	MG-63 cells	<i>H. diffusa</i> injection	<i>H. diffusa</i> injection could induce the apoptosis of MG-63 cells by increasing Bax gene expression in a concentration-dependent manner.	[79]
	MG-63 cells	Aqueous extract	<i>H. diffusa</i> , combined with cisplatin, had a stronger inhibitory effect than the single agents in MG-63 cells with IC ₅₀ 164.6 and 5.0 µL/mL, respectively. As a result, <i>H. diffusa</i> could alter anti-apoptotic (Bax and Bad) and pro-apoptotic protein (Bcl-xl and Bcl-2) expression, and it elevated the levels of caspase-3 and caspase-8.	[80]
	U87 cells	Aqueous extract	The extract suppressed U87 cells growth in a dose- and time-dependent manner.	[4]
Angiogenesis	1. Breast tumor-bearing BALB/c mice 2. Zebrafish embryo model 3. Human endothelial cells 4. C57BL/6 mice	4-Vinyl phenol	4-Vinyl phenol was demonstrated with anti-angiogenic activity <i>in vitro</i> and <i>in vivo</i> .	[81]
Immunomodulatory effect				
	Normal BALB/c mice	Ethanol Extract	The extract has promoted immune responses in normal BALB/c mice.	[82]
	Immunosuppression mice induced by cyclophosphamide	Polysaccharides extracts	The extract could improve the clearance index, phagocytic index, and the index of the thymus and spleen of immunosuppression mice.	[50]
	Inmmunosuppressed mice induced by cyclophosphamide	Total flavonoids extract	The extract enhanced specific and non-specific immunity.	[83]
Antioxidant effects				
		The extract from methanol, acetone and 80% alcohol	The extraction with 80% alcohol has the strongest antioxidant activity on DPPH assay.	[84]
		The extract from water, ethanol, acetone, chloroform, ether, petroleum benzine	Acetone extract had the strongest antioxidant effect.	[85]
	LO ₂ cells	Aqueous extract	The aqueous extract exerted a good antioxidant effect in DPPH assay with a 50% scavenging concentration at 0.153 mg/mL. Aqueous extract treatment reversed H ₂ O ₂ -induced activation of the MEK/ERK pathway and H ₂ O ₂ -induced inhibition of the P13-K/AKT/GSK3b pathway in LO ₂ cells. This may be due to the improvement activity of the aqueous extract of <i>H. diffusa</i> on the antioxidant defense system.	[86]
		Twelve pure compounds isolated from <i>H. diffusa</i>	All compounds showed antioxidant effects on xanthine oxidase inhibition, xanthine-xanthine oxidase cytochrome c and TBA-MDA systems.	[33]

Table 2. Cont.

Activities	Model	Formulation/Dosage/Extract	Reference
Anti-inflammatory effect			
	Lipopolysaccharide-induced renal inflammation mice	Aqueous extract	The extract protected renal tissues, significantly suppressed the production of TNF- α , IL-1, IL-6 and MCP-1, as well as significantly promoted the production of IL-10 in serum and renal tissues. [87]
	RAW 264.7 cells	Total flavonoids extract	The extract treatment on LPS-stimulated RAW 264.7 cells, reduced expression of iNOS, TNF- α , IL-6 and IL-1 β , as well as suppressing phosphorylation of I κ B p38, JNK and ERK1/2 in a concentration-dependent manner, indicating that the anti-inflammatory activity of total flavonoids had a close relationship with the NF- κ B and MAPK signaling pathways. [88]
Neuroprotective effect			
	Rat cortical cells damaged by L-glutamate	Methanolic extract, five flavonoids and four <i>O</i> -acylated iridoid glycosides	All compounds exhibited significant neuroprotective activity in primary cultures of rat cortical cells damaged by L-glutamate. [34]
Anti-fibrosis effect			
	Ras oncogene-transformed R6 cells	Oleanolic acid	Oleanolic acid inhibits the growth of ras oncogene-transformed R6 cells. Oleanolic acid-mediated growth inhibition of transformed cells does not require direct cell–cell contact between normal and ras-transformed cells. [89]

3.1. Anti-Cancer Activity

3.1.1. Anti-Colorectal Cancer Activity

H. diffusa has been used as a major formula for the clinical treatment of colorectal cancer (CRC). *In vitro*, ethanol extract of *H. diffusa* treatment significantly suppresses proliferation and induced apoptosis of HT-29 cells, resulting in DNA fragmentation, loss of plasma membrane asymmetry, collapse of mitochondrial membrane potential, activation of caspase-9 and caspase-3, increase of the ratio of pro-apoptotic Bax to anti-apoptotic Bcl-2, reduction of the mRNA expression levels of cyclin D1, cyclin-dependent kinase 4 and B-cell lymphoma-2 (Bcl-2), upregulation of the expression levels of Bcl-2-associated X protein, prevention of G1-S progression, and reduction of mRNA expression of pro-proliferative PCNA, Cyclin D1 and CDK4. These results indicated that the anti-colorectal cancer cells effect of *H. diffusa* might be carried out via multiple approaches, such as the mitochondria-dependent pathway, IL-6/STAT3 pathway and cell cycle arrest [2,55–58]. The mechanism was also confirmed by animal experiments [58,59]. Meanwhile, the ethanolic extract of *H. diffusa* displayed an inhibition effect on CT-26 cells with inhibitory rates from $35.46\% \pm 3.59\%$ to $71.84\% \pm 3.12\%$ at different concentrations (0.06 mg/mL, 0.08 mg/mL, 0.10 mg/mL, 0.12 mg/mL, 0.14 mg/mL, 0.16 mg/mL, 0.18 mg/mL and 0.20 mg/mL) and showed a stronger inhibition effect with an increase of concentration [60]. Li *et al.* [61] revealed that the ethanolic extract treatment could overcome 5-fluorouracil resistance in HCT-8/5-FU cells by downregulating the expression of P-gp and ABCG2. In addition, 2-hydroxymethyl-1-hydroxy anthraquinone (IC₅₀ 45 µM) and ursolic acid (IC₅₀ 71 µM) isolated from *H. diffusa* exhibited inhibition effects on Caco-2 cell proliferation [5], and the mechanism of the inhibition effect for ursolic acid might include the cleavage of the Poly (ADP-ribose) Polymerase (PARP) [62].

3.1.2. Anti-Leukemia Activity

The anti-leukemia effects of both aqueous and ethanolic extracts of *H. diffusa* have been investigated in several cancer cell lines. *H. diffusa* aqueous extract treatment with 0.01–4150 µg/mL restrained the growth of the CEM cells by enhancing the expression of P53 *in vitro* [63] and influenced murine leukemia WEHI-3 cells, as well as promoting T- and B-cell proliferation in leukemic mice administrated with 16 and 32 mg/kg *in vivo* [64]. The ethanolic extract of *H. diffusa* could trigger an arrest of HL-60 cells at the G₀/G₁ phase and sub-G₁ population, provoke DNA condensation and DNA damage, but elevate the activities of caspase-3, caspase-8 and caspase-9, thus, inhibiting the cell proliferation of HL-60 cells with the half maximal inhibitory concentration (IC₅₀) value of 4.62 mg/mL [65,66]. Wang *et al.* [67] found that 2-hydroxy-3-methyl anthraquinone treatment (0–80 µM) could enhance apoptosis of U937 cells in a dose-dependent manner through the activation of p-p38MAPK and downregulation of p-ERK1/2. Further study verified it could alter the expression of Fas/FasL and activation of caspase-8, thus inducing THP-1 cell apoptosis [68].

3.1.3. Anti-Liver Cancer Activity

Li *et al.* [69] reported the inhibition of aqueous extract of *H. diffusa* on blood metastasis in H22 mice. The body and immune organs weights increased after administration with *H. diffusa* extract at three doses of 0.25, 0.5 and 1.0 mg/kg. *In vitro*, the aqueous extract of *H. diffusa* treatment (1.25–10 mg/mL) remarkably inhibited HepG2 cell proliferation in a dose-dependent manner, probably via the arrest of HepG2 cells at the G₀/G₁ phase and the induction of S phase delay [70]. Treatment with total flavones extract from *H. diffusa* could reverse the invasion of MHCC97-H cells in epithelial-mesenchymal transition induced by TGF-β1 at the dose of 200 µg/mL, and the effect might be carried out by decreasing the level of E-cadherin protein and increasing the expression of vimentin protein [71]. Li *et al.* found that both 1,3-Dihydroxy-2-Methylanthraquinone (79 and 157 µmol/L) and ethyl acetate extract (100 and 200 µg/mL) induced apoptosis on HepG2 cells, resulting in upregulation of Bax, p53, Fas, FasL, p21 and cytoplasmic cytochrome C levels and

caspase-3, -8, -9 proteases activities, while downregulation of Bcl-2, mitochondrial cytochrome C, cyclin E and CDK 2 in a dose-dependent manner [72]. Nine compounds from *H. diffusa*, namely, ethyl 13 (S)-hydroxy-chlorophyllide a, 2-methyl-3-methoxy anthraquinone, 2-hydroxymethyl anthraquinone, 2-hydroxy-3-methyl anthraquinone, 2-hydroxymethyl-1-hydroxy anthraquinone, 2-hydroxy-1-methoxy anthraquinone, 2-hydroxy-3-methyl-1-methoxy anthraquinone, oleanolic acid and ursolic acid, have been researched for their anti-liver cancer effect within the concentration range from 1 to 200 μM . As a result, ursolic acid exhibited a strong inhibition of HepG2 cell survival (IC_{50} 36.63 μM) [5]. Another study revealed that the inhibitory activity of 2-hydroxy-3-methyl anthraquinone (IC_{50} 51 μM) and 2-hydroxy-1-methoxy anthraquinone (IC_{50} 62 μM) might be achieved by activity against protein tyrosine kinases v-src and pp60src [38].

3.1.4. Anti-Lung Cancer Activity

Aqueous extract of *H. diffusa* treatment (0–200 $\mu\text{g}/\text{mL}$) showed a suppression effect on A549 and H1355 cells in a concentration-dependent manner. But this effect was not found in LLC cells [66]. Further, Shi *et al.* [38] confirmed that two compounds of 2-hydroxy-3-methyl anthraquinone (IC_{50} 66 μM) and 2-hydroxy-1-methoxy anthraquinone (IC_{50} 79 μM) from *H. diffusa* could induce apoptosis on SPC-1-A cells with a close relationship to the mitochondrial apoptotic pathway.

3.1.5. Anti-Breast Cancer Activity

Anthraquinones, iridoid glucosides, stigmasterols and alkaloid/flavonoid extracts were evaluated for anti-breast cancer using human breast cancer cell line MCF7. Dong *et al.* [73] found that the crude alkaloid/flavonoid extract, but not its three major components, possessed antitumor activity against the human breast cancer cell line MCF7. However, Liu *et al.* [74] observed that methyl anthraquinone from *H. diffusa* exhibited an inhibition effect on MCF7 cells with half maximal effective concentration (EC_{50}) of 18.62 ± 2.71 and 42.19 ± 3.84 μM for 24 and 48 h, respectively, and induced MCF-7 cells apoptosis via the Ca^{2+} /calpain/caspase-4 pathway. Moreover, compounds of 2-hydroxy-3-methyl anthraquinone (IC_{50} 57 μM) and 2-hydroxy-1-methoxy anthraquinone (IC_{50} 65 μM) inhibited protein tyrosine kinases v-src and pp60src and the growth of Bcap37 cells [38].

3.1.6. Anti-Cervical Tumor Activity

Zhang *et al.* [3] discovered that the aqueous extract of *H. diffusa* treated (0.5 g/kg bw) by intragastric administration on human cervical carcinoma xenograft in nude mice showed an inhibitory effect on cervical cancer cells and induced apoptosis of Hela cells. Meanwhile, anthraquinones, especially 2-hydroxymethyl-1-hydroxy anthraquinone, showed a strong inhibitory effect on Hela cells with IC_{50} 45 μM *in vitro* [5].

3.1.7. Anti-Prostate Cancer Activity

The potential anti-prostate cancer effect of *H. diffusa*, mainly the active compounds, has previously been provided on a variety of cell lines. 2-Methyl-3-methoxy anthraquinone (IC_{50} 64.72–105.90 μM), 2-hydroxy-3-methyl anthraquinone (IC_{50} 28.82–159.20 μM) and ursolic acid (IC_{50} 22.33–36.08 μM) exhibited inhibitory effects on DU145, PC-3 and LNCaP cells [5]. 6-*O*-(*E*)-*p*-coumaroyl scandoside methyl ester and 10(*S*)-hydroxy pheophytin a showed an anti-proliferation effect on PC-3 cells in a dose-dependent manner from 0 to 60 μM , while 10(*S*)-hydroxy pheophytin a also showed a strong anti-proliferation effect on LNCaP cells with a significant effect, with an IC_{50} value of 20 μM [52]. Hu *et al.* [51] isolated three cyclotides (DC 1-3) and studied their anti-prostate cancer effect. Thus, three cyclotides, especially DC 3 (1 mg/kg) showed inhibition against PC3, DU145 and LNCaP cells. In addition, DC3 significantly inhibited development of the tumor in weight and size in the model of a prostate xenograft, and showed significant anti-cancer effect ($p < 0.01$) at a dose of 1 mg/kg, with 40.23% inhibition of the rate of tumor growth (weight).

3.1.8. Anti-Multiple Myeloma Activity

Up to now, the anti-multiple myeloma effect of *H. diffusa* has been proved in RPMI 8226 cells. The polysaccharides extracts (1, 2 and 3 mg/mL) [75], the compound of 2-hydroxymethyl-1-hydroxy anthraquinone (1–200 μ M) [5], as well as *H. diffusa* injection (20, 40 and 60 μ L/mL) [76], exhibited an inhibitory effect on RPMI 8226 cells growth in a dose-dependent manner.

3.1.9. Other Anti-Cancer Effects

Other anti-cancer effects have also been reported during these years. The ethanolic extract of *H. diffusa* (0–200 μ g/mL) suppressed the proliferation of B16F10 cells in a dose-dependent manner [66]. The lipophilic extract (50 and 100 mg/kg) and crude polysaccharide (31.25 and 62.5 mg/kg) from *H. diffusa* showed anti-tumor activities on S-180 cells and a protective effect on chemotherapeutic damage [77]. *H. diffusa* injection could induce the apoptosis of MG-63 cells by increasing the Bax gene expression in a concentration-dependent manner from 50 to 400 μ g/mL [78,79]. When it is used with cisplatin, the combined use exhibited a stronger inhibitory effect than the single agents. This might be due to its property of elevating the levels of Bax, Bad, caspase-3 and caspase-8 expression and decreasing the levels of Bcl-x1 and Bcl-2 [80]. Meanwhile, Zhang *et al.* [4] found that the aqueous extract of *H. diffusa* (2–8 mg/mL) inhibited the growth of U87 cells in a dose-dependent manner by inducing mitochondrial apoptosis via the AKT/ERK pathways. Moreover, the compound, 4-vinyl phenol, was demonstrated to have anti-angiogenic activity in human endothelial cells of HUVEC (IC₅₀ 15.31 μ g/mL) and HMEC₁ (IC₅₀ 21.43 μ g/mL), breast tumor-bearing BALB/c mice (0.2–2 mg/kg), C57BL/6 mice (20–100 μ g/mL matrigel) and zebrafish embryo models (6.25–12.5 μ g/mL matrigel), and this effect had a close relationship with the PI3K/AKT pathway [81].

3.2. Immunomodulatory Effect

Lin *et al.* [64] found that aqueous extract of *H. diffusa* (16 and 32 mg/kg) affected immune responses by promoting T- and B-cell proliferation in leukemic mice in WEHI-3-generated leukemia mice. Meanwhile, Kuo *et al.* [82] discovered that the ethanolic extract (16, 32 and 64 mg/kg) could also promote immune responses in normal BALB/c mice by promoting CD11b, CD19 and Mac-3 levels, increasing phagocytosis activity of macrophages obtained from the peritoneal cavity and increasing NK cell activity and B- and T-cell proliferation. The polysaccharides extracts (2.25, 4.5 and 9.0 mg/mL) could improve the clearance index, phagocytic index and the index of the thymus and spleen of immunosuppression mice [50]. When immunosuppressed mice were orally administrated total flavonoids of *H. diffusa* (15, 30 and 60 mg/kg), the levels of interleukin-2 (IL-2) and interferon- γ (INF- γ) were enhanced and the proliferation of T and B lymphocytes was increased, indicating the immunomodulatory effect of total flavonoids [83].

3.3. Antioxidant Effect

The aqueous, methanolic and 80% acetonetic extracts were evaluated for antioxidant activity and the extraction from 80% alcohol (0.1–4.5 mg/mL) showed the strongest antioxidant activity, by 2,2-diphenyl-1-picrylhydrazyl (DPPH) assay [84]. Yu *et al.* [85] compared the antioxidant effects of aqueous, alcoholic, acetonetic, chloroform, ether and petroleum benzene extracts from *H. diffusa*, and found that the acetonetic extracts (0.03%–0.18%), especially the 0.12% acetone extract, had the strongest antioxidant effect, by determination of peroxide value. In addition, the aqueous extract (0.3–10 mg/mL) treatment could protect human hepatocyte cells from H₂O₂-induced cytotoxicity in a dose-dependent manner as the probable result of the improvement activity of the aqueous extract of *H. diffusa* on the antioxidant defense system by reversing H₂O₂-induced activation of the MEK/ERK pathway and H₂O₂-induced inhibition of the P13-K/AKT/GSK3 β pathway in LO₂ cells [86]. The antioxidant effect of *H. diffusa* may be due to its compounds, like flavonoids and iridoids. Three flavonol glycosides (quercetin 3-O-sambubioside, kaempferol-3-O-[2-O-(E-6-O-feruloyl)- β -D-glucopyranosyl]- β -D-galactopyranoside and quercetin 3-O-sophoroside) and six known iridoid

glycosides (asperuloside, asperulosidic acid methyl ester, (*E*)-6-*O*-*p*-methoxy cinnamoyl scandoside methyl ester, (*E*)-6-*O*-feruloyl scandoside methyl ester and (*E*)-6-*O*-coumaroyl scandoside methyl ester) were determined for their antioxidant effects on xanthine oxidase inhibition, xanthine-xanthine oxidase cytochrome c and TBA-MDA systems. In consequence, asperuloside (IC₅₀ 118.5 ± 0.70 μM) and kaempferol-3-*O*-(2-*O*-β-D-glucopyranosyl)-β-D-galactopyranoside (IC₅₀ 98.7 ± 0.16 μM) showed a minor anti-lipid peroxidation effect and quercetin di-glycosides exerted a remarkable antioxidant effect as superoxide anion scavengers [33].

3.4. Anti-Inflammatory Effect

The aqueous extract (5.0 g/kg bw) treatment exhibited an anti-inflammatory effect in lipopolysaccharide-induced renal inflammation of mice by significantly suppressing the production of tumor necrosis factor-α (TNF-α), IL-1, IL-6 and monocyte chemotactic protein 1 (MCP-1) in renal tissues, as well as significantly promoting the production of IL-10 in serum and renal tissues. Moreover, two main chemotypes, including eight flavonoids and four iridoid glycosides were found in renal tissues after *H. diffusa* treatment, indicating that the anti-inflammatory effect may be due to these constituents [87]. *In vitro*, Chen *et al.* found that the flavonoids extract treatment (50–100 μg/mL) on LPS-stimulated RAW 264.7 cells reduced expression of iNOS, TNF-α, IL-6 and IL-1β, as well as suppressing phosphorylation of IκB p38, JNK and ERK1/2 in a concentration-dependent manner, indicating that the anti-inflammatory activity of total flavonoids had a close relationship with the NF-κB- and MAPK-signaling pathways [88].

3.5. Others

Five flavonol glycosides (kaempferol-3-*O*-[2-*O*-(6-*O*-*E*-feruloyl)-β-D-glucopyranosyl]-β-D-galactopyranoside, quercetin-3-*O*-[2-*O*-(6-*O*-*E*-feruloyl)-β-D-glucopyranosyl]-β-D-galactopyranoside, quercetin-3-*O*-[2-*O*-(6-*O*-*E*-feruloyl)-β-D-glucopyranosyl]-β-D-glucopyranoside, kaempferol-3-*O*-(2-*O*-β-D-glucopyranosyl)-β-D-galactopyranoside and quercetin-3-*O*-(2-*O*-β-D-glucopyranosyl)-β-D-galactopyranoside) and four *O*-acylated iridoid glycosides (6-*O*-*Z*-*p*-methoxy cinnamoyl scandoside methyl ester, 6-*O*-*E*-*p*-methoxy cinnamoyl scandoside methyl ester, 6-*O*-*Z*-*p*-coumaroyl scandoside methyl ester and 6-*O*-*E*-*p*-coumaroyl scandoside methyl ester) isolated from *H. diffusa* exhibited a significant neuroprotective effect on L-glutamate-damaged rat cortical cells in the concentration from 0.1 to 10 μM; further, the structure–activity study proved di-OH in the B ring and an acyl substituent in flavonoids, a *p*-methoxy group in the aromatic ring and a trans double bond in the acyl moiety of acylated iridoid glycosides might be crucial for the biological response [34]. Wu *et al.* [89] found the inhibitory effect of oleanolic acid (2 and 8 μg/mL) isolated from *H. diffusa* against ras-transformed fibroblasts on R6 cells, and this inhibition might cause normal cells to secrete an inhibitory factor against the transformed cells, but did not require direct cell–cell contact.

4. Quality Control

Quality control of herbal medicines is necessary to ensure their stability, efficiency and safety. Modern analytical techniques provide simpler, more accurate and reliable methods for the quality control for *H. diffusa*. Besides the macroscopic and microscopic characters of *H. diffusa* [9], DNA sequence has become a powerful tool for the distinguishing *H. diffusa* from counterfeits, such as *H. corymbosa* and *H. tenelliflora* [8,12]. Chemical fingerprint is a comprehensive method accepted by the Food and Drug Administration, European Medicines Agency, and China Food and Drug Administration [90]. It can provide information about the types of compounds, as well as their relative ratios. A HPLC-MS fingerprint method was applied to 10 batches of *H. diffusa* materials from nine regions in China. The results showed that this method could differentiate samples from different geographical origins or processing methods [91]. Liang *et al.* [92] analyzed the chemical fingerprints of 17 batches of *H. diffusa* and found that the contents of asperuloside and (*E*)-6-*O*-*p*-coumaroyl scandoside methyl ester were quite different in samples collected from different habitats.

Table 3. Quantitative analysis for the quality control of *H. diffusa*.

Analytes	Method	Results	Reference
Deacetyl asperulosidic acid methyl ester	HPLC	The contents of deacetyl asperulosidic acid methyl ester of 22 batches were from 0.31 to 3.34 mg/g.	[93]
Oleanolic acid	TLC	The contents of oleanolic acid of 3 batches were from 1.63% to 1.72%	[94]
Isoscutellarein	HPLC	The contents of isoscutellarein have a close relationship with the collecting times and were also different in leaves (1.11–2.72 mg/g) and stem (0.35–0.94 mg/g).	[95]
<i>p</i> -Coumaric acid	HPLC	The contents of <i>p</i> -coumaric acid in the injection of <i>H. diffusa</i> from four manufacturers ranged from 0.34 to 0.49 mg/mL.	[96]
<i>p</i> -Coumaric acid	HPLC	The contents of <i>p</i> -coumaric acid of 13 batches were from 0.46 to 1.88 mg/mL	[97]
3,4-Dihydroxy methyl benzoate	HPLC	The contents of 3,4-dihydroxy methyl benzoate of 8 batches were from 40.8 to 87.0 µg/g.	[98]
Polysascharides	UV	Polysascharides have been determined by the phenol-sulfuric acid method by spectrophotometry at 490 nm, and the content was 15.10%.	[99]
Ursolic acid Oleanolic acid	HPLC	Six batches have been determined with the contents of 1.75–3.37 mg/g for ursolic acid and 0.50–0.80 mg/g for oleanolic acid, indicating that the ursolic acid and oleanolic acid content in the samples from different sources were significantly different.	[100]
Ursolic acid Oleanolic acid	HPLC	The contents of ursolic acid and oleanolic acid have a close relationship with the collecting time. The range of contents was 1.17–3.75 and 0.19–0.96 mg/g for ursolic acid and oleanolic acid, respectively.	[101]
Ursolic acid Oleanolic acid	HPLC	The contents of ursolic acid and oleanolic acid were 0.51%–0.58% and 0.11%–0.14%, respectively. And the contents of the whole herb were slightly lower than those of the overground part for both of the two compounds.	[102]
Ursolic acid Oleanolic acid	HPLC-MS/MS	The contents of ursolic acid and oleanolic acid for 10 batches were 0.15%–0.65% and 0.06%–0.17%, respectively.	[103]
2-Hydroxy-3-methoxy-7-methyl anthraquinone 2-Hydroxy-1-methoxy anthraquinone	HPLC	The contents were 0.16–0.51 and 0.22–0.49 mg/g for 2-hydroxy-3-methoxy-7-methyl anthraquinone and 2-hydroxy-1-methoxyanthraquinone, respectively.	[104]
Asperuloside E-6- <i>O</i> - <i>p</i> -Coumaroyl scandoside methyl ester E-6- <i>O</i> - <i>p</i> -Coumaroyl scandoside methyl ester-10-methyl ether	HPLC	The contents of asperuloside, E-6- <i>O</i> - <i>p</i> -coumaroyl scandoside methyl ester and E-6- <i>O</i> - <i>p</i> -coumaroyl scandoside methyl ester-10-methyl ether have been determined in twenty-three batches. The result was that the contents of the compounds were significantly varied among the different samples. The concentration ranges were 0–7.885, 1.104–7.159 and 0–1.795 mg/g for asperuloside, E-6- <i>O</i> - <i>p</i> -coumaroyl scandoside methyl ester and E-6- <i>O</i> - <i>p</i> -coumaroyl scandoside methyl ester-10-methyl ether, respectively.	[105]

Table 3. Cont.

Analytes	Method	Results	Reference
3,4-Dihydroxy methyl benzoate <i>p</i> -Coumaric acid Ferulic acid (<i>E</i>)-6- <i>O</i> - <i>p</i> -Coumaroyl scandoside methyl ester	HPLC	Four compounds have been quantified in the injection of <i>H. diffusa</i> with contents of 2.25–2.63, 7.02–7.15, 0.96–1.17 and 7.16–7.33 g/L for 3,4-dihydroxy methyl benzoate, <i>p</i> -coumaric acid, ferulic acid and (<i>E</i>)-6- <i>O</i> - <i>p</i> -coumaroyl scandoside methyl ester, respectively.	[106]
Geniposidic acid Ursolic acid Quercetin <i>p</i> -Coumaric acid	CE	Four compounds have been quantified in the injection of <i>H. diffusa</i> with contents of 1.004, 1.182, 0.110 and 0.067 mg/g for ursolic acid, geniposidic acid, quercetin and <i>p</i> -coumaric acid, respectively.	[107]
Asperuloside acid Asperuloside (<i>E</i>)-6- <i>O</i> -Feruloyl scandoside methyl ester (<i>E</i>)-6- <i>O</i> - <i>p</i> -Coumaroyl scandoside methyl ester Scandoside methyl ester	HPLC	The contents were 1.57–5.93, 1.45–3.86, 1.82–3.23, 1.54–3.82 and 1.49–4.11 mg/g for asperuloside acid, asperuloside, (<i>E</i>)-6- <i>O</i> -feruloyl scandoside methyl ester, (<i>E</i>)-6- <i>O</i> - <i>p</i> -coumaroyl scandoside methyl ester and scandoside methyl ester, respectively, and they were very different in different batches.	[108]
Quercetin-3- <i>O</i> -sambubioside Quercetin-3- <i>O</i> - β -D-glucopyranside Kaempferol-3- <i>O</i> - β -D-glucopyranside Rutin Quercetin Kaempferol	HPLC	Six compounds from eight batches of <i>H. diffusa</i> have been quantified with contents of 1.36–6.32, 0.98–10.23, 0.79–7.98, 4.92–15.78, 0.52–1.72 and 0.75–2.15 mg/g for quercetin-3- <i>O</i> -sambubioside, quercetin-3- <i>O</i> - β -D-glucopyranside, kaempferol-3- <i>O</i> - β -D-glucopyranside, rutin, quercetin and kaempferol, respectively, indicating that the contents for these compounds were quite different from different regions.	[109]
Desacetyl asperulosidic acid Asperuloside Aesculetin Coumaric acid Ferulic acid Quercetin Kaempferol	HPLC	Seven compounds from six batches of <i>H. diffusa</i> have been quantified with contents of 42.48 ± 1.43 , 63.76 ± 1.01 , 1765 ± 0.69 , 881.9 ± 0.74 , 86.99 ± 1.65 , 1395 ± 0.731 and 902.2 ± 0.82 μ g/g for desacetyl asperulosidic acid, asperuloside, aesculetin, coumaric acid, ferulic acid, quercetin, kaempferol, respectively.	[110]

The quantitative analysis for the quality control of *H. diffusa* has mostly focused on the diversity of components by a series of analytical methods, such as UV, HPLC, TLC and LC/MS. Up to now, triterpenes (ursolic acid and oleanolic acid), iridoids ((*E*)-6-*O*-*p*-coumaroyl scandoside methyl ester), geniposidic acid, deacetyl asperulosidic acid methyl ester, asperuloside acid, asperuloside and (*E*)-6-*O*-feruloyl scandoside methyl ester), phenolic acid (*p*-coumaric acid and ferulic acid), flavonoids (quercetin, rutin, quercetin-3-*O*- β -D-glucopyranside, quercetin-3-*O*-sambubioside, kaempferol, kaempferol-3-*O*- β -D-glucopyranside), anthraquinones (2-hydroxy-3-methoxy-7-methyl anthraquinone and 2-hydroxy-1-methoxy anthraquinone), polysaccharides and one miscellaneous compound have been quantified as mark compounds for the quality control of *H. diffusa*. However, there were wide variations in the contents of these compounds, caused by samples from different sources and different collecting times (Table 3). Therefore, it is very urgent that a comprehensive method for ensuring the quality of *H. diffusa* be established.

5. Pharmacokinetics

The investigations about pharmacokinetics of *H. diffusa* are very scarce. After oral administration of *H. diffusa* in lipopolysaccharide-induced renal inflammation in mice, most compounds, including flavonoids, iridoid glycosides and anthraquinone, were found in plasma, and 12 compounds (eight flavonoids and four iridoid glycosides) were found in kidney, determined by UPLC-Q-TOF-MS/MS. The results indicated that flavonoids, iridoids and anthraquinones might be responsible for the protective effect of *H. diffusa* on renal inflammation [87]. Liu *et al.* [111] found that *p*-coumaric acid was a major metabolite of (*E*)-6-*O*-*p*-coumaroyl scandoside methyl ester in rat plasma after oral administration of a dose of 20 mg/kg. Compared with direct administration of *p*-coumaric acid, the absorption and elimination of *p*-coumaric acid were slower with administration of (*E*)-6-*O*-*p*-coumaroyl scandoside methyl ester. This was also confirmed by Yan *et al.* at 2011 [112]. Moreover, Ganbold *et al.* [62] investigated the bioavailability of *H. diffusa* by production of post-absorption samples using the Caco-2 cell model and confirmed that the decoction has good permeability ($P_{app} = 3.575 \times 10^{-6}$ cm/s) *in vitro* with no cytotoxic effect.

6. Conclusions

Although *H. diffusa* has been used in China for thousands of years as a heat-clearing and detoxifying medicine, it has become popular for its anti-cancer effect, especially in the Taiwan district. Modern research on *H. diffusa* has provided much evidence for its anti-cancer effect using *in vitro* and *in vivo* experiments and has tried to clarify the mechanism of its action. Meanwhile, its other activities, such as anti-oxidant, anti-inflammatory, anti-fibroblasts, immunomodulatory and neuroprotective effects, have been reported. The achievement of these therapeutic effects is due to the chemical composition of *H. diffusa*. One hundred and seventy-one compounds have been reported, including iridoids, flavonoids, anthraquinones, phenolic acids and their derivatives, sterols, triterpenes, polysaccharides, cyclotides, coumarins, alkaloids and volatile oils. Among these constituents, iridoids, flavonoids and anthraquinones are three main ingredients and may play an essential role in its activities. However, there is no official quality standard for the quality control of *H. diffusa*. The contents of bioactive compounds are significantly different in the samples from different sources and different collecting times. So, a feasible and reliable approach is urgently needed in considering the botanical origin and bioactive effects. Moreover, a relatively small number of pharmacokinetics studies have been summarized, and, therefore, it is difficult to evaluate the function of *H. diffusa* in the human body. Altogether, this review gives comprehensive information about *H. diffusa* and provides evidence for its clinical application and further development.

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References

1. Tao, C.; Taylor, C.M. Rubiaceae. In *Flora of China*; Wu, Z.Y., Raven, P.H., Hong, D.Y., Eds.; Science Press: Beijing, China; Missouri Botanical Garden Press: St. Louis, MO, USA, 2011; Volume 19, pp. 147–174.
2. Lin, J.M.; Li, Q.Y.; Chen, H.W.; Lin, H.; Lai, Z.J.; Peng, J. *Hedyotis diffusa* Willd extract suppresses proliferation and induces apoptosis via IL-6-inducible STAT3 pathway inactivation in human colorectal cancer cells. *Oncol. Lett.* **2015**, *9*, 1962–1970. [[PubMed](#)]
3. Zhang, P.Y.; Zhang, B.; Gu, J.; Hao, L.; Hu, F.F.; Han, C.H. The study of the effect of *Hedyotis diffusa* on the proliferation and the apoptosis of the cervical tumor in nude mouse model. *Cell Biochem. Biophys.* **2015**, *72*, 783–789. [[CrossRef](#)] [[PubMed](#)]
4. Zhang, Y.; Xie, R.F.; Xiao, Q.G.; Li, R.; Shen, X.L.; Zhu, X.G. *Hedyotis diffusa* Willd extract inhibits the growth of human glioblastoma cells by inducing mitochondrial apoptosis via AKT/ERK pathways. *J. Ethnopharmacol.* **2014**, *158*, 404–411. [[CrossRef](#)] [[PubMed](#)]
5. Meng, Q.X.; Roubin, H.R.; Hanranhan, R.J. Ethnopharmacological and bioactivity guided investigation of five TCM anticancer herbs. *J. Ethnopharmacol.* **2013**, *148*, 229–238. [[CrossRef](#)] [[PubMed](#)]
6. Chao, T.H.; Fu, P.K.; Chang, C.H.; Chang, S.N.; Mao, F.C.; Lin, C.H. Prescription patterns of Chinese herbal products for post-surgery colon cancer patients in Taiwan. *J. Ethnopharmacol.* **2014**, *156*, 702–708. [[CrossRef](#)] [[PubMed](#)]
7. Yeh, Y.C.; Chen, H.Y.; Yang, S.H.; Lin, Y.H.; Chiu, J.H.; Lin, Y.H.; Chen, C.L. *Hedyotis diffusa* combined with scutellaria barbata Are the core treatment of Chinese herbal medicine used for breast cancer patients: A population-based Study. *Evid. Based Complement Alternat. Med.* **2014**, *2014*. [[CrossRef](#)] [[PubMed](#)]
8. Li, M.; Wong, Y.L.; Jiang, L.L.; Wong, K.L.; Wong, Y.T.; Lau, C.B.; Shaw, P.C. Application of novel loop-mediated isothermal amplification (LAMP) for rapid authentication of the herbal tea ingredient *Hedyotis diffusa* Willd. *Food Chem.* **2013**, *141*, 2522–2525. [[CrossRef](#)] [[PubMed](#)]
9. Lee, H.Z.; Bau, D.T.; Kuo, C.L.; Tsai, R.Y.; Chen, C.Y.; Chang, Y.H. Clarification of the phenotypic characteristics and anti-tumor activity of *Hedyotis diffusa*. *Am. J. Chin. Med.* **2011**, *39*, 201–213. [[CrossRef](#)] [[PubMed](#)]
10. Lau, C.B.; Cheng, L.; Cheng, B.W.; Yue, G.G.; Wong, E.C.; Lau, C.P.; Leung, P.C.; Fung, K.P. Development of a simple chromatographic method for distinguishing between two easily confused species, *Hedyotis diffusa* and *Hedyotis corymbosa*. *Nat. Prod. Res.* **2012**, *26*, 1446–1450. [[CrossRef](#)] [[PubMed](#)]
11. Wang, L.; Zhou, C.; Mai, H.Z. Analysis of volatile compounds in *Hedyotis diffusa* and *Hedyotis corymbosa*. *J. Chin. Mater. Med.* **2003**, *26*, 563–564.
12. Liu, Z.M.; Hao, M.G.; Wang, J.L. Application of allele-specific primer in the identification of *Hedyotis diffusa*. *J. Chin. Mater. Med.* **2004**, *27*, 484–487.
13. Yang, Y.B.; Yang, X.Q.; Ding, Z.T. Chemical constituents from *Hedyotis diffusa*. *Chin. J. Yunnan Univ. (Nat. Sci.)* **2007**, *29*, 187–189.
14. Liang, Z.T.; He, M.F.; Fong, W.F.; Jiang, Z.H.; Zhao, Z.Z. A comparable, chemical and pharmacological analysis of the traditional Chinese medicinal herbs *Oldenlandia diffusa* and *O. corymbosa* and a new valuation of their biological potential. *Phytomedicine* **2008**, *15*, 259–267. [[CrossRef](#)] [[PubMed](#)]
15. Liu, E.H.; Zhou, T.; Li, G.B.; Li, J.; Huang, X.N.; Pan, F.; Gao, N. Characterization and identification of iridoid glucosides, flavonoids and anthraquinones in *Hedyotis diffusa* by high-performance liquid chromatography/electrospray ionization tandem mass spectrometry. *J. Sep. Sci.* **2012**, *35*, 263–272. [[CrossRef](#)] [[PubMed](#)]
16. Nishihama, Y.; Masuda, K.; Yamaki, M.; Takagi, S.; Sakina, K. Three new iridoid glucosides from *Hedyotis diffusa*. *DlantaMedica* **1981**, *43*, 28–33. [[CrossRef](#)] [[PubMed](#)]
17. Zhang, Y.Y.; Luo, J.B. Studies on the chemical constituents in Herb of *Hedyotis diffusa*. *J. Chin. Mater. Med.* **2008**, *31*, 522–524.

18. Ji, B.Y.; Fan, C.Q.; Fei, L.X.; Ma, Y. Advance on the chemical and pharmacological effects studies of *Hedyotis diffusa*. *Chin. J. Exp. Tradit. Med. Form.* **2014**, *20*, 235–240.
19. Si, J.Y.; Chen, D.H.; Pan, R.L.; Zhao, X.H. Chemical constituents of *Hedyotis Diffusa*. *Nat. Prod. Res. Dev.* **2006**, *18*, 942–944.
20. Zhang, Y.Y.; Chen, Y.; Fan, C.L.; Ye, W.C.; Luo, J.B. Two new iridoids from *Hedyotis diffusa*. *Fitoterapia* **2010**, *81*, 515–517. [[CrossRef](#)] [[PubMed](#)]
21. Liu, J.Z.; Wang, L. Studies on chemical constituents of *Hedyotis diffusa* Willd. *J. Hebei Med. Univ.* **2007**, *28*, 188–190.
22. Li, C.M.; Zhao, Y.Y.; Guo, Z.M.; Zhang, X.L.; Xue, X.Y.; Liang, X.M. Effective 2D-RPLC/RPLC enrichment and separation of micro-components from *Hedyotis diffusa* Willd and characterization by using ultra-performance liquid chromatography/quadrupole time-of-flight mass spectrometry. *J. Pharm. Biomed. Anal.* **2014**, *99*, 35–44. [[CrossRef](#)] [[PubMed](#)]
23. Chen, W.C.; Gu, D.W.; Zhang, H.; Zhu, Z.Y.; Zhang, G.Q.; Chai, Y.F. HPLC-TOEMS in rapid separation and identification of chemical components in *Oldenlandia diffusa* and its injection preparations. *Acad. J. Second Mil. Med. Univ.* **2010**, *31*, 292–296. [[CrossRef](#)]
24. Li, C.; Xue, X.; Zhou, D.; Zhang, F.; Xu, Q.; Ren, L.; Liang, X. Analysis of iridoid glucosides in *Hedyotis diffusa* by high-performance liquid chromatography/electrospray ionization tandem mass spectrometry. *J. Pharm. Biomed. Anal.* **2008**, *48*, 205–211. [[CrossRef](#)] [[PubMed](#)]
25. Li, D.X.; Schmitz, O.J. Comprehensive two-dimensional liquid chromatography tandem diode array detector (DAD) and accurate mass QTOF-MS for the analysis of flavonoids and iridoid glycosides in *Hedyotis diffusa*. *Anal. Bioanal. Chem.* **2015**, *407*, 231–240. [[CrossRef](#)] [[PubMed](#)]
26. Xu, G.H.; Kim, Y.H.; Chi, S.W.; Choo, S.J.; Ryoo, I.J.; Ahn, J.S.; Yoo, I.D. Evaluation of human neutrophil elastase inhibitory effect of iridoid glycosides from *Hedyotis diffusa*. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 513–515. [[CrossRef](#)] [[PubMed](#)]
27. Zhang, Q.M.; Sun, Z.Y. Study on chemical constituents of *Oldenlandia diffusa*. *Chin. J. Chin. Mater. Med.* **2014**, *37*, 2216–2218.
28. Ren, R.A. *Identification of Chinese Drug*; Shanghai Scientific & Technological Publishers: Shanghai, China, 1986; p. 491.
29. Wu, K.S.; Zhang, K.; Tan, G.S.; Zeng, G.Y.; Zhou, Y.J. Study on constituents of *Oldenlandia diffusa*. *Chin. Pharm. J.* **2005**, *40*, 817–818.
30. Liang, S.Y.; Chen, F.L.; Tang, Q.F.; Luo, J.B.; Zeng, Y.C. Study of Chemical Constituents from Herba *Hedyotis diffusa*. *Tradit. Chin. Drug Res. Clin. Pharm.* **2012**, *23*, 655–657.
31. Zhou, Y.J.; Wu, K.S.; Zeng, G.R.; Tang, J.B.; Xu, K.P.; Li, F.S.; Tang, G.S. Study on chemical constituents of *Oldenlandia diffusa*. *Chin. J. Chin. Mater. Med.* **2007**, *32*, 590–593.
32. Zhang, H.J.; Chen, Y.G.; Huang, R. Study on flavonoids constituents of *Oldenlandia diffusa*. *Chin. J. Chin. Mater. Med.* **2005**, *28*, 385–387.
33. Lu, C.M.; Yang, J.J.; Wang, P.Y.; Lin, C.C. A new acylated flavonol glycoside and antioxidant effects of *Hedyotis diffusa*. *Planta Med.* **2000**, *66*, 374–377. [[CrossRef](#)] [[PubMed](#)]
34. Kim, Y.; Park, E.J.; Kim, J.; Kim, Y.; Kim, S.R.; Kim, Y.Y. Neuroprotective constituents from *Hedyotis diffusa*. *J. Nat. Prod.* **2001**, *64*, 75–78. [[CrossRef](#)] [[PubMed](#)]
35. Ren, F.Z.; Liu, G.S.; Zhang, L.; Niu, G.Y. Study on Chemical constituents of *Hedyotis diffusa*. *Chin. Pharm. J.* **2005**, *40*, 502–504.
36. Kang, X.D.; Li, X.; Mao, Y.; Zhao, C.C.; Li, N.; Meng, D.L. Chemical constituents of *Hedyotis diffusa* Willd. *J. Shenyang Pharm. Univ.* **2007**, *24*, 479–481.
37. Liu, Y.Q.; Ying, W.J.; Liu, Y.; Feng, Y.N.; Lv, Q.T. Summarization on the chemical constituents of *Oldenlandia Diffusa* Willd. *Shandong J. Tradit. Chin. Med.* **2014**, *33*, 709–712.
38. Shi, Y.; Wang, C.H.; Gong, X.G. Apoptosis-inducing effects of two anthraquinones from *Hedyotis diffusa* Willd. *Biol. Pharm. Bull.* **2008**, *31*, 1075–1078. [[CrossRef](#)] [[PubMed](#)]
39. Yu, L.; Li, J.M.; Jiang, Z.; Guo, X.J. A new anthraquinone from *Hedyotis diffusa*. *Chin. J. Med. Chem.* **2008**, *18*, 298–300.
40. Tai, D.F.; Lin, Y.M.; Chen, F.C. Component of *Hedyotis diffusa* willd. *Chemistry.* **1979**, *3*, 60–61.
41. Huang, W.H.; Li, Y.B.; Jiang, J.Q. Chemical constituents from *Hedyotis diffusa*. *Chin. J. Chin. Mater. Med.* **2008**, *33*, 524–526.

42. Kang, X.D.; Li, X.; Mao, Y. A new anthraquinone from *Hedyotis diffusa* Willd. *Chin. J. Chin. Mater. Med.* **2006**, *16*, 368–370.
43. Zhou, Y.; Gao, W.Y.; Wang, Y.; Liu, X.J. Studies on constituents of *Oldenlandia diffusa*. *Chin. Tradit. Herb. Drug* **2007**, *38*, 55–57.
44. Lv, H.C.; He, J. A study on chemical constituents of *Oldenlandia diffusa* (Willd) Roxb. *Nat. Prod. Res. Dev.* **1996**, *8*, 34–37.
45. Ruehle, P.H.; Browne, C.E.; Vickery, E.H.; Beller, N.R.; Eisenbraun, E.J.; Loghry, R.A.; Van der Helm, D. Synthesis and antifertility activity of 3,9-dihydroxy-5,6,6a alpha,6b beta,11,12,12a beta,12b alpha-octahydrodibenzo[a,g]biphenylene, a structural relative of diethylstilbestrol. *J. Med. Chem.* **1980**, *23*, 1410–1414. [[CrossRef](#)] [[PubMed](#)]
46. Huang, W.; Li, Y.; Jiang, J. Chemical constituents from *Hedyotis diffusa*. *Chin. J. Chin. Mater. Med.* **2009**, *34*, 712–714.
47. Tan, N.H.; Wang, S.M.; Yang, Y.B.; Tian, F. Anticancer activity and principles of *Hedyotis diffusa*. *Nat. Prod. Res. Dev.* **2002**, *14*, 33–36.
48. Liu, Z.G.; Luo, J.B.; Chen, F.L. The Pilot Study of Volatile Compounds in *Hedyotis diffusa* from different sources. *Tradit. Chin. Drug Res. Clin. Pharm.* **2005**, *16*, 132–134.
49. Yang, S.; Yang, W.W.; Hu, J.F.; Lv, Q.F.; Rong, R.; Jiang, H.Q.; Gong, L.L. GC-MS combined with Kovats Index analysis for volatile compounds in *Hedyotis diffusa*. *Chin. J. Exp. Tradit. Med. Form.* **2012**, *18*, 93–95.
50. Ma, H.; Cheng, Y.L.; Zhang, J.J.; Cao, G.S.; Yang, P.M. Effect of preliminary immune activity and structural identification of a polysaccharide extracted from *Oldenlandia diffusa*. *Chin. J. Exp. Tradit. Med. Form.* **2014**, *20*, 37–40.
51. Hu, E.; Wang, D.G.; Chen, J.Y.; Tao, X.L. Novel cyclotides from *Hedyotis diffusa* induce apoptosis and inhibit proliferation and migration of prostate cancer cells. *Int. J. Clin. Exp. Med.* **2015**, *8*, 4059–4065. [[PubMed](#)]
52. Li, M.; Jiang, R.W.; Hon, P.M.; Cheng, L.; Li, L.L.; Zhou, J.R.; Shaw, P.C.; But, P.P. Authentication of the anti-tumor herb *Baihuasheshicao* with bioactive marker compounds and molecular sequences. *Food Chem.* **2010**, *119*, 1239–1245. [[CrossRef](#)]
53. Wang, X.; Cheng, W.M.; Yao, X.M.; Guo, X.J. Qualitative analysis of the chemical constituents in *Hedyotis diffusa* by HPLC-TOF-MS. *Nat. Prod. Res.* **2012**, *26*, 167–172. [[CrossRef](#)] [[PubMed](#)]
54. Qi, J.Y.; Fan, W.P.; Ju, P.P. Extraction and deputation of polysaccharide from *Hedyotis diffusa* willd. *Acta Universitatis Medicinalis Nanjing* **2001**, *21*, 558.
55. Lin, M.H.; Lin, J.M.; Wei, L.H.; Xu, W.; Hong, Z.F.; Cai, Q.Y.; Peng, J.; Zhu, D.Z. *Hedyotis diffusa* Willd extract inhibits HT-29 cell proliferation via cell cycle arrest. *Exp. Ther. Med.* **2012**, *4*, 307–310. [[PubMed](#)]
56. Lin, J.M.; Chen, Y.Q.; Wei, L.H.; Chen, X.Z.; Xu, W.; Hong, Z.F. *Hedyotis diffusa* Willd extract induces apoptosis via activation of the mitochondrion-dependent pathway in human colon carcinoma cells. *Int. J. Oncol.* **2010**, *37*, 1331–1338. [[PubMed](#)]
57. Lin, J.M.; Wei, L.H.; Xu, W.; Hong, Z.F.; Liu, X.X.; Peng, J. Effect of *Hedyotis diffusa* Willd extract on tumor angiogenesis. *Mol. Med. Rep.* **2011**, *4*, 1283–1288. [[PubMed](#)]
58. Lin, J.M.; Wei, L.H.; Shen, A.L.; Cai, Q.Y.; Xu, W.; Li, H. *Hedyotis diffusa* Willd extract suppresses Sonic hedgehog signaling leading to the inhibition of colorectal cancer angiogenesis. *Int. J. Oncol.* **2013**, *42*, 651–656. [[CrossRef](#)] [[PubMed](#)]
59. Cai, Q.Y.; Lin, J.M.; Wei, L.H.; Zhang, L.; Wang, L.L.; Zhan, Y.Z. *Hedyotis diffusa* Willd inhibits colorectal cancer growth *In Vivo* via inhibition of STAT3 signaling pathway. *Int. J. Mol. Sci.* **2012**, *13*, 6117–6128. [[CrossRef](#)] [[PubMed](#)]
60. Wu, Z.P.; Jin, C.G.; Li, J.; Chen, X.Q.; Yao, Q.; Zhu, Q.S. Inhibition of colon cancer cells by ethanol extract of *Oldenlandia diffusa*. *J. Kunming Med. Univ.* **2013**, *34*, 31–34.
61. Li, Q.Y.; Wang, X.F.; Shen, A.L.; Zhang, Y.C.; Chen, Y.Q.; Thomas, J.S.; Lin, J.M.; Peng, J. *Hedyotis diffusa* Willd overcomes 5-fluorouracil resistance in human colorectal cancer HCT-8/5-FU cells by downregulating the expression of P-glycoprotein and ATP-binding cassette subfamily G member 2. *Exp. Ther. Med.* **2015**, *10*, 1845–1850. [[PubMed](#)]
62. Ganbolda, M.; Barkera, J.; Ma, R.; Jones, L.; Carew, M. Cytotoxicity and bioavailability studies on a decoction of *Oldenlandia diffusa* and its fractions separated by HPLC. *J. Ethnopharmacol.* **2010**, *131*, 396–403. [[CrossRef](#)] [[PubMed](#)]

63. Zhu, D.C.; Pan, R.B.; Wang, Q. Research on the mechanisms of inhibiting effects of the aqueous extract of *Hedyotis diffusa* Willd on CEM cells. *Lishizhen Med. Mater. Med. Res.* **2014**, *25*, 827–829.
64. Lin, C.C.; Kuo, C.L.; Lee, M.H.; Hsu, S.C.; Huang, A.C.; Tang, N.Y.; Lin, J.P.; Yang, J.S.; Lu, C.C.; Chiang, J.H. Extract of *Hedyotis diffusa* Willd influences murine leukemia WEHI-3 cells *in vivo* as well as promoting T- and B-cell proliferation in leukemic mice. *In Vivo* **2011**, *25*, 633–640. [[PubMed](#)]
65. Chen, X.H.; Gao, R.L.; Qian, X.D.; Wang, X.; Tan, P.L.; Yin, L.M.; Zhou, Y.H. Inhibition effect of *Hedyotis diffusa* wild injection on HL-60 cells and its mechanism. *J. Exp. Hematol.* **2008**, *16*, 1035–1038.
66. Kuo, Y.J.; Yang, J.S.; Lu, C.C.; Chiang, S.Y.; Lin, J.G.; Chung, J.G. Ethanol extract of *Hedyotis diffusa* willd upregulates G₀/G₁ phase arrest and induces apoptosis in human leukemia cells by modulating caspase cascade signaling and altering associated genes expression was assayed by cDNA microarray. *Environ. Toxicol.* **2015**, *30*, 1162–1177. [[CrossRef](#)] [[PubMed](#)]
67. Wang, N.; Li, D.Y.; Niu, H.Y.; Zhang, Y.; He, P.; Wang, J.H. 2-Hydroxy-3-methylanthraquinone from *Hedyotis diffusa* Willd induces apoptosis in human leukemic U937 cells through modulation of MAPK pathways. *Arch. Pharm. Res.* **2013**, *36*, 752–758. [[CrossRef](#)] [[PubMed](#)]
68. Wang, J.H.; Shu, L.H.; Yang, L.L.; Zhang, M.; Zhang, M.; He, P. 2-Hydroxy-3-methylanthraquinone from *Hedyotis diffusa* WILLD Induces apoptosis via alteration of Fas/FasL and activation of caspase-8 in human Leukemic THP-1 Cells. *Arch. Med. Res.* **2011**, *42*, 577–583. [[CrossRef](#)] [[PubMed](#)]
69. Li, J.; Sun, J.; Song, J. Experimental research on effect of *Hedyotis diffusa* Willd on blood metastasis in H22 mice. *Lishizhen Med. Mater. Med. Res.* **2012**, *23*, 2434–2435.
70. Chen, X.Z.; Cao, Z.Y.; Chen, T.S.; Zhang, Y.Q.; Liu, Z.Z.; Su, Y.T. Water extract of *Hedyotis Diffusa* Willd suppresses proliferation of human HepG2 cells and potentiates the anticancer efficacy of low-dose 5-fluorouracil by inhibiting the CDK2-E2F1 pathway. *Oncol. Rep.* **2012**, *28*, 742–748. [[PubMed](#)]
71. Zhang, Y.B.; Zhu, J.; Xiao, J.X.; Guo, Y.H.; Liao, Z.J.; Xu, R. Effect and mechanism of total flavones of *Oldenlandia diffusa* willd on epithelial-mesenchymal transition of cell line MHCC97-H induced by TGF- β 1. *J. Xi'an Jiaotong Univ. (Med. Sci.)* **2016**, *37*, 279–282.
72. Li, Y.L.; Zhang, J.; Min, D.; Hongyan, Z.; Lin, N.; Li, Q.S. Anticancer effects of 1,3-dihydroxy-2-methyl anthraquinone and the ethyl acetate fraction of *Hedyotis diffusa* willd against HepG2 carcinoma cells mediated via apoptosis. *PLoS ONE* **2016**. [[CrossRef](#)]
73. Dong, Q.; Ling, B.; Gao, B.; Maley, J.; Sammynaiken, R.; Yang, J. *Hedyotis diffusa* water extract diminished the cytotoxic effects of chemotherapy drugs against human breast cancer MCF7 cells. *Nat. Prod. Commun.* **2014**, *9*, 699–700. [[PubMed](#)]
74. Liu, Z.; Liu, M.; Liu, M.; Li, J.C. Methyl anthraquinone from *Hedyotis diffusa* WILLD induces Ca²⁺-mediated apoptosis in human breast cancer cells. *Toxicol. In Vitro* **2010**, *24*, 142–147. [[CrossRef](#)] [[PubMed](#)]
75. Lin, S.Y.; Shen, C.Y.; Jiang, J.P.; Wu, L.Q.; Dai, T.Y.; Qian, W.B. Meng HT Apoptosis of multiple myeloid cells induced by polysaccharides extracts from *Hedyotis diffusa* and its mechanism. *Chin. J. Hematol.* **2013**, *34*, 337–340.
76. Zhang, X.; Ye, B.D.; Lin, S.Y. Effects of *Hedyotis diffusa* Willd injection on the proliferation of RPMI 8226 cells. *Chin. J. Integr. Tradit. West. Med.* **2012**, *32*, 1658–1662.
77. Zhao, H.R.; Li, R.; Lin, Y.N.; Cheng, N.L. Influence of extraction on process of *Hedyotis diffusa* on anti-tumor activity. *J. Chin. Pharm. Univ.* **2002**, *33*, 510–513.
78. Huang, Y.L.; Tang, Y.J.; Wang, J.L.; Xie, K.G.; Huang, K. Effect of *Hedyotis diffusa* willd injection on osteosarcoma MG-63 cells bax gene expression. *Chongqing Med.* **2014**, *43*, 4708–4710.
79. Xie, K.G.; Tang, Y.J.; Huang, Y.L.; Huang, K.; Lu, L.; Lin, J.J.; Lu, X.Z. Effect of different concentration and action time spreading *Hedyotis* herb injection induced MG-63 cells apoptosis. *Med. Innov. China* **2016**, *13*, 255–263.
80. Pu, F.; Chen, F.; Lin, S.; Chen, S.; Zhang, Z.; Wang, B.; Shao, Z. The synergistic anticancer effect of cisplatin combined with *Oldenlandia diffusa* in osteosarcoma MG-63 cell line *in vitro*. *OncoTargets Ther.* **2016**, *9*, 255–263.
81. Yue, G.G.L.; Lee, J.K.M.; Kwok, H.F.; Cheng, L.; Wong, E.C.W.; Jiang, L.; Yu, H.; Leung, H.W.; Wong, Y.L.; Leung, P.C.; *et al.* Novel PI3K/AKT targeting anti-angiogenic activities of 4-vinylphenol, a new therapeutic potential of a well-known styrene metabolite. *Sci. Rep.* **2015**, *5*. [[CrossRef](#)] [[PubMed](#)]
82. Kuo, Y.J.; Lin, J.P.; Hsiao, Y.T.; Chou, G.L.; Tsai, Y.H.; Chiang, S.Y.; Lin, J.G.; Chuang, J.G. Ethanol extract of *Hedyotis diffusa* Willd affects immune responses *in vivo*. *In Vivo* **2015**, *29*, 453–460. [[PubMed](#)]

83. Wang, Y.L.; Zhang, Y.; Fang, M.; Li, Q.J.; Jiang, Q.; Ming, L. Immunomodulatory effects of total flavones of *Oldenlandia diffusa* Willd. *Chin. Pharmacol. Bull.* **2005**, *21*, 444–447.
84. Yang, X.Z.; Hao, Z.Y.; Zhu, Y.C.; Dong, Y. Effects of different solvents and extraction methods on antioxidant activity of *Hedyotis diffusa* Extract. *Guizhou Agric. Sci.* **2014**, *42*, 43–45.
85. Yu, X.; Du, Z.J.; Chen, Y.J.; Huang, T.Q. Study on antioxidant effect from *Oldenlandia diffusa* Willd. *Food Ferment. Ind.* **2002**, *28*, 10–13.
86. Gao, X.; Li, C.; Tang, Y.L.; Zhang, H.; Chan, S.W. Effect of *Hedyotis diffusa* water extract on protecting human hepatocyte cells (LO₂) from H₂O₂-induced cytotoxicity. *Pharm. Biol.* **2015**. [[CrossRef](#)] [[PubMed](#)]
87. Ye, J.H.; Liu, M.H.; Zhang, X.L.; He, J.Y. Chemical profiles and protective effect of *Hedyotis diffusa* Willd in lipopolysaccharide-induced renal inflammation mice. *Int. J. Mol. Sci.* **2015**, *16*, 27252–27269. [[CrossRef](#)] [[PubMed](#)]
88. Chen, Y.; Lin, Y.; Li, Y.; Li, C. Total flavonoids of *Hedyotis diffusa* Willd inhibit inflammatory responses in LPS-activated macrophages via suppression of the NF- κ B and MAPK signaling pathways. *Exp. Ther. Med.* **2016**, *11*, 1116–1122. [[CrossRef](#)] [[PubMed](#)]
89. Wu, P.K.; Tai, W.C.S.; Liang, Z.T.; Zhao, Z.Z.; Hsiao, W.L.W. Oleanolic acid isolated from *Oldenlandia diffusa* exhibits a unique growth inhibitory effect against ras-transformed fibroblasts. *Life Sci.* **2009**, *85*, 113–121. [[CrossRef](#)] [[PubMed](#)]
90. Liu, L.S.; Liu, M.H.; He, J.Y. *Hypericum japonicum* Thunb. ex murray: Phytochemistry, pharmacology, quality control and pharmacokinetics of an important herbal medicine. *Molecules* **2014**, *19*, 10733–10754. [[CrossRef](#)] [[PubMed](#)]
91. Yang, T.; Yang, Y.H.; Yang, J.Y.; Chen, B.M.; Chen, Y.X.; Yu, S.Y.; Duan, J.P.; Ouyang, H.T.; Cheng, J.P. Finger print of *Hedyotis diffusa* Willd by LC-MS. *Chin. Med. Herb.* **2007**, *4*, 21–23.
92. Liang, Z.T.; Jiang, Z.H.; Ho, H.; Zhao, Z.Z. Comparative analysis of *Oldenlandia diffusa* and its substitutes by high performance liquid chromatographic fingerprint and mass spectrometric analysis. *Planta Med.* **2007**, *73*, 1502–1508. [[CrossRef](#)] [[PubMed](#)]
93. Fan, C.Q.; Li, R.R.; Jin, Y.; Li, H.X.; Feng, X.F. Quality standard of *Hedyotis diffusae*. *Chin. J. Exp. Tradit. Med. Form.* **2014**, *20*, 98–101.
94. Lu, W.B. Quantitative determination of oleanolic acid in *Oldenlandia diffusa* (Willd) Roxb. by TLC-scanning. *Lishizhen Med. Mater. Med. Res.* **2001**, *12*, 961–962.
95. Cao, G.S.; Yang, P.M.; Wang, X.F.; Li, H.; Gao, P. Determination of isoseutellarein in different parts of *Hedyotis diffusa* in different harvest time by HPLC. *Chin. J. Exp. Tradit. Med. Form.* **2014**, *20*, 49–51.
96. Zhang, C.H.; Guo, X.J.; Xue, X.F.; Yang, L.; Ran, G.M.; Li, F.M. Determination of *p*-coumaric acid in *Baihua Sheshicao* injection by HPLC. *Chin. Pharm. J.* **2004**, *39*, 854–855.
97. Zhang, C.H.; Guo, X.J.; Bao, L.D.; Qin, F.; Li, M. Determination of *p*-coumaric acid in *Hedyotis diffusae* Willd. From different source by reversed-phase high-performance liquid chromatography. *Chin. J. Chromatogr.* **2005**, *23*, 180–182.
98. Ma, L.; Li, J.M.; Chen, Y.Q.; Li, Y.; Guo, X.J. Determination of 3,4-dihydroxy Methyl Benzoate in *Hedyotis diffusa* Willd by HPLC. *Lishizhen Med. Mater. Med. Res.* **2009**, *20*, 528–529.
99. Ling, Y.Z. Separation and content determination of polysaccharides in *Hedyotis diffusa* Willd. *Biotechnology* **2005**, *15*, 48–50.
100. Yang, Y.C.; Wei, M.C.; Chiu, H.F.; Huang, T.C. Development and validation of a modified ultrasound-assisted extraction method and a HPLC method for the quantitative determination of two triterpenic acids in *Hedyotis diffusa*. *Nat. Prod. Commun.* **2013**, *8*, 1683–1686. [[PubMed](#)]
101. Zhang, Y.; Tan, X.H.; Cui, X.B.; Jiang, G.B.; Zhu, Y.L. Quantitative determination of ursolic acid and oleanolic acid in *Baihuasheshicao* (Herba *Hedyotis diffusae*) produced from different places by HPLC. *J. Beijing Univ. Tradit. Chin. Med.* **2010**, *33*, 274–276.
102. Zhou, S.Q.; Leng, G.H. Mensuration of content of oleanolic acid and malonic acid of *Hedyotis diffusa* Willd by HPLC. *J. Anhui Agric. Sci.* **2006**, *34*, 1785–1787.
103. Yang, Y.H.; Chen, Y.X. Determination of oleanolic acid and ursolic acid in *Oldenlandia diffusa* (Willd) Roxb. by LC/MS. *Herb. Med.* **2008**, *27*, 589–591.
104. Liu, Y.Q.; Ying, W.J.; Zuo, L.; Man, Q.Q.; Lv, H.T.; Jiang, H.Q.; Gong, L.L. Determination of two anthraquinones in *Hedyotis diffusa* by HPLC. *Chin. J. Exp. Tradit. Med. Form.* **2014**, *20*, 42–44.

105. Liang, Z.T.; Jiang, Z.H.; Leung, K.S.Y.; Zhao, Z.Z. Determination of iridoid glucosides for quality assessment of herba oldenlandiae by high-performance liquid chromatography. *Chem. Phann. Bull.* **2006**, *54*, 1131–1137. [[CrossRef](#)]
106. Zhang, H.F.; Zhang, H.Y.; Li, Y.; Guo, X.J. Simultaneous determination of four components in *Hedyotis diffusa* oral solution by reversed-phase HPLC. *Chin. J. Chin. Mater. Med.* **2008**, *33*, 2329–2331.
107. Cheung, H.Y.; Cheung, S.H.; Law, M.L.; Lai, W.P. Simultaneous determination of key bioactive components in *Hedyotis diffusa* by capillary electrophoresis. *J. Chromatogr. B* **2006**, *834*, 195–198. [[CrossRef](#)] [[PubMed](#)]
108. Yang, P.M.; Cao, G.S.; Li, F.; Gao, P. Contents of five iridoids in *Ooldenlandia diffusa* (Wind) Roxb. based on HPLC-DAD. *Chin. Hosp. Pharm. J.* **2015**, *35*, 9–12.
109. Cao, G.S.; Yang, P.M.; Li, F.; Li, J. Determination of six active flavonoids in *Oldenlandia diffusa* based on HPLC-DAD. *Chin. J. Exp. Tradit. Med. Form.* **2014**, *20*, 52–55.
110. Zhai, X.; Lv, Y. Simultaneous determination of 7 active components in *Hedyotis diffuse* by HPLC. *Chin. Pharm.* **2016**, *19*, 70–72.
111. Liu, K.; Yan, L.Q.; Yao, G.C.; Guo, X.J. Estimation of *p*-coumaric acid as metabolite of *E*-6-*O*-*p*-coumaroyl scandoside methyl ester in rat plasma by HPLC and its application to a pharmacokinetic study. *J. Chromatogr. B* **2006**, *831*, 303–306. [[CrossRef](#)] [[PubMed](#)]
112. An, L.Z.; Wang, Y.F.; Yu, Q.R. HPLC-TOF-MS analysis of metabolites of *Oldenlandia diffusa* effective extracts in rats. *Chin. J. Chin. Mater. Med.* **2011**, *36*, 1301–1304.



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